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The reactive stroma response regulates the immune landscape in prostate cancer

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How to cite this article: Thomas R, Jerome JM, Krieger KL, Ashraf N, Rowley DR. The reactive stroma response regulates the immune landscape in prostate cancer. *J Transl Genet Genom* 2024;8:249-77. <https://dx.doi.org/10.20517/jtgg.2024.15>

Received: 28 Mar 2024 **First Decision:** 12 Jun 2024 **Revised:** 10 Jul 2024 **Accepted:** 17 Jul 2024 **Published:** 24 Jul 2024

Academic Editor: Xiaolin Zi **Copy Editor:** Fangyuan Liu **Production Editor:** Fangyuan Liu

Abstract

Prostate cancer remains the most commonly diagnosed and the second leading cause of cancer-related deaths in men in the United States. The neoplastic transformation of prostate epithelia, concomitant with modulations in the stromal compartment, known as reactive stromal response, is critical for the growth, development, and progression of prostate cancer. Reactive stroma typifies an *emergent response* to disrupted tissue homeostasis commonly observed in wound repair and pathological conditions such as cancer. Despite the significance of reactive stroma in prostate cancer pathobiology, our understanding of the ontogeny, phenotypic and functional heterogeneity, and reactive stromal regulation of the immune microenvironment in prostate cancer remains limited. Traditionally characterized to have an immunologically "cold" tumor microenvironment, prostate cancer presents significant challenges for advancing immunotherapy compared to other solid tumors. This review explores the detrimental role of reactive stroma in prostate cancer, particularly its immunomodulatory function. Understanding the molecular characteristics and dynamic transcriptional program of the reactive stromal populations in tandem with tumor progression could offer insights into enhancing immunotherapy efficacy against prostate cancer.

Keywords: Prostate cancer, emergent/repair biology, reactive stroma, tumor microenvironment, immune landscape, immunotherapy



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INTRODUCTION

The prostate, an exocrine gland located beneath the urinary bladder and surrounding the urethra, is essential for male reproductive function. During embryonic development, epithelial buds from the urogenital sinus interact with mesenchyme, driving prostate differentiation and secretory duct formation^[1-3]. Cancer of the prostate gland, the most diagnosed cancer in men and the second-leading cause of male cancer deaths in the United States, typically progresses slowly^[4-6]. Prostate cancer incidence and mortality rates are closely linked to aging. Additionally, in the United States, African American men have a higher incidence rate of developing aggressive prostate cancer compared to their Caucasian counterparts^[4,7]. While localized disease responds well to surgery or radiation, 20% to 50% of patients experience biochemical recurrence within a decade, leading to advanced or metastatic cancer^[7]. The prostate stroma plays a critical role in promoting the progression of advanced prostate cancer^[8].

In adult differentiated biology, the physical and biochemical interactions between the epithelial cells together with cellular and non-cellular components of the stroma regulate normal prostate function and homeostasis^[9]. The prostate stroma is composed of smooth muscle cells, tissue-resident mesenchymal cells, extracellular matrix (ECM) proteins, nerves, blood vessels, and a spectrum of immune cells [Figure 1]^[9-11]. During development, androgen secreted by the testis is the chief regulator of prostate gland growth, and the hormonal action is mediated through the androgen receptor (AR)^[12]. However, tissue recombination experiments have revealed that the androgenic effects on prostate gland growth and development are not solely dependent on epithelial AR but require paracrine signaling induced by the AR-positive prostate mesenchyme^[13,14]. Disruptions in the functional coupling between the stromal and epithelial interactions in the adult prostate are associated with glandular dysfunction typified in aging and disease, although specific mechanisms remain undefined. Considerable evidence suggests these biological disruptions are associated with a transition into a repair-centric, emergency, or emergent tissue systems biology state to affect rapid repair until the biological priority resets to functional, differentiated biology^[15]. However, an unresolved/chronic repair state of the stroma, defined as reactive stroma, has been correlated with promoting the evolution of pathological processes including benign prostate hyperplasia (BPH) and prostate cancer^[15-18] [Figure 1].

Currently, there is very limited information available on both tissue-resident immune cells in healthy adult prostate and the modulations that occur in the immune landscape in a disease state like cancer. Often co-evolving with cancer, the reactive stroma is an environment enriched with growth factors and characterized by increased angiogenesis, an increased inflammatory response, and an extensively remodeled ECM resulting in a desmoplastic reaction^[11,19]. The reactive stroma of solid tumors, including prostate cancer, has been shown to be immunosuppressive and associated with induced resistance to tumor-targeted immunotherapies; however, the mechanisms remain complex^[20,21]. Recent studies have shown that either reprogramming subsets of stromal cells or immunotherapies targeting stromal antigens can disrupt the pro-tumorigenic microenvironment niche and enhance endogenous or vaccine-induced antitumor immunity^[20-24]. In this review, we will discuss the current knowledge about stromal evolution in prostate cancer tumorigenesis and its known regulation of the tissue-immune landscape. Understanding this will help in developing effective therapeutic strategies that can be leveraged to co-target peri-tumoral reactive stroma to reprogram the immune suppressive tumor microenvironment (TME) and render it permissive to antitumor immunotherapies.

PROSTATE GLAND AT HOMEOSTASIS

An integrative network involving various cellular and acellular components regulates the structure, function, and homeostasis of the prostate gland. The acinar epithelial cells constitute the functional

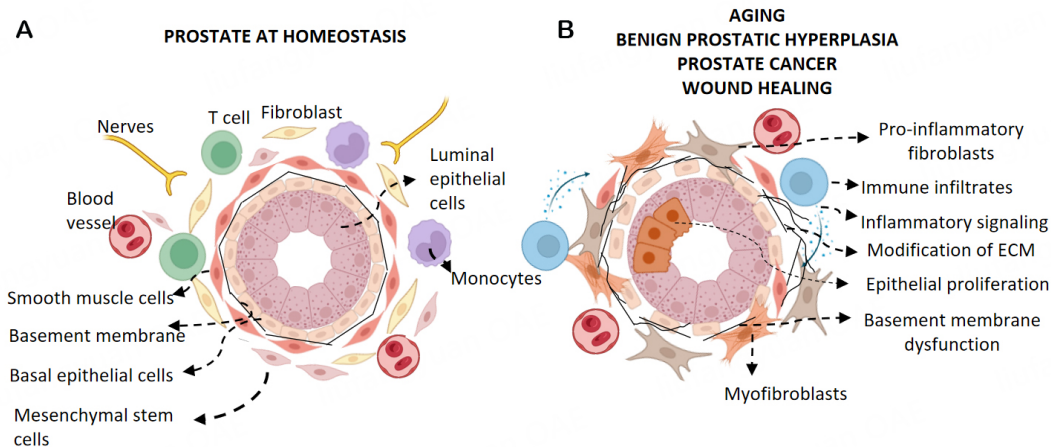


Figure 1. Reactive/emergent stromal response in prostate. (A) The prostate gland at homeostasis maintains a well-organized tissue architecture with specific cellular components functioning in a balanced state. (B) Aging and conditions like benign prostatic hyperplasia (BPH) and cancer disrupt this homeostasis, triggering an emergency/emergent (repair) processes in the prostate tissue to restore homeostasis.

parenchyma of the prostate gland. The stromal cells synthesize ECM components and provide mechanical support to the secretory epithelium. In addition to these functions, immune cells actively participate in the surveillance of organ integrity, while the vascular system provides oxygen and nutrient support to the organ. Moreover, the contractile activity of the smooth muscle of the stroma is pivotal for the proper functioning of the prostate gland. This activity is regulated by neuronal inputs, with the sympathetic nervous system acting via the hypogastric nerve, and parasympathetic nervous system via the pelvic nerve^[1,2,25].

The bulk stroma of the prostate consists of fibroblasts, mesenchymal stem cells, and smooth muscle cells. The homeostasis and normal functioning of the prostate gland are dependent on intercellular communications between the epithelial and stromal compartment, mediated through the paracrine and apocrine secretions from both cell types^[25-30]. Various secretory effectors like wntless-related integration sites (WNTs), transforming growth factors (TGF) α and β , fibroblast growth factors (FGF), insulin growth factors (IGF), epidermal growth factor (EGF), platelet-derived growth factors (PDGF), vascular endothelial growth factor (VEGF), prostaglandins, endothelin, sonic hedgehog, and nitrous oxide orchestrate cellular proliferation, differentiation, and the regulation of cell death in both epithelial and stromal cells of the prostate gland. These processes are mediated via the respective cognate receptors^[27,30,31]. Furthermore, a well-defined laminin-positive basement membrane demarcates the epithelial acini from the fibromuscular stroma of the prostate gland^[32]. Along with key junctional complexes within the epithelium, the structural integrity of this basement membrane is crucial for maintaining the functionality of the acini and preventing the dissemination of pathogenic microbes into the stromal tissue. A breach in basement membrane integrity also serves as a critical precursor to invasive progression and systemic pathogenesis like prostate cancer metastases^[33,34].

Steroid regulation of the prostate

Steroids, particularly testosterone, play an important role in maintaining the structural and functional integrity of the prostate gland. At the subcellular level, androstenedione is also essential for maintaining the function of prostatic epithelium. In addition to androgens, estrogen contributes to the regulation of prostate function by primarily affecting stromal cell proliferation and angiogenesis^[2,31,35,36]. The impact of estrogen on prostatic epithelium is intricate. While it has been associated with inducing hyperplasia, metaplasia, and

keratinization of the epithelium, as well as neoplasia in the prostate of adult rodent models^[37-39], studies have also shown the administration of estrogen-induced BPH in dogs. Moreover, the estrogenic effect has been demonstrated to induce aberrations in the prostatic epithelium in both primates and humans^[39,40]. Therefore, maintaining the estrogen/testosterone (E/T) ratio is integral for maintaining normal function and homeostasis of the prostate gland^[39]. Besides androgens and estrogens, progesterone, prolactin (a hypophyseal hormone), and insulin regulate prostatic function and growth^[2].

Steroid action in the prostate gland is mediated through respective intracellular hormone receptors. The human prostate expresses AR, estrogen receptors α and β (ER α and ER β), estrogen-responsive G protein-coupled receptor 30 (GPR30), progesterone receptor (PR), and glucocorticoid receptor (GR)^[2]. Following puberty, the steady state phase of the prostate gland is maintained by balancing cell proliferation and cell death, a process regulated by AR signaling in both the epithelium and stromal cells of the prostate^[41]. During the prenatal and postnatal differentiated stage, ER α is primarily expressed in the stroma and smooth muscle cells, while ER β is expressed in the epithelium^[42-44]. The compartmentalization of expression and the differential affinity of both ER α and β to bind to ligands and cofactors suggests the diverse functional role of estrogen within the prostate gland. Experiments in mice show that ER α -regulated transcription of cytokine genes in the mesenchyme regulates prostate differentiation and morphology during development. Meanwhile, estrogen-mediated signaling in the prostate epithelium, mediated via ER β , has been shown to be important for epithelial function^[39].

Isoforms of PR (PRA and PRB) are predominantly expressed in the stromal and smooth muscle cells of the prostate. The interaction with the prostate epithelium is crucial for PR expression in the stromal cells. Notably, PR activation has been observed to inhibit stromal expansion, which contrasts ER α regulation of stromal cells^[45]. These observations suggest the coordinated activity of PR and ER α in maintaining epithelium-stromal homeostasis in the prostate gland; however, the mechanisms remain unclear. Glucocorticoids exert pleiotropic effects systemically through the GR receptors. In the prostate, GR and AR share overlapping cisomes and transcriptomic signatures. AR activation has been shown to downregulate GR expression in the prostate epithelium, indicating a critical negative feedback regulation between these two hormone receptors. Consequently, in castration conditions, GR signaling can bypass AR inhibition, promoting therapeutic resistance and prostate cancer cell survival^[46].

IMMUNE LANDSCAPE IN PROSTATE GLAND

Tumor-infiltrating immune cells influence the progression of prostate cancer and its response to treatment, yet understanding the immune microenvironment crucial for normal prostate function remains limited. Recent studies utilizing bulk and single-cell sequencing techniques on normal and non-cancerous prostate tissue have identified a diverse array of leukocytes, including mononuclear phagocytes (MNPs), mast cells, natural killer (NK) cells, B cells, and tissue-resident T cells. Cross-species analysis in mice has revealed that T cells and MNPs, which persist even in prostate cancer, primarily populate the healthy prostate. Cross-analyses of single-cell RNA sequencing datasets in humans have identified six distinct classes of MNPs in the normal prostate, including monocytes, conventional dendritic cells (cDC1 and cDC2 subsets), proliferating macrophages, and various macrophage subclasses, including MAC1, MAC2, and MAC-MT^[47,48].

Tissue-resident macrophages are crucial during embryonic development and in maintaining adult tissue homeostasis^[49]. The diversity observed among MNPs within the prostate suggests potential tissue-specific functions. Transcriptomic analyses reveal that certain macrophage subclasses, such as MAC-MT, are exclusive to the prostate gland and exhibit heightened expression of zinc transporter genes, implicating their

involvement in zinc homeostasis critical for prostatic fluid synthesis^[50,51]. Remarkably, targeted depletion of macrophages using antibody targeting colony-stimulating factor 1 receptor (CSF1R) resulted in decreased zinc concentration specifically within the prostate gland, confirming the presumed role of MAC-MT macrophages in zinc regulation^[47]. Furthermore, the distinct functions of dendritic cell subsets, such as cDC1 and cDC2, in antigen presentation and T-cell activation, respectively, underscore the dual importance of MNPs in both maintaining prostatic function and regulating immune responses^[47,48].

Contrary to conventional theory, there is growing evidence to suggest that tissue-resident macrophages are seeded in the embryonic stage and self-maintained throughout adulthood. Transcriptomic analyses revealed that MAC-MT, with upregulated levels of zinc transporters genes, *SLC39A8* and *SLC30A1*, might be seeded in the prostate from the prenatal stage, as it is transcriptionally similar to the yolk sac-derived macrophages^[47]. The expression of the zinc transporters and metallothionein genes were also identified to be highly expressed in embryonically seeded macrophages in the murine prostate gland, further highlighting the developmental origin of these specialized immune cells. Conversely, MAC2 was suggested to be monocyte-derived and specific to the prostate gland^[47-49,52]. These insights underscore the intricate interplay between immune cell populations and tissue-specific functions within the prostate, providing valuable insights into potential therapeutic targets for prostate cancer and related disorders.

HORMONAL DYSREGULATION IN AN AGING PROSTATE

Hormonal imbalances, tissue atrophy, and chronic inflammation are characteristic features of an aging prostate^[53]. Additionally, with aging, there is an increased likelihood of transitioning from adult differentiated biology to repair-centric/emergent systems biology in tissues, resulting in the activation of reactive stromal response. This reactive stroma plays a crucial role in regulating epithelial proliferation and modulating the immune microenvironment. Notably, hormonal dysregulation is one of the primary contributors to the transformation of the prostate stroma into a reactive phenotype^[53].

Aging men experience an upregulation of estrogen production due to declining testosterone levels, a process exacerbated by comorbidities like obesity and type 2 diabetes^[53-56]. Testosterone deficiency is implicated in inducing chronic inflammation within the prostate tissue, as testosterone plays a crucial role in inhibiting the pro-inflammatory response of prostate stromal cells by activating AR and inhibiting the secretion of inflammatory cytokines and growth factors^[57,58]. Additionally, testosterone protects against inflammation caused by uropathogenic bacteria like *Escherichia coli* by downregulating the Janus Kinases (JAK)/signal transducer and activator of transcription 1 (STAT1) signaling pathway in the prostate epithelium^[59]. Indeed, chronic inflammation of the prostate gland induced by bacteria was observed to induce premalignant and malignant lesions in the prostate gland of Mongolian gerbils^[60].

Obesity exacerbates hormonal imbalance by increasing aromatase activity, leading to the conversion of testosterone to estradiol, the most potent form of estrogen in men^[61]. The increase in the estrogen/testosterone (E/T) ratio due to aging can lead to estrogen dominance, promoting stromal cell proliferation and fibrosis, which can accelerate clinical progression in BPH, and induce premalignant lesions in the prostate gland^[62,63]. Racial disparities in prostate cancer incidence and mortality rates, particularly among African American men, have been linked to dysregulated estrogenic action on the prostate gland, with higher serum levels of estradiol observed in Non-Hispanic black men compared to Non-Hispanic white men^[39,64-68]. Additionally, exposure to elevated estrogen levels during early gestation has been suggested to be a contributing factor to racial differences in prostate cancer risk, which needs to be substantiated through population-based studies. However, *in vivo* rodent studies support the induction of abnormalities in the prostate gland by early estrogenic exposure^[37,69-71].

Multiplex profiling has revealed a significant enrichment of T and B lymphocytes in the mouse prostate gland due to aging. Specifically, a strong correlation was observed between age and enrichment of programmed cell death protein 1 positive (PD-1⁺) CD4⁺ and CD8⁺ T cells in the mouse prostate gland. PD-1, a cell membrane protein, plays a critical role in inhibiting both T and B-cell immune response, and is a marker for T-cell "exhaustion"^[47,53,72]. Intriguingly, estrogen modulates immune cells of both myeloid and lymphoid lineages in a tissue-context-dependent manner^[73]. Given the presence of both myeloid and lymphoid cells in prostate gland homeostasis^[47,48], understanding the dynamic changes in the immune profile within the prostate gland in the context of testosterone deficiency and estrogenic dominance is crucial for comprehending disease development in an aging prostate.

TUMOR-STROMA INTERACTIONS

While tissue-emergent/reactive stromal responses in carcinomas are often considered secondary to epithelial changes, the limited progression of many epithelial tumors from *in situ* lesions despite harboring genetic abnormalities associated with malignancy raises questions about the driving molecular factors of neoplasms^[74,75]. Recombinant studies in murine models have provided evidence suggesting that the stromal microenvironment is a key determinant in promoting prostate carcinogenesis^[76-80]. Moreover, exogenous insults directly affecting the stroma have been identified as critical initiators of the carcinogenic process in various other solid tumors. For instance, ultraviolet radiation-induced dermal atrophy has been shown to precede keratinocyte tumors, while chemicals in cigarette smoke metabolically promote cancer by inducing autophagy and premature aging in the host stromal microenvironment in an organ such as the breast^[75,81,82]. In another example, obesity-induced metabolites derived from gut microbiota induce senescence in hepatic stellate cells, which then secrete inflammatory and tumor-promoting factors that facilitate the development of hepatocellular carcinoma in mice exposed to chemical carcinogens^[83]. Collectively, these instances suggest that the tissue stroma may indeed play a primary role in initiating and promoting cancer development.

Thus, the experimental evidence cited above suggests that aging or insult-driven changes of the stroma create a permissible emergent/reactive tissue or organ environment (soil) that promotes the growth of monoclonal or polyclonal tumors (field cancerization). However, based on this view, sustainable treatment or cure for cancer will be difficult to attain as long as the reactive soil persists^[75]. Therefore, there is a pressing need to characterize the stromal compartment of solid tumors. One of the main limitations in characterizing reactive stroma in prostate cancer is the heterogeneity of cancer-associated fibroblasts (CAFs) that make up the TME. Stromal heterogeneity is partially explained by the fact that CAFs can be derived from the activation of tissue-resident fibroblasts, mesenchymal stem cells, vimentin-positive periacinar cells, circulating bone marrow-derived precursors, vessel-associated pericytes, and endothelial cells^[10,11]. Spatial transcriptomic analysis of radical prostatectomy-derived tissue, in addition to stromal cell lineages identified from single-cell sequencing analyses of mouse prostate stroma, suggests the presence of reactive stromal cells with different transcriptional programs and functions within the prostate cancer TME^[84,85]. Additionally, the phenotypic plasticity of the activated stromal cells further underscores the dynamic nature of the reactive stroma^[11]. Hence, characterizing a moving target such as TME to understand tumorigenesis, development, and progression becomes a challenging endeavor.

STROMAL RESPONSE IN PROSTATE CANCER

The coordinated host emergent response to tissue injury involves the collective action of cells that make up the connective tissue/stroma and the extracellular matrix (ECM) products. The normal reactive stromal response to injury is self-limited and regulated spatially and temporally to re-establish tissue integrity and reset homeostasis. The mechanisms underlying reactive/emergent stromal response include the release of

inflammatory and growth signals, basement membrane dysfunction initiating cell-to-matrix and intercellular interactions regulating cell proliferation, migration, and differentiation of both stromal and epithelial cells, fibroplasia, angiogenesis, ECM remodeling, and wound contraction [Figure 1]. However, the persistence of this repair process resulting in a chronic, non-healing wound and fibrosis, can affect any tissue and organ system in the human body^[86-88].

In 1863, Rudolf Virchow first recognized the association between wound healing phases in tissue and tumorigenesis. Clinical similarities suggested shared common cellular and molecular signatures between the two conditions. This insight led Scottish pathologist Dr. Alexander Haddow to deduce cancers as wounds that overheat^[89,90]. However, clinical evidence indicated that in cancer, reactive stroma is not self-limited and is tumor-promoting, which led Dr. Harold Dvorak to postulate that "tumors are wounds that do not heal"^[86]. Owing to Paget's "seed and soil hypothesis", reactive stroma has emerged as essential soil regulating multiple aspects of tumorigenesis, including initiation, development, progression to metastases, and most importantly, development of therapeutic resistance^[91].

Reactive stroma is heterogeneous in its makeup, exhibiting both organ- and tumor-specific characteristics. The prevalence and abundance of reactive stroma serve as disease-defining factors and are associated with poor prognosis in several solid tumors, including colon carcinoma, head and neck cancer, HER2-negative early breast cancer, squamous cell carcinoma, and rectal cancer^[20,92-96]. In prostate cancer, reactive stroma co-evolves with tumor development, and its relative abundance is quantified as reactive stromal grade (RSG). An RSG of 3 represents when more than 50% of the prostate tumor area is composed of reactive stroma, and the latter is associated with earlier biochemical recurrence and worse prognosis^[97,98]. Reactive stroma in prostate cancer is composed of CAFs that can transdifferentiate into cancer-associated myofibroblasts (myCAF) or inflammatory fibroblasts (iCAF), with an expanded and modified ECM with collagen deposition, dense microvessels, and immune infiltrates [Figure 1]^[11,17,99]. Tissue recombinant experiments demonstrated that prostate cancer-derived CAFs promote tumor growth *in vivo* while normal fibroblasts inhibit the process, thus confirming the critical nature of reactive stroma in prostate tumor growth and development^[100-102].

MOLECULAR FEATURES OF REACTIVE STROMA

TGF- β signaling

Cytokines, such as TGF- β , play a crucial role in regulating cell fate and reactive stromal response. TGF- β induces the differentiation of stromal cells into vimentin and smooth muscle alpha-actin-positive myCAF, thereby initiating a wound repair-like reactive stroma. Concurrently, TGF- β also modulates the composition of the ECM by inducing the expression of collagen 1 and tenascin-C in stromal cells^[10,17]. TGF- β is also critical in modulating the immune reaction. It can suppress interleukin 2 (IL-2) synthesis and T-cell proliferation, as well as regulate the differentiation of both CD4⁺ T cells and regulatory T cells (Tregs). Cytokines such as IL-10, IL-4, and TGF- β secreted by reactive stromal cells and immune-suppressive cells like Tregs can increase the polarization of M2 macrophages. M2 macrophages have been shown to enhance angiogenesis, tissue remodeling, and modulate the immune microenvironment by expressing human leukocyte antigen (HLA-DR) and programmed death-ligand 1 (PD-L1), resulting in the suppression of the immune system^[103-105].

Although the expression of TGF- β increases from prostatic intraepithelial neoplasia (PIN) to the development of prostate cancer lesions^[10,17], the biological activity of TGF- β within TME is not cancer-specific. TGF- β is a fundamental regulator of different cellular processes in adult differentiated biology^[106]. Thus, the biological activity of TGF- β activity is essentially indistinguishable whether in a wound repair,

tissue fibrosis, or TME. TGF- β , secreted by both epithelial cells and fibroblasts, is critical for coordinating tissue repair and homeostasis through its context-dependent pleiotropic functions. For example, TGF- β induces cytostasis in non-transformed epithelial cells, while in endothelial and mesenchymal cells, it stimulates proliferation. Additionally, TGF- β can both induce and suppress apoptosis, suggesting that other critical signaling inputs mediate TGF- β -induced cellular survival and cell death^[107]. Epithelial-to-mesenchymal transition (EMT) is an important cellular event common in wound repair, fibrosis, and cancer. TGF- β -induced signaling, either independently or in coordination with critical mediators of tissue homeostasis such as Wnt and NOTCH, induces EMT by repressing inter-epithelial adhesion and junctional complexes while upregulating mesenchymal markers including ECM remodeling enzymes like matrix metalloproteinases (MMPs), facilitating cell migration^[108]. Apart from TGF- β , growth factors such as FGFs and PDGF, which are critical for wound healing and tissue remodeling, also regulate reactive stromal biology in prostate cancer^[109,110].

Remodeled ECM

ECM remodeling serves as another critical regulator of tissue biology, integrity, and, most importantly, maintaining homeostasis. Beyond providing physical support, the ECM plays a pivotal role in cell adhesion, migration, initiating angiogenesis, tissue development, and repair^[111]. The cells of the reactive stroma actively overexpress ECM proteins (collagens, elastin, fibronectin, tenascin-C, and hyaluronic acid), MMPs, fibroblast activation protein (FAP), and lysyl oxidases^[17,111-113]. MMPs, in particular, play a critical role in remodeling the ECM and inducing modifications that influence its topographical and mechanical properties. In healthy tissue, ECM remodeling is a tightly regulated process that involves the counterbalance of ECM synthesis with the activity of MMPs, MMP inhibitors, and lysyl oxidases^[114].

Damage to the ECM due to tissue injury initiates an emergent response referred to as a "wound healing cascade" to repair damaged tissue and reset tissue homeostasis^[111]. In cancer, ECM is damaged and exposure to chronic remodel signals generated from both the cancer cells and reactive stromal cells results in a chronic wound-healing cascade. Consequently, the dysregulated ECM remodeling and turnover results in the development of fibrotic tissue (desmoplasia) with enhanced stiffening around the tumors. This ECM alteration affects every aspect of tumor biology, including the regulation of proliferation, differentiation, gene expression, cell adhesion, migration, invasion, *etc.*^[111]. Beyond its regulatory control, ECM also influences immunogenicity, oxygenation, and the response of cancer to treatments. Thus, a reactive stroma composed of extensively remodeled ECM is correlated with poor prognosis in cancer^[111,115].

Activated angiogenic niche

Cells within the vasculature network, primarily endothelial cells (EC), pericytes, and vascular smooth muscle cells, constitute additional critical components of the tissue stroma and are essential regulators of prostate homeostasis^[10]. In healthy tissue, the non-angiogenic EC interacts with a complex basement membrane composed of collagen IV, laminin, perlecan (heparan sulfate proteoglycan), and entactin/nidogen via integrin^[116]. However, in response to wounding and cancer, the reactive stromal cells (myofibroblasts or CAFs) secrete ECM such as collagen I and IV, fibronectin, secreted protein acidic and rich in cysteine (SPARC), tenascin, heparan sulfate proteoglycans, connective tissue growth factor, and VEGF. On interaction with these reactive stromal products, the quiescent EC transitions into an activated status/angiogenic switch, resulting in the formation of phenotypically distinct blood vessels with aberrant branching and enhanced leakiness^[10,116]. Hypoxia is another critical feature common to both tissue injury and TME. Hypoxia-induced activation of hypoxia-inducible factors (HIFs) in both epithelial and stromal cells results in the secretion of proangiogenic factors that modulate vessel maturation. Thus, the new growth of vascular network in response to reactive stromal response in the TME is critical in regulating the proliferation, growth, and progression of solid tumors^[117-119].

In addition to providing nutritional support and oxygen supply, activated EC express chemokines and adhesion molecules, facilitating the recruitment of leukocytes, monocytes, and neutrophils^[120,121]. Pericytes, a class of tissue-resident mesenchymal stem cells, play a critical role in the stabilization of neovessels and regulating MMP activity^[10,99,116]. In both wound repair and cancer, pericytes deviate from the EC and vascular basement membrane, undergoing a phenotypic transformation that regulates angiogenesis, inflammation, fibrosis, tissue regeneration, and re-epithelialization^[122,123]. All these functional aspects of pericytes in an emergent wound repair scenario are critical in establishing a pre-metastatic niche, which is critical for tumor growth and progression^[124].

STROMAL MEMORY

The concept of cellular "memory" can be defined as when cells maintain an altered phenotypic or functional state proceeding with an initial environmental stimuli/insult. Myeloid lineage cells like monocytes, NK cells, macrophages, and neutrophils exhibit innate immune memory, a manifestation of cellular memory^[125,126]. Research over the past two decades has demonstrated that these cells exhibit protective or cross-protective mechanisms against recurring infections through heightened activation of the innate immune response. This heightened response is driven by pattern recognition receptors (PRRs) on myeloid cells, allowing recognition of pathogen-associated molecular patterns (PAMPs). Activation of PRRs by PAMPs triggers the expression of genes involved in inflammatory and immune responses^[127-129].

Fibroblast memory

Emerging evidence suggests that non-immune cells also possess a memory of past insults such as inflammation, enabling them to mount rapid responses to emergent situations like injury or infection. The biological mechanisms regulating cellular memory are multifaceted, involving various processes such as alterations in chromosomal accessibility due to epigenetic modifications, increase in expression of activation receptors, and priming of cellular signaling networks^[121,130-133].

Naik *et al.* were the first to discover that epithelial stem cells exposed to inflammation retain cellular memory, leading to enhanced repair responses to future tissue-related injuries^[121,134]. This phenomenon extends to fibroblasts, which develop an inflammatory memory upon exposure to exogenous challenges such as lipopolysaccharide (LPS) or endogenous inflammatory signals like tumor necrosis factor α (TNF- α)^[121]. For instance, human gingival fibroblasts pretreated with LPS showed no tolerance but maintained cytokine and chemokine expression after secondary LPS treatment^[135]. Similarly, in conditions like rheumatoid arthritis, fibroblasts such as synoviocytes exhibit gene-specific priming by altering chromatin following chronic exposure to inflammatory signals like TNF- α , leading to enhanced and prolonged chemokines and cytokine production upon subsequent interferon (γ) stimulation^[132,136]. Klein *et al.* have also observed that LPS primes synovial fibroblasts to sustain inflammatory responses by changing the epigenetic configuration at gene promoters regulating LPS-induced cellular responses^[131].

In patients with tendinopathy, stromal fibroblast activation markers such as podoplanin and vascular cell adhesion molecule (VCAM-1) are notably elevated compared to healthy tendon tissues. This elevation persists even after the gradual decline in inflammatory gene signatures following the removal of stimuli like IL-1 β , suggesting that activated fibroblast memory maintains a persistent activated state rather than sustaining inflammatory responses^[130]. Additionally, in rheumatoid arthritis, sustained synovial inflammation is also attributed to persistent activation of nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) signaling induced by TNF- α , facilitated by upregulation of TNF receptors and proximal signaling components, and downregulation of negative feedback inhibitors involved in the homeostatic balance of the NF- κ B signaling pathway^[136]. Epigenetic modifications are also known to regulate epithelial stem cell memory^[121,131,132,134].

Endothelial cell memory

Similar to fibroblasts, EC is also activated in response to inflammation. EC activation can be distinguished as a "delayed (type II)" or "immediate (type I)" response. In the delayed response, upon activation, EC expresses chemokines, VCAM-1, intercellular adhesion molecule (ICAM-1), and E-selectin several hours post-stimulus due to a requirement for *de novo* transcription and translation^[137,138]. In contrast, in the type I response, there is no delay following stimulation and response due to the preformation of adhesion molecules and chemoattractant resulting from a preceding inflammatory stimulus. This suggests EC memory^[138-141]. Studies have demonstrated that EC exposed to homocysteine, an independent risk factor for developing atherosclerosis, has an augmented response to inflammatory mediators such as LPS and thrombin^[142]. Additionally, recent studies have also shown that EC stores a metabolic memory of an earlier transient hyperglycemia in the vasculature in diabetic patients, resulting in epigenetic changes, cardiovascular complications, chronic inflammation, and oxidative stress in later stages^[143-145].

As integral components of the tissue structure, the ability of stromal cells to adapt to environmental stimuli while retaining memory of past exposures is essential for maintaining tissue homeostasis^[125,146,147]. Various environmental stimuli, such as injury or infection, can trigger the innate cellular memory in stromal cells, leading to an activated phenotypic and functionally emergent state through different molecular mechanisms, as described. This emergent microenvironment state of tissues in diseases like cancer can have a detrimental impact on disease initiation and development, progression, and response to treatments^[21,125,148-150]. For instance, chronic inflammation of organs due to injury-causing agents or infections is known to induce cancers, such as esophageal, lung, gastric, and colon cancer, which are often metastatic, treatment-resistant, and lethal^[151-158]. Therefore, targeting specific stromal memories involved in maintaining a reactive/emergent stromal response emerges as a promising therapeutic strategy to mitigate the detrimental effects of microenvironment priming and enhance treatment efficacy in cancer patients^[21,125,159].

IMMUNE REGULATION IN CANCER

Traditionally, cancer research has focused on the intrinsic biology of cancer cells to identify potential molecular determinants crucial for tumor growth, development, and progression. However, there has been a recent shift in attention toward the role of non-cancerous cellular components in the TME, particularly immune cells, in controlling tumor growth and development. This shift has been clinically validated and garnered attention because of its potential to be curative in subsets of cancer patients^[150]. Consequently, there has been an increase in research efforts aimed at understanding the mechanisms regulating the reactivity of immune cells toward various types of tumors. This shift in focus reflects a growing recognition of the intricate interplay between cancer cells and the TME, highlighting the importance of comprehensively understanding the latter for the development of effective cancer therapies.

Paul Ehrlich's hypothesis on immune cells suppressing carcinoma development led to the "immune surveillance hypothesis" later proposed by Burnet and Thomas^[160]. While the tumor-specific immune response was validated in inbred mouse strains, discordant results from immune-deficient mouse models initially cast doubt on the concept^[161]. However, the development of defined immune-deficient models and epidemiological data from human studies reaffirmed the relevance of cancer immune surveillance, leading to the broader concept of "cancer-immuno-editing". This concept recognizes the dual role of host-protecting and tumor-sculpting properties of the immune system. Cancer-immuno-editing involves three stages: tumor elimination by the immune system (immune surveillance), a phase of equilibrium where

tumor variants resistant to surveillance are selected (immune-sculpting), and the emergence of overt tumors in an immunocompetent host (immune-escape)^[161-167].

Role of stroma in immunomodulation of the TME

The stroma plays a pivotal role in immunomodulation, posing a significant challenge to immunotherapies in various solid tumors like pancreatic duct adenocarcinoma (PDAC), non-small cell lung cancer, ovarian cancer, hepatocellular carcinoma, and prostate cancer^[23,168-172]. However, targeting the stroma to enhance immunotherapy efficacy and hinder tumor progression has been largely overlooked. Understanding the complex interplay between tumor-stroma-immune components is crucial for developing innovative therapies to modulate the TME and improve targeted cancer treatments^[21,161].

Stromal cells significantly contribute to cancer-immuno-editing by modulating the immune system through the secretion of various chemical messengers, such as chemokines, cytokines, and prostaglandins, as well as the ECM. In PDAC, elevated levels of the chemokine-chemokine (C-X-C motif) ligand 10 (CXCL10), positively correlated with high stromal content, are associated with decreased median overall survival in patients. CXCL10 expression is linked to the presence of Tregs, which exerts immunosuppressive effects, compromising immune surveillance against cancer^[21,173,174]. Additionally, the immune regulatory chemokine CXCL5 secreted by tumor-associated macrophages (TAMs), CAFs, EC, and cancer cells themselves plays a crucial role in recruiting neutrophils to the TME. Neutrophils, in turn, modulate the TME, promoting tumor growth and progression, and induce anti-inflammatory M2 macrophage polarization, impairing immune surveillance. High expression of CXCL5 is associated with poor patient survival in various cancers, including renal, pancreatic, liver, and cervical cancer^[21,175-177].

In solid tumors, myCAF-expressing FAP exhibits immunosuppressive properties by secreting large amounts of stromal cell-derived factor-1 (SDF-1), hindering T-cell-tumor interactions and attracting Tregs. Ablation of FAP⁺ stromal cells leads to hypoxia-induced cancer cell death mediated by interferon- γ and TNF- α ^[178-181]. Additionally, CAF-secreted TGF- β inhibits host immune surveillance by impairing dendritic cell, M1 macrophage, NK cells, and CD8⁺ T-cell function, while promoting Treg and Th17 cell differentiation and suppressing B cell proliferation and IgA secretion^[182,183]. Moreover, TGF- β restricts T-cell infiltration, diminishing tumor response to PD-L1 blockade^[184].

TGF- β induces ECM remodeling, while its suppression of ECM-modulating proteins like MMP-1, -8, and -13 results in the formation of fibrotic and desmoplastic ECM matrix, which is associated with cancer recurrence and chemoresistance^[185-187]. Desmoplasia impedes T-cell recruitment into tumor nests, causing T-cell accumulation in peri-tumoral regions and promoting immune escape. Additionally, within these peri-tumoral regions, T cells are exposed to paracrine signals, resulting in their suppression^[23]. The matricellular protein periostin (POSTN) is highly expressed by both tumor and stromal cells. Its elevated expression is associated with poor prognosis in various cancers, including prostate, lung, pancreatic, ovarian, breast, colorectal, hepatocellular, bladder, and osteosarcoma^[188]. POSTN promotes PD-1 expression in TAMs via integrin-ILK-NF- κ B signaling. PD-1-expressing TAMs were observed to induce PD-L1 expression in colorectal cancer cells, promoting immune escape^[189]. Collectively, the reactive stromal response plays a critical role in tumor development, progression, and modulation of the immune landscape in TME [Figure 2].

THE IMMUNE LANDSCAPE IN PROSTATE CANCER

In the past decade, the understanding of the immune landscape in cancer has evolved significantly. Studies have utilized cell surface markers to identify various immune cell populations within the TME, including

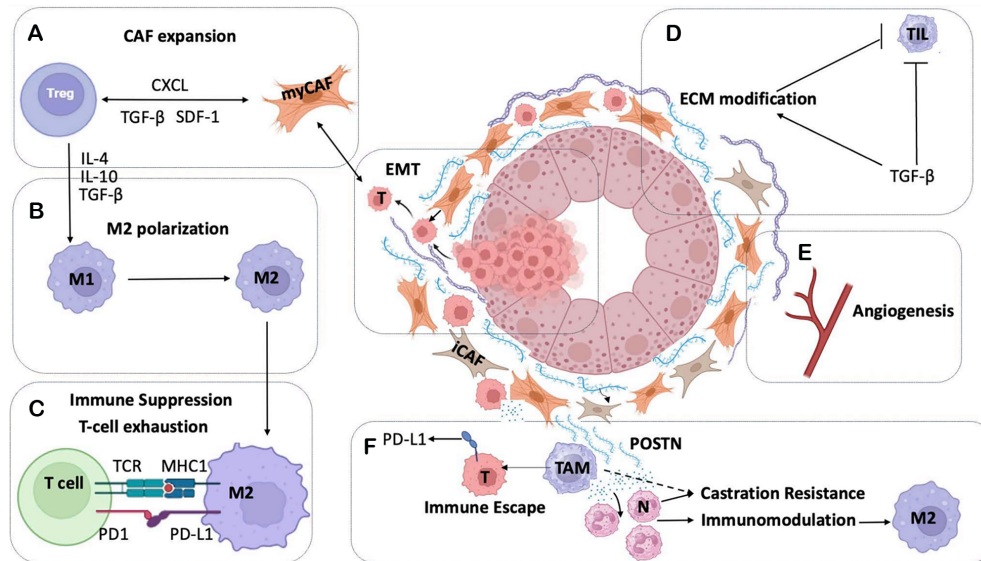


Figure 2. Emergent stromal response regulate immunosuppressive landscape in solid tumors. (A) The reciprocal interactions between tumor cells (T) and expanding CAFs within the reactive TME results in the secretion of TGF- β and various chemokines (CXCL) facilitating the recruitment and regulation of Tregs. (B) Treg secreted cytokines (IL-4, IL-10 and TGF- β) trigger polarization of M1 macrophages to the M2 phenotype. (C) PD-L1-expressing M2 macrophages induce T-cell exhaustion. (D) TGF- β in the TME derived from different cellular sources can modulate the extracellular matrix (ECM) composition. Both TGF- β and the modified ECM impedes tumor-infiltrating lymphocytes (TIL) both molecularly and mechanically. (E) The reactive stroma induces angiogenesis, further supporting tumor growth and survival. (F) Matricellular protein-periostin (POSTN), expressed during reactive stromal response attract TAMs. TAMs facilitate immune evasion in tumor cells by inducing the expression of PD-L1. TAMs also recruit neutrophils, which induces an immunosuppressive TME by causing M2 polarization. Both TAM and neutrophils also induce therapeutic resistance.

T cells, B cells, NK cells, macrophages, monocytes, and granulocytes. Solid tumors are commonly classified as having "hot" or "cold" immune microenvironments based on the presence or absence of these immune cell populations within the tumor margins^[167,190,191]. Prostate tumors are typically classified as having a "cold" TME, characterized by elevated PD-L1 expression and lower levels of tumor-infiltrating immune cells, like CD3⁺ T cells, CD20⁺ B cells, and CD68⁺ macrophages compared to BPH^[192,193]. In addition to elevated levels of Tregs and myeloid-derived suppressor cells (MDSCs), contributing to an immunosuppressive microenvironment in prostate cancer^[167,194], the disease also exhibits reduced tumor antigens due to its low tumor mutational burden^[195]. Additionally, AR signaling suppresses major histocompatibility complex 1 (MHC1) expression and T-cell response, further complicating the development of immunotherapy to target prostate cancer^[167,196-198].

In primary prostate cancer, both the cancerous epithelium and stromal cells express inflammatory factors like TNF- α and IL-6, which induce reactive oxygen species, leading to inflammation, immunosuppression, and tissue damage^[199]. TNF- α and IL-6 promote treatment-resistant/castration-resistant prostate cancer (CRPC) by affecting stromal and prostate cancer cells^[199,200]. IL-6 specifically, mediated by bone morphogenic protein (BMP) and CD105, induces androgen receptor splice variant 7 (AR-V7) expression in prostate cancer cells and fibroblasts, a key mechanism in CRPC progression. IL-6-mediated AR-V7 expression in fibroblasts induced resistance to anti-AR inhibitors in prostate cancer cells. However, neutralizing CD105 downregulated AR-V7 in both prostate cancer cells and fibroblasts, resensitizing the cancer cells to these inhibitors^[201]. Additionally, paracrine interactions between epithelial and stromal cells stimulate prostate stromal cells to secrete chemokines such as CXCL-1, CXCL-2, CXCL-3, and IL-8 (CXCL-8)^[199], which recruit leukocytes like neutrophils, macrophages, monocytes, and MDSCs into the microenvironment via C-X-C chemokine receptor type 2 (CXCR2) activation^[202-204]. In a murine prostate

cancer model, CXCR2 knockout or antagonist administration reduced tumor growth and shifted tumor-associated macrophages toward a pro-inflammatory M1 phenotype^[205]. Elevated neutrophil count correlates with worse overall survival in localized prostate cancer^[206], while elevated IL-8 levels in prostate cancer patients with metastatic disease starting on androgen deprivation therapy (ADT) were associated with shorter progression time to castration resistance and overall survival^[207]. Therefore, understanding the mechanisms governing the inflammatory and immunosuppressive TME from localized disease to metastasis is essential for developing effective immunotherapies for advanced prostate cancer.

To characterize the immune microenvironment in solid tumors like prostate cancer, numerous scientific groups and biotechnological companies have devoted considerable efforts to developing tools for genetic, transcriptomic, metabolic, and proteomic-based profiling of the immune landscape. These approaches encompass serum, spatial, and single-cell-based techniques^[47,208-212]. Notably, there have been significant strides in developing immune-related gene signatures to elucidate key immune cell components within prostate cancer, such as macrophage-based gene signatures^[210], metabolic syndrome-based index scores^[213], immune subtyping^[214,215], immune-based risk scoring^[216], and long non-coding RNA signatures^[217]. These advancements significantly contribute to the capacity to develop biomarkers for diagnostic and prognostic purposes, enhancing the ability to assess the risk, progression, and sensitivity to immunotherapies in prostate cancer.

Research efforts aimed at transforming "cold" immune microenvironments into "hot" ones in solid tumors, including prostate cancer, are underway to enhance the efficacy of immune checkpoint inhibitors (ICIs) like PD-1/programmed death-ligand 1 (PD-L1) or cytotoxic T-lymphocyte-associated antigen 4 (CTLA4) inhibitors^[197]. However, a significant challenge in this strategy, particularly in prostate cancer, is the reactive stromal response, which can molecularly and mechanically shield tumor cells from the antitumor immune response. Additionally, the dysregulation of critical enzymes involved in cellular energetics within tumors, including prostate cancer, is emerging as a hallmark feature associated with tumor evasion, though the mechanisms are complex^[218,219]. Therefore, comprehending the dynamic intercellular crosstalk between tumor-stroma-immune cells and its subsequent modulation of the TME, which can exclude T-cell infiltration or inhibit T-cell function, may be crucial for achieving sustainable efficacy with immunotherapies, including ICIs, in prostate cancer^[23,220].

CURRENT STATE OF PROSTATE CANCER IMMUNOTHERAPIES

The current landscape of immune-based therapeutics for prostate cancer encompasses several approaches, including cancer vaccines, ICIs, adoptive cell therapies, targeted antibodies, and oncolytic viral therapy^[196,221-225]. Cancer vaccines aim to stimulate the patient's immune system, eliciting a response against tumor-specific or tumor-associated antigens (TAA). One notable example is Sipuleucel-T, an FDA-approved autologous vaccine that utilizes dendritic cells (DC) stimulated to target prostatic acid phosphatase (PAP), a protein highly expressed in prostate cancer. Clinical data show evidence of improved median survival and prolonged overall survival among men with metastatic castration-resistant prostate cancer (mCRPC) treated with Sipuleucel-T compared to those treated with mainline treatments (anti-hormone treatment ± chemotherapy). While Sipuleucel-T treatment has demonstrated a broad and durable systemic immune response, clinical data suggest that the treatment provides greater benefits to patients with a lower disease burden. This observation may be attributed to the existence of a robust immune system at the initial stages of cancer development, in contrast to more advanced disease states. In addition, treatment with Sipuleucel-T did not significantly affect mCRPC disease progression. This may be due to the delayed onset of antitumor response by Sipuleucel-T, and therefore, a timeline of diagnosis of mCRPC may play an important role in determining the maximum possible benefit from Sipuleucel-T treatment^[226,227]. Additional

mechanistic underpinnings that resulted in the limited efficacy of DC vaccine for clinical management of prostate cancer are described further in this review. However, several vaccine targets are currently under evaluation against prostate cancer, including oncofetal antigen-5T4, carcinoembryonic antigen (CEA), PSA, prostate-specific membrane antigen (PSMA), survivin, tumor-associated antigens (TAA), and personalized neoantigens. These selected proteins serve as targets due to their high expression levels in cancer cells compared to their normal counterparts^[228].

Other prominent strategies in prostate cancer immunotherapy involve the use of immune modulators to disrupt immune checkpoints, such as PD-1/PD-L1 and CTLA-4, exploited by cancer cells to evade immune detection and responses, often leading to T-cell exhaustion^[196]. In addition to these well-established targets, ongoing clinical investigations in other urological cancer types, such as bladder cancer, explore immune modulators targeting the immunosuppressive activity of CD73, indoleamine 2,3-dioxygenase (IDO), and lymphocyte activation gene 3 (LAG3)^[229-232] (Clinical Trial Registration Numbers NCT03454451, NCT05843448, NCT04586244). Immunomodulation treatments also include the activation of co-stimulatory pathways to promote or enhance T-cell functions by downregulating immunosuppressive components like Tregs in the TME. Key targets in this category include inducible co-stimulator (ICOS), OX40, Toll-like receptors (TLRs), CD137, and IL-2/IL-2R^[233-236].

Adoptive or cell-based immunotherapy represents another autologous approach, where the patient's immune cells, such as T cells, are isolated, expanded *in vitro*, and modified with chimeric antigen receptors (CARs) that can specifically target antigens expressed by tumor cells, thereby eliminating them. Adoptive immunotherapy targets currently under evaluation for prostate cancer include prostate stem cell antigen (PSCA) and PSMA^[237]. Beyond T cells, both NK and tumor-infiltrating lymphocytes (TILs) can also be enhanced and reintroduced into patients^[238,239]. Monoclonal antibodies constitute another class of treatments developed to block specific cell membrane receptors from binding to its target ligand, thereby impeding its functional impact on cancer growth and proliferation. Commonly targeted membrane receptors in prostate cancer include delta-like proteins (DLL), Notch, human epidermal growth factor receptor 2 (HER2), and tumor-associated calcium signal transducer 2 (TROP2)^[240-242]. Antibodies can also be modified to carry cytotoxic payloads, specifically chemotherapeutics, for their active delivery to tumors^[243]. Bi-specific T-cell-engaging antibodies or BiTEs bind to cancer cells and T cells, activating the latter^[244]. Oncolytic viral therapy involves the use of different DNA (Adenovirus and Herpes simplex virus) and RNA (Reovirus) viruses, often modified to infect tumor cells and induce cell death. This approach can elicit an immune response that further aids in the elimination of both localized and metastatic tumors^[245].

While appealing, the immunosuppressive and "cold" TME in prostate cancer poses a significant challenge for the immunotherapy strategies described. Therefore, emerging therapeutic approaches aim to target both cancer cells and the TME. One strategy involves directly targeting stromal markers upregulated in the TME, such as FAP, which is associated with poor prognosis in various solid tumors. Targeting FAP-expressing CAFs using CAR-T therapy shows promise in improving tumor-targeted cytotoxicity of CAR-T cells targeting solid tumors. In summary, therapies targeting both cancer cells and the TME hold the potential for effectively treating prostate cancer and improving patient outcomes. [Table 1](#) provides a summary of various immunotherapies currently under evaluation in clinical trials, while [Table 2](#) outlines potential therapies for targeting tumor stroma^[20,245,263].

MACROPHAGE THERAPY FOR METASTATIC CASTRATION RESISTANCE PROSTATE CANCER

Immunotherapy exhibits limited efficacy in advanced prostate cancer patients with mCRPC, mainly due to

the immunosuppressive TME. Targeting TAMs, the predominant immunosuppressive cells in prostate cancer, presents a promising therapeutic avenue for mCRPC treatment^[264]. Macrophages, once viewed as phagocytic sentinels, now demonstrate diverse roles in maintaining tissue homeostasis. Tissue-resident macrophages oversee the microenvironment, ensuring tissue integrity, facilitating cellular communication, and regulating immunological balance^[265]. Conversely, monocyte-derived macrophages, recruited during inflammation, adopt either the pro-inflammatory "M1" or anti-inflammatory/reparative "M2" phenotypes, each characterized by distinct gene expression and metabolic pathways. "M1" macrophages rely on glycolysis, producing inflammatory cytokines like IL-1 β , IL-12, TNF- α , and reactive oxygen species, while "M2" macrophages employ oxidative phosphorylation, secreting molecules such as arginase-1 and TGF- β ^[266-268].

Despite the simplicity of the "M1/M2" dichotomy, current single-cell transcriptomic data suggest a more complex landscape, acknowledging the high degree of macrophage plasticity and tissue-specificity^[269]. This complexity has propelled macrophage reprogramming to the forefront as a promising therapeutic. Macrophages' dynamic transition between "M1" and "M2" states in response to environmental cues has become a focal point in disease treatment strategies, particularly by directing them toward an "M1" phenotype to initiate inflammation and restore homeostasis.

In mCRPC, TAMs play dual roles: facilitate tumor progression, and induce immunosuppression within the TME^[270]. Research also underscores TAMs' significant function in fostering resistance to anti-androgen therapies. For instance, macrophages can induce ECM remodeling, reminiscent of wound healing processes. Such macrophage-mediated ECM modifications correlate with anti-androgen resistance, particularly through the activation of fibronectin-1 (FN1)-integrin alpha 5 (ITGA5)-tyrosine kinase Src signaling cascade, induced by the cytokine Activin-A^[271]. Reprogramming these TAMs to an M1-like state may disrupt these pro-tumorigenic activities. Transitioned M1 macrophages could potentially reverse the immunosuppressive TME, attenuate ECM-mediated drug resistance, and amplify the efficacy of current therapies^[272].

Upregulated Src kinase activity in prostate cancer bone metastases, mediated by the Activin-A Receptor as well, is associated with macrophage density and several ECM-receptor pathways^[273]. Targeting this activity with the specific Src inhibitor eCF506 has shown promise in blocking enzalutamide resistance, emphasizing its therapeutic potential in mCRPC management^[271]. Furthermore, strategies that modulate TAMs can engender a more antitumoral phenotype^[274]. ICIs can induce M1 macrophage polarization, and the depletion of Treg cells - protectors of the tumor-friendly milieu - by anti-CTLA-4 antibodies is contingent upon macrophage-mediated actions^[275]. These insights affirm the premise that macrophage reprogramming could play a crucial role in enhancing the impact of immunotherapies in mCRPC.

An alternate, emerging approach is through chimeric antigen receptor macrophages (CAR-M), which leverages the natural tumor-homing ability of myeloid cells. This therapy has shown potential advantages in infiltrating solid tumors and can release pro-inflammatory cytokines to improve the TME^[276]. Furthermore, the combination of CAR-M with CAR-T cells has demonstrated synergistic action against cancer cells, exceeding the effects of either therapy alone. This synergy suggests that CAR-M and CAR-T can complement each other, enhancing tumor responses. Despite its promising antitumor activity demonstrated in animal experiments, CAR-M therapy faces several challenges that need to be addressed. These include optimizing the CAR structure by incorporating tandem activation domains or pro-inflammatory cytokines to enhance its effectiveness and safety for clinical application^[277].

Table 1. Selected clinical trials of immunotherapies targeting cancer

Type	Treatment	Combination	Target	Tumor type	Phase	Status	Identifier
Peptide vaccine	NY-ESO-1 Protein	CpG 7909	NY-ESO-1	Adv. PCa	I	Complete	NCT00292045
	UV1	GM-CSF	hTERT	CSPC, mPC	I/II	Unknown status	NCT01784913
mAb	Denosumab	N/A	RANKL	non-metastatic CRPC	III	Complete	NCT00286091
Bisphosphonate	Zoledronic acid	N/A	Osteogenic niche	mCRPC and mBC	IV	Recruiting	NCT04549207
Inhibitors	Ibrutinib	N/A	BTK, MMP-2, MMP-9	PCa	II	Complete	NCT02643667
	Ibrutinib	Trastuzumab	BTK, MMP-2, MMP-9	HER2 ⁺ BC	I/II	Ongoing	NCT03379428
Selected clinical trials of macrophage immunotherapies							
M2/M1 reprogramming	LY3022855	N/A	M-CSFR	mCRPC, mBC	I	Complete	NCT02265536
	MCS110	Carboplatin, gemcitabine	M-CSF	TNBC	II	Complete	NCT02435680
	IPI-549	Tecentriq, abraxane, bevacizumab	PI3K- γ	BC, renal cell carcinoma	II	Ongoing	NCT03961698
	852A	N/A	TLR7	BC, ovarian, endometrial, and cervical cancers	II	Complete	NCT00319748
Antibodies	Imiquimod	Abraxane	TLR7	Adv. BC	II	Complete	NCT00821964
	CP-870,893	N/A	CD40	Adv. solid tumors	I	Complete	NCT02225002
	CP-870,893	Paclitxel, carboplatin	CD40	Solid tumors	I	Complete	NCT00607048
	Hu5F9-G4	Cetuximab	CD47/SIRPa	Solid tumors, Adv. CC	I/II	Complete	NCT02953782
CAR-M	PLX3397	Eribulin	CSF-1R	BC	I/II	Complete	NCT01596751
	CT-0508	N/A	HER2	HER2 ⁺ solid tumors, including PCa	I	Recruiting	NCT04660929
Cytokines	GM-CSF	Carboplatin, cabazitaxel	HSCs	mNEPC, mPC	II	Recruiting	NCT04709276
	Proscavax (GM-CSF, PSA, IL-2)	N/A	PSA	PCa	II	Unknown status	NCT03579654
Inhibitors	Cabiralizumab	Paclitaxel, carboplatin, nivolumab	CSF-1R	TNBC	I/II	Ongoing	NCT04331067
	Daratumumab	N/A	CSF-1R	PCa	I	Ongoing	NCT03177460
	Carlumab	N/A	CCL2	PCa	II	Complete	NCT00992186
	AZD-5069	Enzalutamide	CXCR2	mCRPC	I/II	Terminated	NCT03177187
Dendritic cell therapies for PCa							
DC Vaccine	Sipuleucel-T	N/A	PAP	mCRPC	III	Complete	NCT00065442
	Stapuldencel-T	Docetaxel, prednisone	PAP	mCRPC	III	Complete	NCT02111577
	With tumor mRNA	N/A	hTERT, survivin	mCRPC	I/II	Ongoing	NCT01197625

Thus, the intricate relationship between macrophages, ECM components, and other immune cells within the TME underscores the potential of therapeutic interventions that manipulate these interactions, particularly through M1/M2 reprogramming and CAR-M therapy. Such a strategy could potentially disrupt critical resistance mechanisms and forge a more robust immune response against tumors in their advanced stages.

THE PITFALLS OF DENDRITIC CELL THERAPY IN PROSTATE CANCER

DC therapy has encountered significant challenges, primarily attributed to the inconsistency in clinical responses^[278]. This is compounded by the fact that no DC therapies have received FDA approval since

Table 2. Potential immunotherapies targeting the TME

TME target	Molecular target	Immunotherapies	Potential desired effects	References
Fibroblasts	FAP	FAP-CAR-T; FAP BiTEs	Destroy CAFs to disrupt tumor-stroma interactions and suppress tumor growth	[246-249]
	SDF-1/CXCL12	CDXR4 antagonist (Plerixafor)	Disrupt SDF-1 signaling to reduce tumor growth and metastasis	[250]
	HGF/c-MET	mAb or nAb	Inhibit tumor growth and metastasis	[251]
Endothelium	VEGF	Immunomodulatory agents (thalidomide; lenalidomide)	Inhibit angiogenesis and cancer-stroma adherence, and stimulate the immune system	[252,253]
		Bevacizumab; ramucirumab	Inhibit angiogenesis to reduce tumor blood supply	[254,255]
	PDGF	Olaratumab	Inhibit stromal cell recruitment and activation to disrupt angiogenesis, stromal support, and bone metastasis	[256]
ECM	MMPs	MMP inhibitors	Inhibit ECM remodeling to reduce tumor cell migration	[257]
	Collagen	Collagenase	Reduce ECM stiffness and density to improve drug and immune cell infiltration	[258]
	TNC	mIL12-R6N mAb	Antitumor activity	[259]
	CTGF	mAb	Modulate the TME to reduce fibrosis and enhance the efficacy of other treatments	[260]
	Integrins	mAb	Disrupt cell-ECM interactions, inhibiting tumor cell migration	[261]
	FN1	mAb	Inhibit cell proliferation and migration	[262]
	TGF- β	nAb	Reduce immunosuppression and increase immune cell infiltration in the TME	[262]

Sipuleucel-T in 2010. While certain patients exhibit enhanced immune reactions and positive clinical outcomes, such as reduced PSMA levels and tumor regression, these effects are not universally observed^[279,280]. This variability raises questions about the therapy's reliability, making its therapeutic value uncertain.

The effectiveness of DC therapy hinges on the selection of appropriate TAAs and the successful maturation of the DC themselves. Common TAAs in prostate cancer - such as PSA, PSMA, PAP, and PSCA - show variable expression across different tumors, affecting the efficacy of the therapy^[281,282]. Additionally, the immunogenicity of these antigens may not always be sufficient to induce a strong immune response^[283]. Compounding these issues are the technical complexities in producing functionally mature DCs. The *in vitro* generation process, influenced by factors like the source of DCs, culture conditions, and maturation stimuli, is intricate and can significantly affect the therapy's success. Hence, the lack of standardized protocols for antigen selection and DC maturation further complicates the development of an effective therapy^[284].

The prostate cancer TME presents another hurdle, often characterized by immunosuppressive elements that can impede the activity of cytotoxic T cells, undermining the effectiveness of DC therapy. Furthermore, even when initial immune responses are elicited, sustaining these responses over time remains a challenge, frequently leading to disease progression^[285]. This lack of durable response necessitates repeated administrations or combination therapies, increasing treatment complexity and costs. The dynamic nature of the TME, with its evolving mechanisms of immunomodulation, makes it a moving target for DC therapy. Efforts to understand and manipulate this environment could be key to enhancing the therapy's effectiveness and durability.

DC therapy directly targeting the stromal compartment itself in prostate cancer offers a novel therapeutic avenue. For instance, immunotherapies directly targeting CAFs for depletion or reprogramming have the

potential to reduce or eliminate tumor-promoting and immunosuppressive properties. However, this strategy faces challenges due to heterogeneity, phenotypic plasticity, and complex interactions within the tumor-stroma ecosystem, which can impede the efficacy of the immune response. Understanding and effectively manipulating these interactions are crucial for the success of DC therapy in targeting the stroma, potentially leading to more effective control of prostate cancer growth and metastases^[286].

Despite being generally safe, DC therapy can induce adverse events like flu-like symptoms, injection site reactions, and potential autoimmunity^[287,288]. These adverse events require careful monitoring to ensure patient safety. In conclusion, while DC therapy in prostate cancer represents a significant advancement in cancer immunotherapy, it is constrained by challenges such as inconsistent clinical responses, antigen selection, DC maturation difficulties, immune suppressive TME, transient immune responses, and safety issues. An important future approach should be focused on identifying biomarkers that could predict responses to DC therapy, thereby refining patient selection. Additionally, exploring synergies between DC therapy and other immunomodulatory approaches may unlock new avenues for more effective and comprehensive cancer treatment strategies.

SUMMARY

The survival of an organism is dependent on the maintenance of robust and dynamic systemic homeostatic mechanisms regulating physiological responses to both internal stimuli (wound repair, inflammation, and diseases) and external stimuli (food, pathogens, toxic pollutants, and drugs). Homeostasis is coordinated by the different functional systems within the body via a multitude of long-range (endocrine), short-range (paracrine, juxtacrine, neuronal signaling at synaptic junctions), and self (autocrine) cellular signaling. At the tissue level, the intracellular machinery of tissue-resident cells needs to coordinate and integrate complex signals from cellular and non-cellular components of the tissue environment. However, in the case of cancer, accumulating genomic or epigenetic aberrations in cancerous cells can decouple their functional interactions within a tissue, resulting in the development of neoplasia. Despite the dependence of cancer cells on multicellular interactions with the respective TME as well as systemic physiological environments, collectively referred to as "systems biology of cancer", conventional research continues to follow a reductionist approach, predominantly focusing on cancer-specific intracellular factors, overlooking broader systems influences on cancer pathobiology^[279,280].

Although localized or locally advanced prostate cancer patients undergo definitive therapy with curative intent, up to 50% experience recurrence, progressing to mCRPC. While immunomodulatory therapies like ICIs have advanced as first- or second-line treatments, yielding promising results in various cancers, including non-small cell lung cancer (NSCLC) and colorectal cancer, their efficacy in advanced prostate cancer remains limited. Prostate cancer's immunologically "cold" TME underscores the need for a systems biology approach to identify and characterize spatial and temporal TME signatures. These signatures are crucial determinants in immunomodulation, disease progression, and treatment response^[7,281-283].

In this review, we have highlighted the critical role of reactive stromal response in the evolution of cancer pathobiology through immunomodulation of the TME [Figure 2]. The reactive stromal response is an emergency/emergent response of the tissue to undergo rapid repair and reset homeostasis, a biological priority. The multifaceted role of reactive stromal cells includes their intercellular communication with tissue-resident and immune cells in the systemic circulation to enhance tissue repair while minimizing damage. Thus, to preserve homeostatic balance, the reactive stromal response encompasses immune-regulatory functions^[284]. Therefore, adopting a cancer systems biology approach is crucial for fully grasping the dynamic interactions within the reactive/repair-centric TME. This methodology will enable researchers

to identify key regulatory factors influencing cancer progression, survival, and resistance to immunotherapy, ultimately uncovering potential therapeutic targets. For instance, by integrating phenotypic and functional variations of the TME with molecular characteristics of cancer, an integrative model can be developed to delineate dynamic immunomodulation concurrent with tumor progression at the systems level. This approach will facilitate the discovery of new biomarkers that can be used for predicting immunotherapy response, aid in patient stratification, and inform on effective drug combinations to overcome drug resistance. However, a drawback of systems biology is its reliance on large sets of high-quality patient data collected over various time scales and concepts, which necessitates advanced downstream analyses and computations. Therefore, developing cost-effective and accessible technologies with user-friendly algorithms to integrate data from different omics platforms can revolutionize personalized cancer pathobiology modeling. These integrative, hypothesis-driven, and predictive models can advance our understanding of disease mechanisms and improve personalized treatment strategies^[279,280].

DECLARATIONS

Authors' Contributions

Conceptualized and designed the structural framework of the manuscript, performed comprehensive literature review, drafted the original manuscript, critically reviewed and revised the manuscript for intellectual content, approved the final version of the manuscript: Thomas R

Contributed to the conceptualization of the study; performed comprehensive literature review, and collaborated on writing of the manuscript: Jerome M, Krieger K

Contributed to the final revisions of the manuscript, critically reviewed the manuscript for intellectual content and accuracy of the information reported particularly in [Tables 1](#) and [2](#): Jerome M, Krieger K, Ashraf N

Contributed significantly to the conceptualization of the manuscript, critically reviewed the content and provided crucial feedback on the manuscript, contributed to the comprehensive revisions of the initial and final draft of the manuscript, approved the final version of the manuscript: Rowley D

Availability of data and materials

Not applicable.

Financial support and sponsorship

This study was supported by National Cancer Institute of Health (NCI) (grant No: R01 CA221946, P30 CA125123), Early Investigator Research Award from the Department of Defense Prostate Cancer Research Program (W81WXH-21-1-0154), National Institutes of Health (K99MD018671), and CPRIT RP210027-Baylor College of Medicine Comprehensive Cancer Training Program.

Conflicts of interest

All authors declared that there are no conflicts of interest.

Ethical approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

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REFERENCES

1. Cunha GR, Donjacour AA, Cooke PS, et al. The endocrinology and developmental biology of the prostate. *Endocr Rev* 1987;8:338-62. DOI
2. Kumar VL, Majumder PK. Prostate gland: structure, functions and regulation. *Int Urol Nephrol* 1995;27:231-43. DOI PubMed
3. Singh O, Bolla SR. Anatomy, abdomen and pelvis, prostate. Treasure Island: StatPearls; 2023. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK540987/> [Last accessed on 22 Jul 2024].
4. Rawla P. Epidemiology of prostate cancer. *World J Oncol* 2019;10:63-89. DOI PubMed PMC
5. Schrecengost R, Knudsen KE. Molecular pathogenesis and progression of prostate cancer. *Semin Oncol* 2013;40:244-58. DOI PubMed PMC
6. Siegel RL, Giaquinto AN, Jemal A. Cancer statistics, 2024. *CA Cancer J Clin* 2024;74:12-49. DOI PubMed
7. Shore ND, Moul JW, Pienta KJ, Czernin J, King MT, Freedland SJ. Biochemical recurrence in patients with prostate cancer after primary definitive therapy: treatment based on risk stratification. *Prostate Cancer Prostatic Dis* 2024;27:192-201. DOI PubMed PMC
8. Krušlin B, Ulamec M, Tomas D. Prostate cancer stroma: an important factor in cancer growth and progression. *Bosn J Basic Med Sci* 2015;15:1-8. DOI PubMed PMC
9. Levesque C, Nelson PS. Cellular constituents of the prostate stroma: key contributors to prostate cancer progression and therapy resistance. *CSH Perspect Med* 2018;8:a030510. DOI PubMed PMC
10. Pederzoli F, Raffo M, Pakula H, Ravera F, Nuzzo PV, Loda M. Stromal cells in prostate cancer pathobiology: friends or foes? *Br J Cancer* 2023;128:930-9. DOI PubMed PMC
11. Barron DA, Rowley DR. The reactive stroma microenvironment and prostate cancer progression. *Endocr Relat Cancer* 2012;19:R187-204. DOI PubMed PMC
12. Davey RA, Grossmann M. Androgen receptor structure, function and biology: from bench to bedside. *Clin Biochem Rev* 2016;37:3-15. PubMed PMC
13. Wen S, Chang HC, Tian J, Shang Z, Niu Y, Chang C. Stromal androgen receptor roles in the development of normal prostate, benign prostate hyperplasia, and prostate cancer. *Am J Pathol* 2015;185:293-301. DOI PubMed PMC
14. Cunha GR. Epithelio-mesenchymal interactions in primordial gland structures which become responsive to androgenic stimulation. *Anat Rec* 1972;172:179-95. DOI PubMed
15. Schauer IG, Rowley DR. The functional role of reactive stroma in benign prostatic hyperplasia. *Differentiation* 2011;82:200-10. DOI PubMed PMC
16. Schauer IG, Ressler SJ, Tuxhorn JA, Dang TD, Rowley DR. Elevated epithelial expression of interleukin-8 correlates with myofibroblast reactive stroma in benign prostatic hyperplasia. *Urology* 2008;72:205-13. DOI PubMed PMC
17. Tuxhorn JA, Ayala GE, Smith MJ, Smith VC, Dang TD, Rowley DR. Reactive stroma in human prostate cancer: induction of myofibroblast phenotype and extracellular matrix remodeling. *Clin Cancer Res* 2002;8:2912-23. PubMed
18. Tuxhorn JA, Ayala GE, Rowley DR. Reactive stroma in prostate cancer progression. *J Urol* 2001;166:2472-83. DOI PubMed
19. Kai F, Drain AP, Weaver VM. The extracellular matrix modulates the metastatic journey. *Dev Cell* 2019;49:332-46. DOI PubMed PMC
20. Valkenburg KC, de Groot AE, Pienta KJ. Targeting the tumour stroma to improve cancer therapy. *Nat Rev Clin Oncol* 2018;15:366-81. DOI PubMed PMC
21. Puré E, Lo A. Can targeting stroma pave the way to enhanced antitumor immunity and immunotherapy of solid tumors? *Cancer Immunol Res* 2016;4:269-78. DOI PubMed PMC
22. Quail DF, Joyce JA. Microenvironmental regulation of tumor progression and metastasis. *Nat Med* 2013;19:1423-37. DOI PubMed PMC
23. Xiao Z, Todd L, Huang L, et al. Desmoplastic stroma restricts T cell extravasation and mediates immune exclusion and immunosuppression in solid tumors. *Nat Commun* 2023;14:5110. DOI PubMed PMC
24. Zhang FF, Qiao Y, Xie Y, et al. Epitope-based minigene vaccine targeting fibroblast activation protein α induces specific immune responses and anti-tumor effects in 4 T1 murine breast cancer model. *Int Immunopharmacol* 2022;112:109237. DOI
25. Gorbach IN, Novikov DK. [Detection of leukocyte sensitization to tuberculosis mycobacterial antigens in newborns by the method of migration suppression in vitro]. *Vopr Okhr Materin Det* 1976;21:66-8. PubMed
26. Giacomini A, Grillo E, Rezzola S, et al. The FGF/FGFR system in the physiopathology of the prostate gland. *Physiol Rev* 2021;101:569-610. DOI
27. Cunha GR. Mesenchymal-epithelial interactions: past, present, and future. *Differentiation* 2008;76:578-86. DOI PubMed
28. Roberson KM, Edwards DW, Chang GC, Robertson CN. Isolation and characterization of a novel human prostatic stromal cell culture: DuK50. *In Vitro Cell Dev Biol Anim* 1995;31:840-5. DOI PubMed
29. Webber MM, Trakul N, Thraves PS, et al. A human prostatic stromal myofibroblast cell line WPMY-1: a model for stromal-epithelial interactions in prostatic neoplasia. *Carcinogenesis* 1999;20:1185-92. DOI

30. Simons BW, Hurley PJ, Huang Z, et al. Wnt signaling though beta-catenin is required for prostate lineage specification. *Dev Biol* 2012;371:246-55. [DOI](#) [PubMed](#) [PMC](#)
31. Farnsworth WE. Prostate stroma: physiology. *Prostate* 1999;38:60-72. [DOI](#)
32. Mostofi FK, Sesterhenn IA, Davis CJ. A pathologist's view of prostatic carcinoma. *Cancer* 1993;71:906-32. [DOI](#) [PubMed](#)
33. Steukers L, Glorieux S, Vandekerckhove AP, Favoreel HW, Nauwynck HJ. Diverse microbial interactions with the basement membrane barrier. *Trends Microbiol* 2012;20:147-55. [DOI](#) [PubMed](#) [PMC](#)
34. Brekken RA, Stupack D. Extracellular matrix in tumor biology. In: *Biology of extracellular matrix*. Cham: Springer International Publishing; 2017. [DOI](#)
35. Welén K, Damber JE. Androgens, aging, and prostate health. *Rev Endocr Metab Disord* 2022;23:1221-31. [DOI](#) [PubMed](#) [PMC](#)
36. Stanworth RD, Jones TH. Testosterone for the aging male; current evidence and recommended practice. *Clin Interv Aging* 2008;3:25-44. [DOI](#)
37. Prins GS, Huang L, Birch L, Pu Y. The role of estrogens in normal and abnormal development of the prostate gland. *Ann N Y Acad Sci* 2006;1089:1-13. [DOI](#) [PubMed](#) [PMC](#)
38. Shapiro E, Huang H, Masch RJ, McFadden DE, Wilson EL, Wu XR. Immunolocalization of estrogen receptor alpha and beta in human fetal prostate. *J Urol* 2005;174:2051-3. [DOI](#) [PubMed](#)
39. Prins GS, Korach KS. The role of estrogens and estrogen receptors in normal prostate growth and disease. *Steroids* 2008;73:233-44. [DOI](#) [PubMed](#) [PMC](#)
40. Coffey DS, Walsh PC. Clinical and experimental studies of benign prostatic hyperplasia. *Urol Clin North Am* 1990;17:461-75. [DOI](#) [PubMed](#)
41. Vickman RE, Franco OE, Moline DC, Vander Griend DJ, Thumbikat P, Hayward SW. The role of the androgen receptor in prostate development and benign prostatic hyperplasia: a review. *Asian J Urol* 2020;7:191-202. [DOI](#) [PubMed](#) [PMC](#)
42. Schulze H, Claus S. Histological localization of estrogen receptors in normal and diseased human prostates by immunocytochemistry. *Prostate* 1990;16:331-43. [DOI](#) [PubMed](#)
43. Schulze H, Barrack ER. Immunocytochemical localization of estrogen receptors in the normal male and female canine urinary tract and prostate. *Endocrinology* 1987;121:1773-83. [DOI](#) [PubMed](#)
44. Prins GS, Birch L. Neonatal estrogen exposure up-regulates estrogen receptor expression in the developing and adult rat prostate lobes. *Endocrinology* 1997;138:1801-9. [DOI](#) [PubMed](#)
45. Yu Y, Liu L, Xie N, et al. Expression and function of the progesterone receptor in human prostate stroma provide novel insights to cell proliferation control. *J Clin Endocrinol Metab* 2013;98:2887-96. [DOI](#) [PubMed](#) [PMC](#)
46. Arora VK, Schenkein E, Murali R, et al. Glucocorticoid receptor confers resistance to antiandrogens by bypassing androgen receptor blockade. *Cell* 2013;155:1309-22. [DOI](#) [PubMed](#) [PMC](#)
47. Tuong ZK, Loudon KW, Berry B, et al. Resolving the immune landscape of human prostate at a single-cell level in health and cancer. *Cell Rep* 2021;37:110132. [DOI](#) [PubMed](#) [PMC](#)
48. Henry GH, Malewska A, Joseph DB, et al. A cellular anatomy of the normal adult human prostate and prostatic urethra. *Cell Rep* 2018;25:3530-3542.e5. [DOI](#) [PubMed](#) [PMC](#)
49. Mass E, Ballesteros I, Farlik M, et al. Specification of tissue-resident macrophages during organogenesis. *Science* 2016;353:aaf4238. [DOI](#) [PubMed](#) [PMC](#)
50. Costello LC, Franklin RB. A comprehensive review of the role of zinc in normal prostate function and metabolism; and its implications in prostate cancer. *Arch Biochem Biophys* 2016;611:100-12. [DOI](#) [PubMed](#) [PMC](#)
51. Singh KK, Desouki MM, Franklin RB, Costello LC. Mitochondrial aconitase and citrate metabolism in malignant and nonmalignant human prostate tissues. *Mol Cancer* 2006;5:14. [DOI](#) [PubMed](#) [PMC](#)
52. Ginhoux F, Guilliams M. Tissue-resident macrophage ontogeny and homeostasis. *Immunity* 2016;44:439-49. [DOI](#) [PubMed](#)
53. Cannarella R, Condorelli RA, Barbagallo F, La Vignera S, Calogero AE. Endocrinology of the aging prostate: current concepts. *Front Endocrinol* 2021;12:554078. [DOI](#) [PubMed](#) [PMC](#)
54. Feldman HA, Longcope C, Derby CA, et al. Age trends in the level of serum testosterone and other hormones in middle-aged men: longitudinal results from the Massachusetts male aging study. *J Clin Endocrinol Metab* 2002;87:589-98. [DOI](#)
55. O'Donnell AB, Araujo AB, McKinlay JB. The health of normally aging men: The massachusetts male aging study (1987-2004). *Exp Gerontol* 2004;39:975-84. [DOI](#) [PubMed](#)
56. Huhtaniemi I. Late-onset hypogonadism: current concepts and controversies of pathogenesis, diagnosis and treatment. *Asian J Androl* 2014;16:192-202. [DOI](#) [PubMed](#) [PMC](#)
57. Rastrelli G, Vignozzi L, Corona G, Maggi M. Testosterone and benign prostatic hyperplasia. *Sex Med Rev* 2019;7:259-71. [DOI](#) [PubMed](#)
58. Vignozzi L, Cellai I, Santi R, et al. Antiinflammatory effect of androgen receptor activation in human benign prostatic hyperplasia cells. *J Endocrinol* 2012;214:31-43. [DOI](#)
59. Ho CH, Fan CK, Yu HJ, et al. Testosterone suppresses uropathogenic escherichia coli invasion and colonization within prostate cells and inhibits inflammatory responses through JAK/STAT-1 signaling pathway. *PLoS One* 2017;12:e0180244. [DOI](#) [PubMed](#) [PMC](#)
60. Quintar AA, Gonçalves BF, Taboga SR, Maldonado CA. The mongolian gerbil (*Meriones unguiculatus*) as a model for inflammation-promoted prostate carcinogenesis. *Cell Biol Int* 2017;41:1234-8. [DOI](#)
61. Cohen PG. Obesity in men: the hypogonadal-estrogen receptor relationship and its effect on glucose homeostasis. *Med Hypotheses*

- 2008;70:358-60. DOI PubMed
62. Yang Y, Sheng J, Hu S, et al. Estrogen and G protein-coupled estrogen receptor accelerate the progression of benign prostatic hyperplasia by inducing prostatic fibrosis. *Cell Death Dis* 2022;13:533. DOI PubMed PMC
63. Ellem SJ, Risbridger GP. Aromatase and regulating the estrogen: androgen ratio in the prostate gland. *J Steroid Biochem Mol Biol* 2010;118:246-51. DOI
64. Rebbeck TR. Prostate cancer disparities by race and ethnicity: from nucleotide to neighborhood. *Cold Spring Harb Perspect Med* 2018;8:a030387. DOI PubMed PMC
65. Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics, 2021. *CA Cancer J Clin* 2021;71:7-33. DOI
66. Chowdhury-Paulino IM, Ericsson C, Vince R Jr, Spratt DE, George DJ, Mucci LA. Racial disparities in prostate cancer among black men: epidemiology and outcomes. *Prostate Cancer Prostatic Dis* 2022;25:397-402. DOI PubMed PMC
67. Hinata N, Fujisawa M. Racial differences in prostate cancer characteristics and cancer-specific mortality: an overview. *World J Mens Health* 2022;40:217-27. DOI PubMed PMC
68. Rohrmann S, Nelson WG, Rifai N, et al. Serum estrogen, but not testosterone, levels differ between black and white men in a nationally representative sample of Americans. *J Clin Endocrinol Metab* 2007;92:2519-25. DOI
69. Henderson BE, Bernstein L, Ross RK, Depue RH, Judd HL. The early in utero oestrogen and testosterone environment of blacks and whites: potential effects on male offspring. *Br J Cancer* 1988;57:216-8. DOI PubMed PMC
70. Platz EA, Giovannucci E. The epidemiology of sex steroid hormones and their signaling and metabolic pathways in the etiology of prostate cancer. *J Steroid Biochem Mol Biol* 2004;92:237-53. DOI PubMed
71. Giusti RM, Iwamoto K, Hatch EE. Diethylstilbestrol revisited: a review of the long-term health effects. *Ann Intern Med* 1995;122:778-88. DOI PubMed
72. Fox JJ, Hashimoto T, Navarro HI, Garcia AJ, Shou BL, Goldstein AS. Highly multiplexed immune profiling throughout adulthood reveals kinetics of lymphocyte infiltration in the aging mouse prostate. *Aging* 2023;15:3356-80. DOI PubMed PMC
73. Khan D, Ansar Ahmed S. The immune system is a natural target for estrogen action: opposing effects of estrogen in two prototypical autoimmune diseases. *Front Immunol* 2015;6:635. DOI PubMed PMC
74. Eifert C, Powers RS. From cancer genomes to oncogenic drivers, tumour dependencies and therapeutic targets. *Nat Rev Cancer* 2012;12:572-8. DOI PubMed
75. Dotto GP. Multifocal epithelial tumors and field cancerization: stroma as a primary determinant. *J Clin Invest* 2014;124:1446-53. DOI PubMed PMC
76. Cunha GR, Hayward SW, Wang YZ, Ricke WA. Role of the stromal microenvironment in carcinogenesis of the prostate. *Int J Cancer* 2003;107:1-10. DOI
77. Cunha GR, Hayward SW, Wang YZ. Role of stroma in carcinogenesis of the prostate. *Differentiation* 2002;70:473-85. DOI PubMed
78. Wang Y, Sudilovsky D, Zhang B, et al. A human prostatic epithelial model of hormonal carcinogenesis. *Cancer Res* 2001;61:6064-72. PubMed
79. Ricke WA, Ishii K, Ricke EA, et al. Steroid hormones stimulate human prostate cancer progression and metastasis. *Int J Cancer* 2006;118:2123-31. DOI
80. Hayward SW, Wang Y, Cao M, et al. Malignant transformation in a nontumorigenic human prostatic epithelial cell line. *Cancer Res* 2001;61:8135-42. PubMed
81. Hu B, Castillo E, Harewood L, et al. Multifocal epithelial tumors and field cancerization from loss of mesenchymal CSL signaling. *Cell* 2012;149:1207-20. DOI PubMed PMC
82. Salem AF, Al-Zoubi MS, Whitaker-Menezes D, et al. Cigarette smoke metabolically promotes cancer, via autophagy and premature aging in the host stromal microenvironment. *Cell Cycle* 2013;12:818-25. DOI PubMed PMC
83. Yoshimoto S, Loo TM, Atarashi K, et al. Obesity-induced gut microbial metabolite promotes liver cancer through senescence secretome. *Nature* 2013;499:97-101. DOI
84. Berglund E, Maaskola J, Schultz N, et al. Spatial maps of prostate cancer transcriptomes reveal an unexplored landscape of heterogeneity. *Nat Commun* 2018;9:2419. DOI PubMed PMC
85. Kwon OJ, Zhang Y, Li Y, et al. Functional heterogeneity of mouse prostate stromal cells revealed by single-cell RNA-Seq. *iScience* 2019;13:328-38. DOI PubMed PMC
86. Huet E, Jaroz C, Nguyen HQ, et al. Stroma in normal and cancer wound healing. *FEBS J* 2019;286:2909-20. DOI
87. Varga J, Brenner DA, Phan SH. Fibrosis research: methods and protocols. Berlin: Springer; 2008.
88. Clark RAF. The molecular and cellular biology of wound repair. New York: Plenum Press; 1996. DOI
89. Schäfer M, Werner S. Cancer as an overhealing wound: an old hypothesis revisited. *Nat Rev Mol Cell Biol* 2008;9:628-38. DOI PubMed
90. Haddow A. Molecular repair, wound healing, and carcinogenesis: tumor production a possible overhealing? Amsterdam: Elsevier; 1973. pp. 181-234. DOI
91. Langley RR, Fidler IJ. The seed and soil hypothesis revisited--the role of tumor-stroma interactions in metastasis to different organs. *Int J Cancer* 2011;128:2527-35. DOI PubMed PMC
92. Plava J, Cihova M, Burikova M, Matuskova M, Kucerova L, Miklikova S. Recent advances in understanding tumor stroma-mediated chemoresistance in breast cancer. *Mol Cancer* 2019;18:67. DOI PubMed PMC
93. Mesker WE, Junggeburst JM, Szuhai K, et al. The carcinoma-stromal ratio of colon carcinoma is an independent factor for survival

- compared to lymph node status and tumor stage. *Cell Oncol* 2007;29:387-98. DOI PubMed PMC
94. Almagush A, Alabi RO, Troiano G, et al. Clinical significance of tumor-stroma ratio in head and neck cancer: a systematic review and meta-analysis. *BMC Cancer* 2021;21:480. DOI PubMed PMC
 95. He R, Li D, Liu B, et al. The prognostic value of tumor-stromal ratio combined with TNM staging system in esophagus squamous cell carcinoma. *J Cancer* 2021;12:1105-14. DOI PubMed PMC
 96. Zhu Y, Jin Z, Qian Y, Shen Y, Wang Z. Prognostic value of tumor-stroma ratio in rectal cancer: a systematic review and meta-analysis. *Front Oncol* 2021;11:685570. DOI PubMed PMC
 97. Ruder S, Gao Y, Ding Y, et al. Development and validation of a quantitative reactive stroma biomarker (qRS) for prostate cancer prognosis. *Hum Pathol* 2022;122:84-91. DOI PubMed PMC
 98. Ayala G, Tuxhorn JA, Wheeler TM, et al. Reactive stroma as a predictor of biochemical-free recurrence in prostate cancer. *Clin Cancer Res* 2003;9:4792-801. PubMed
 99. Tran LL, Dang T, Thomas R, Rowley DR. ELF3 mediates IL-1 α induced differentiation of mesenchymal stem cells to inflammatory iCAFs. *Stem Cells* 2021;39:1766-77. DOI PubMed PMC
 100. Olumi AF, Grossfeld GD, Hayward SW, Carroll PR, Tlsty TD, Cunha GR. Carcinoma-associated fibroblasts direct tumor progression of initiated human prostatic epithelium. *Cancer Res* 1999;59:5002-11. DOI PubMed PMC
 101. Hayashi N, Cunha GR. Mesenchyme-induced changes in the neoplastic characteristics of the Dunning prostatic adenocarcinoma. *Cancer Res* 1991;51:4924-30. PubMed
 102. Proia DA, Kuperwasser C. Stroma: tumor agonist or antagonist. *Cell Cycle* 2005;4:1022-5. DOI
 103. Mun JY, Leem SH, Lee JH, Kim HS. Dual relationship between stromal cells and immune cells in the tumor microenvironment. *Front Immunol* 2022;13:864739. DOI PubMed PMC
 104. Sanjabi S, Oh SA, Li MO. Regulation of the immune response by TGF- β : from conception to autoimmunity and infection. *Cold Spring Harb Perspect Biol* 2017;9:a022236. DOI PubMed PMC
 105. Zhang F, Wang H, Wang X, et al. TGF- β induces M2-like macrophage polarization via SNAIL-mediated suppression of a pro-inflammatory phenotype. *Oncotarget* 2016;7:52294-306. DOI PubMed PMC
 106. Penn JW, Grobbelaar AO, Rolfe KJ. The role of the TGF-beta family in wound healing, burns and scarring: a review. *Int J Burns Trauma* 2012;2:18-28. DOI
 107. Zhang Y, Alexander PB, Wang XF. TGF- β family signaling in the control of cell proliferation and survival. *Cold Spring Harb Perspect Biol* 2017;9:a022145. DOI PubMed PMC
 108. Xu J, Lamouille S, Derynck R. TGF-beta-induced epithelial to mesenchymal transition. *Cell Res* 2009;19:156-72. DOI PubMed PMC
 109. Farooq M, Khan AW, Kim MS, Choi S. The role of fibroblast growth factor (FGF) signaling in tissue repair and regeneration. *Cells* 2021;10:3242. DOI PubMed PMC
 110. Pierce GF, Mustoe TA, Altrock BW, Deuel TF, Thomason A. Role of platelet-derived growth factor in wound healing. *J Cell Biochem* 1991;45:319-26. DOI
 111. Diller RB, Tabor AJ. The role of the extracellular matrix (ECM) in wound healing: a review. *Biomimetics* 2022;7:87. DOI PubMed PMC
 112. Rodriguez-Pascual F, Rosell-Garcia T. Lysyl oxidases: functions and disorders. *J Glaucoma* 2018;27 Suppl 1:S15-9. DOI PubMed
 113. Wang LC, Lo A, Scholler J, et al. Targeting fibroblast activation protein in tumor stroma with chimeric antigen receptor T cells can inhibit tumor growth and augment host immunity without severe toxicity. *Cancer Immunol Res* 2014;2:154-66. DOI PubMed PMC
 114. Nissen NI, Karsdal M, Willumsen N. Collagens and cancer associated fibroblasts in the reactive stroma and its relation to cancer biology. *J Exp Clin Cancer Res* 2019;38:115. DOI PubMed PMC
 115. Henke E, Nandigama R, Ergün S. Extracellular matrix in the tumor microenvironment and its impact on cancer therapy. *Front Mol Biosci* 2019;6:160. DOI PubMed PMC
 116. Hughes CC. Endothelial-stromal interactions in angiogenesis. *Curr Opin Hematol* 2008;15:204-9. DOI PubMed PMC
 117. Nishida N, Yano H, Nishida T, Kamura T, Kojiro M. Angiogenesis in cancer. *Vasc Health Risk Manag* 2006;2:213-9. DOI PubMed PMC
 118. Li Y, Zhao L, Li XF. Hypoxia and the tumor microenvironment. *Technol Cancer Res Treat* 2021;20:15330338211036304. DOI PubMed PMC
 119. Zhang Y, Coleman M, Brekken RA. Perspectives on hypoxia signaling in tumor stroma. *Cancers* 2021;13:3070. DOI PubMed PMC
 120. Crola Da Silva C, Lamerant-Fayel N, Paprocka M, et al. Selective human endothelial cell activation by chemokines as a guide to cell homing. *Immunology* 2009;126:394-404. DOI PubMed PMC
 121. Crowley T, Buckley CD, Clark AR. Stroma: the forgotten cells of innate immune memory. *Clin Exp Immunol* 2018;193:24-36. DOI PubMed PMC
 122. Jiang Z, Zhou J, Li L, et al. Pericytes in the tumor microenvironment. *Cancer Lett* 2023;556:216074. DOI
 123. Thomas H, Cowin AJ, Mills SJ. The importance of pericytes in healing: wounds and other pathologies. *Int J Mol Sci* 2017;18:1129. DOI PubMed PMC
 124. Paiva AE, Lousado L, Guerra DAP, et al. Pericytes in the premetastatic niche. *Cancer Res* 2018;78:2779-86. DOI PubMed PMC
 125. Kirk T, Ahmed A, Rognoni E. Fibroblast memory in development, homeostasis and disease. *Cells* 2021;10:2840. DOI PubMed PMC

126. Netea MG, Quintin J, van der Meer JW. Trained immunity: a memory for innate host defense. *Cell Host Microbe* 2011;9:355-61. DOI PubMed
127. Wang T, Hu Y, Dusi S, et al. "Open Sesame" to the complexity of pattern recognition receptors of myeloid-derived suppressor cells in cancer. *Front Immunol* 2023;14:1130060. DOI PubMed PMC
128. Akira S, Uematsu S, Takeuchi O. Pathogen recognition and innate immunity. *Cell* 2006;124:783-801. DOI PubMed
129. Netea MG, van der Meer JW. Trained immunity: an ancient way of remembering. *Cell Host Microbe* 2017;21:297-300. DOI PubMed
130. Dakin SG, Buckley CD, Al-Mossawi MH, et al. Persistent stromal fibroblast activation is present in chronic tendinopathy. *Arthritis Res Ther* 2017;19:16. DOI PubMed PMC
131. Klein K, Frank-Bertoncelj M, Karouzakis E, et al. The epigenetic architecture at gene promoters determines cell type-specific LPS tolerance. *J Autoimmun* 2017;83:122-33. DOI
132. Sohn C, Lee A, Qiao Y, Loupasakis K, Ivashkiv LB, Kalliolias GD. Prolonged tumor necrosis factor α primes fibroblast-like synoviocytes in a gene-specific manner by altering chromatin. *Arthritis Rheumatol* 2015;67:86-95. DOI PubMed PMC
133. Koch SR, Lamb FS, Hellman J, Sherwood ER, Stark RJ. Potentiation and tolerance of toll-like receptor priming in human endothelial cells. *Transl Res* 2017;180:53-67.e4. DOI PubMed PMC
134. Naik S, Larsen SB, Gomez NC, et al. Author correction: inflammatory memory sensitizes skin epithelial stem cells to tissue damage. *Nature* 2018;560:E2. DOI
135. Ara T, Kurata K, Hirai K, et al. Human gingival fibroblasts are critical in sustaining inflammation in periodontal disease. *J Periodontol Res* 2009;44:21-7. DOI
136. Lee A, Qiao Y, Grigoriev G, et al. Tumor necrosis factor α induces sustained signaling and a prolonged and unremitting inflammatory response in rheumatoid arthritis synovial fibroblasts. *Arthritis Rheum* 2013;65:928-38. DOI PubMed PMC
137. Springer TA. Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm. *Cell* 1994;76:301-14. DOI PubMed
138. Wolff B, Burns AR, Middleton J, Rot A. Endothelial cell "memory" of inflammatory stimulation: human venular endothelial cells store interleukin 8 in Weibel-Palade bodies. *J Exp Med* 1998;188:1757-62. DOI PubMed PMC
139. Bonfanti R, Furie BC, Furie B, Wagner DD. PADGEM (GMP140) is a component of Weibel-Palade bodies of human endothelial cells. *Blood* 1989;73:1109-12. DOI PubMed
140. McEver RP, Beckstead JH, Moore KL, Marshall-Carlson L, Bainton DF. GMP-140, a platelet alpha-granule membrane protein, is also synthesized by vascular endothelial cells and is localized in Weibel-Palade bodies. *J Clin Invest* 1989;84:92-9. DOI PubMed PMC
141. Zimmerman GA, McIntyre TM, Mehra M, Prescott SM. Endothelial cell-associated platelet-activating factor: a novel mechanism for signaling intercellular adhesion. *J Cell Biol* 1990;110:529-40. DOI PubMed PMC
142. Séguin C, Abid MR, Spokes KC, et al. Priming effect of homocysteine on inducible vascular cell adhesion molecule-1 expression in endothelial cells. *Biomed Pharmacother* 2008;62:395-400. DOI PubMed PMC
143. Ceriello A, Ihnat MA, Thorpe JE. Clinical review 2: the "metabolic memory": is more than just tight glucose control necessary to prevent diabetic complications? *J Clin Endocrinol Metab* 2009;94:410-5. DOI PubMed
144. Testa R, Bonfigli AR, Prattichizzo F, La Sala L, De Nigris V, Ceriello A. The "Metabolic Memory" theory and the early treatment of hyperglycemia in prevention of diabetic complications. *Nutrients* 2017;9:437. DOI PubMed PMC
145. Yao Y, Song Q, Hu C, et al. Endothelial cell metabolic memory causes cardiovascular dysfunction in diabetes. *Cardiovasc Res* 2022;118:196-211. DOI
146. Muhl L, Genové G, Leptidis S, et al. Single-cell analysis uncovers fibroblast heterogeneity and criteria for fibroblast and mural cell identification and discrimination. *Nat Commun* 2020;11:3953. DOI PubMed PMC
147. Amersfoort J, Eelen G, Carmeliet P. Immunomodulation by endothelial cells - partnering up with the immune system? *Nat Rev Immunol* 2022;22:576-88. DOI PubMed PMC
148. Yeo SY, Lee KW, Shin D, An S, Cho KH, Kim SH. A positive feedback loop bi-stably activates fibroblasts. *Nat Commun* 2018;9:3016. DOI PubMed PMC
149. Wynn TA. Cellular and molecular mechanisms of fibrosis. *J Pathol* 2008;214:199-210. DOI PubMed PMC
150. Rosenberg SA. Raising the bar: the curative potential of human cancer immunotherapy. *Sci Transl Med* 2012;4:127ps8. DOI PubMed PMC
151. Lin R, Zhang C, Zheng J, et al. Chronic inflammation-associated genomic instability paves the way for human esophageal carcinogenesis. *Oncotarget* 2016;7:24564-71. DOI PubMed PMC
152. Hibino S, Kawazoe T, Kasahara H, et al. Inflammation-induced tumorigenesis and metastasis. *Int J Mol Sci* 2021;22:5421. DOI PubMed PMC
153. Bockerstett KA, DiPaolo RJ. Regulation of gastric carcinogenesis by inflammatory cytokines. *Cell Mol Gastroenter Hepatol* 2017;4:47-53. DOI PubMed PMC
154. Qian S, Golubnitschaja O, Zhan X. Chronic inflammation: key player and biomarker-set to predict and prevent cancer development and progression based on individualized patient profiles. *EPMA J* 2019;10:365-81. DOI PubMed PMC
155. He S, Xu J, Liu X, Zhen Y. Advances and challenges in the treatment of esophageal cancer. *Acta Pharm Sin B* 2021;11:3379-92. DOI PubMed PMC

156. Ashrafi A, Akter Z, Modareszadeh P, et al. Current landscape of therapeutic resistance in lung cancer and promising strategies to overcome resistance. *Cancers* 2022;14:4562. DOI PubMed PMC
157. Marin JJ, Al-Abdulla R, Lozano E, et al. Mechanisms of resistance to chemotherapy in gastric cancer. *Anti Agent Med Chem* 2016;16:318-34. DOI
158. Wang Q, Shen X, Chen G, Du J. Drug resistance in colorectal cancer: from mechanism to clinic. *Cancers* 2022;14:2928. DOI PubMed PMC
159. Scott AK, Rafuse M, Neu CP. Mechanically induced alterations in chromatin architecture guide the balance between cell plasticity and mechanical memory. *Front Cell Dev Biol* 2023;11:1084759. DOI PubMed PMC
160. Burnet FM. The Concept of immunological surveillance. *Prog Exp Tumor Res* 1970;13:1-27. DOI
161. Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoediting: from immunosurveillance to tumor escape. *Nat Immunol* 2002;3:991-8. DOI PubMed
162. Penn I. Posttransplant malignancies. *Transplant Proc* 1999;31:1260-2. DOI PubMed
163. Sheil AG. Cancer after transplantation. *World J Surg* 1986;10:389-96. DOI PubMed
164. Penn I. Malignant melanoma in organ allograft recipients. *Transplantation* 1996;61:274-8. DOI PubMed
165. Penn I. Sarcomas in organ allograft recipients. *Transplantation* 1995;60:1485-91. DOI PubMed
166. Pham SM, Kormos RL, Landreneau RJ, et al. Solid tumors after heart transplantation: lethality of lung cancer. *Ann Thorac Surg* 1995;60:1623-6. DOI
167. Nair SS, Weil R, Dovey Z, Davis A, Tewari AK. The tumor microenvironment and immunotherapy in prostate and bladder cancer. *Urol Clin North Am* 2020;47:e17-54. DOI PubMed
168. Zhong C, Li Y, Yang J, et al. Immunotherapy for hepatocellular carcinoma: current limits and prospects. *Front Oncol* 2021;11:589680. DOI PubMed PMC
169. Jia D, Zhou Z, Kwon OJ, et al. Stromal FOXF2 suppresses prostate cancer progression and metastasis by enhancing antitumor immunity. *Nat Commun* 2022;13:6828. DOI PubMed PMC
170. Lander VE, Belle JI, Kingston NL, et al. Stromal reprogramming by FAK inhibition overcomes radiation resistance to allow for immune priming and response to checkpoint blockade. *Cancer Discov* 2022;12:2774-99. DOI
171. Yang D, Duan MH, Yuan QE, et al. Suppressive stroma-immune prognostic signature impedes immunotherapy in ovarian cancer and can be reversed by PDGFRB inhibitors. *J Transl Med* 2023;21:586. DOI PubMed PMC
172. Bremnes RM, Dønnem T, Al-Saad S, et al. The role of tumor stroma in cancer progression and prognosis: emphasis on carcinoma-associated fibroblasts and non-small cell lung cancer. *J Thorac Oncol* 2011;6:209-17. DOI
173. Nishikawa H, Koyama S. Mechanisms of regulatory T cell infiltration in tumors: implications for innovative immune precision therapies. *J Immunother Cancer* 2021;9:e002591. DOI PubMed PMC
174. Lunardi S, Jamieson NB, Lim SY, et al. IP-10/CXCL10 induction in human pancreatic cancer stroma influences lymphocytes recruitment and correlates with poor survival. *Oncotarget* 2014;5:11064-80. DOI PubMed PMC
175. Vilgelm AE, Richmond A. Chemokines modulate immune surveillance in tumorigenesis, metastasis, and response to immunotherapy. *Front Immunol* 2019;10:333. DOI PubMed PMC
176. Hussain S, Peng B, Cherian M, Song JW, Ahirwar DK, Ganju RK. The roles of stroma-derived chemokine in different stages of cancer metastases. *Front Immunol* 2020;11:598532. DOI PubMed PMC
177. Deng J, Jiang R, Meng E, Wu H. CXCL5: a coachman to drive cancer progression. *Front Oncol* 2022;12:944494. DOI PubMed PMC
178. Kraman M, Bambrough PJ, Arnold JN, et al. Suppression of antitumor immunity by stromal cells expressing fibroblast activation protein- α . *Science* 2010;330:827-30. DOI
179. Poznansky MC, Olszak IT, Foxall R, Evans RH, Luster AD, Scadden DT. Active movement of T cells away from a chemokine. *Nat Med* 2000;6:543-8. DOI PubMed
180. Kryczek I, Wei S, Keller E, Liu R, Zou W. Stroma-derived factor (SDF-1/CXCL12) and human tumor pathogenesis. *Am J Physiol Cell Physiol* 2007;292:C987-95. DOI PubMed
181. Schreiber H, Rowley DA. Cancer. Awakening immunity. *Science* 2010;330:761-2. DOI PubMed
182. Yang L, Pang Y, Moses HL. TGF- β and immune cells: an important regulatory axis in the tumor microenvironment and progression. *Trends Immunol* 2010;31:220-7. DOI PubMed PMC
183. Shi X, Young CD, Zhou H, Wang X. Transforming growth factor- β signaling in fibrotic diseases and cancer-associated fibroblasts. *Biomolecules* 2020;10:1666. DOI PubMed PMC
184. Mariathasan S, Turley SJ, Nickles D, et al. TGF β attenuates tumour response to PD-L1 blockade by contributing to exclusion of T cells. *Nature* 2018;554:544-8. DOI
185. Zippi M, De Toma G, Minervini G, et al. Desmoplasia influenced recurrence of disease and mortality in stage III colorectal cancer within five years after surgery and adjuvant therapy. *Saudi J Gastroenterol* 2017;23:39-44. DOI PubMed PMC
186. Whatcott CJ, Posner RG, Von Hoff DD, Han H. Chapter 8 desmoplasia and chemoresistance in pancreatic cancer. In: Grippo PJ, Munshi HG, editors. Pancreatic cancer and tumor microenvironment. India: Trivandrum; 2012. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK98939/> [Last accessed on 22 Jul 2024].
187. DeClerck YA. Desmoplasia: a response or a niche? *Cancer Discov* 2012;2:772-4. DOI

188. González-González L, Alonso J. Periostin: a matricellular protein with multiple functions in cancer development and progression. *Front Oncol* 2018;8:225. [DOI](#) [PubMed](#) [PMC](#)
189. Wei T, Wang K, Liu S, et al. Periostin deficiency reduces PD-1⁺ tumor-associated macrophage infiltration and enhances anti-PD-1 efficacy in colorectal cancer. *Cell Rep* 2023;42:112090. [DOI](#)
190. Mortezaee K. Immune escape: a critical hallmark in solid tumors. *Life Sci* 2020;258:118110. [DOI](#) [PubMed](#)
191. Galon J, Bruni D. Approaches to treat immune hot, altered and cold tumours with combination immunotherapies. *Nat Rev Drug Discov* 2019;18:197-218. [DOI](#) [PubMed](#)
192. Hussein MR, Al-Assiri M, Musalam AO. Phenotypic characterization of the infiltrating immune cells in normal prostate, benign nodular prostatic hyperplasia and prostatic adenocarcinoma. *Exp Mol Pathol* 2009;86:108-13. [DOI](#) [PubMed](#)
193. von Amsberg G, Alsdorf W, Karagiannis P, et al. Immunotherapy in advanced prostate cancer-light at the end of the tunnel? *Int J Mol Sci* 2022;23:2569. [DOI](#) [PubMed](#) [PMC](#)
194. Anker JF, Naseem AF, Mok H, Schaeffer AJ, Abdulkadir SA, Thumbikat P. Multi-faceted immunomodulatory and tissue-tropic clinical bacterial isolate potentiates prostate cancer immunotherapy. *Nat Commun* 2018;9:1591. [DOI](#) [PubMed](#) [PMC](#)
195. Vareki S. High and low mutational burden tumors versus immunologically hot and cold tumors and response to immune checkpoint inhibitors. *J Immunother Cancer* 2018;6:157. [DOI](#) [PubMed](#) [PMC](#)
196. Wang I, Song L, Wang BY, Rezazadeh Kalebasty A, Uchio E, Zi X. Prostate cancer immunotherapy: a review of recent advancements with novel treatment methods and efficacy. *Am J Clin Exp Urol* 2022;10:210-33. [PubMed](#) [PMC](#)
197. Ma Z, Zhang W, Dong B, et al. Docetaxel remodels prostate cancer immune microenvironment and enhances checkpoint inhibitor-based immunotherapy. *Theranostics* 2022;12:4965-79. [DOI](#) [PubMed](#) [PMC](#)
198. Chesner L, Graff J, Polesso F, et al. Abstract B041: AR suppresses MHC class I expression and T-cell response in prostate cancer. *Cancer Res* 2023;83:B041. [DOI](#)
199. Kogan-Sakin I, Cohen M, Paland N, et al. Prostate stromal cells produce CXCL-1, CXCL-2, CXCL-3 and IL-8 in response to epithelia-secreted IL-1. *Carcinogenesis* 2009;30:698-705. [DOI](#)
200. Tse BW, Scott KF, Russell PJ. Paradoxical roles of tumour necrosis factor-alpha in prostate cancer biology. *Prostate Cancer* 2012;2012:128965. [DOI](#) [PubMed](#) [PMC](#)
201. Smith BN, Mishra R, Billet S, et al. Antagonizing CD105 and androgen receptor to target stromal-epithelial interactions for clinical benefit. *Mol Ther* 2023;31:78-89. [DOI](#) [PubMed](#) [PMC](#)
202. Zhou C, Gao Y, Ding P, Wu T, Ji G. The role of CXCL family members in different diseases. *Cell Death Discov* 2023;9:212. [DOI](#) [PubMed](#) [PMC](#)
203. Bullock K, Richmond A. Suppressing MDSC recruitment to the tumor microenvironment by antagonizing CXCR2 to enhance the efficacy of immunotherapy. *Cancers* 2021;13:6293. [DOI](#) [PubMed](#) [PMC](#)
204. Korbecki J, Kupnicka P, Chlubek M, Gorący J, Gutowska I, Baranowska-Bosiacka I. CXCR2 receptor: regulation of expression, signal transduction, and involvement in cancer. *Int J Mol Sci* 2022;23:2168. [DOI](#) [PubMed](#) [PMC](#)
205. Di Mitri D, Mirinda M, Vasilevska J, et al. Re-education of tumor-associated macrophages by CXCR2 blockade drives senescence and tumor inhibition in advanced prostate cancer. *Cell Rep* 2019;28:2156-2168.e5. [DOI](#) [PubMed](#) [PMC](#)
206. Bahig H, Taussky D, Delouya G, et al. Neutrophil count is associated with survival in localized prostate cancer. *BMC Cancer* 2015;15:594. [DOI](#) [PubMed](#) [PMC](#)
207. Sharma J, Gray KP, Harshman LC, et al. Elevated IL-8, TNF- α , and MCP-1 in men with metastatic prostate cancer starting androgen-deprivation therapy (ADT) are associated with shorter time to castration-resistance and overall survival. *Prostate* 2014;74:820-8. [DOI](#)
208. Minas TZ, Candia J, Dorsey TH, et al. Serum proteomics links suppression of tumor immunity to ancestry and lethal prostate cancer. *Nat Commun* 2022;13:1759. [DOI](#) [PubMed](#) [PMC](#)
209. Wallace TA, Prueitt RL, Yi M, et al. Tumor immunobiological differences in prostate cancer between African-American and European-American men. *Cancer Res* 2008;68:927-36. [DOI](#)
210. Zhu W, Wu J, Huang J, et al. Multi-omics analysis reveals a macrophage-related marker gene signature for prognostic prediction, immune landscape, genomic heterogeneity, and drug choices in prostate cancer. *Front Immunol* 2023;14:1122670. [DOI](#) [PubMed](#) [PMC](#)
211. Chen C, Luo J, Wang X. Identification of prostate cancer subtypes based on immune signature scores in bulk and single-cell transcriptomes. *Med Oncol* 2022;39:123. [DOI](#) [PubMed](#)
212. Guo T, Wang J, Yan S, et al. A combined signature of glycolysis and immune landscape predicts prognosis and therapeutic response in prostate cancer. *Front Endocrinol* 2022;13:1037099. [DOI](#) [PubMed](#) [PMC](#)
213. Ren C, Wang Q, Wang S, et al. Metabolic syndrome-related prognostic index: predicting biochemical recurrence and differentiating between cold and hot tumors in prostate cancer. *Front Endocrinol* 2023;14:1148117. [DOI](#) [PubMed](#) [PMC](#)
214. Li N, Yu K, Lin Z, Zeng D. Development of a novel immune subtyping system expanded with immune landscape and an 11-gene signature for predicting prostate cancer survival. *J Oncol* 2022;2022:1183173. [DOI](#) [PubMed](#) [PMC](#)
215. Keam SP, Halse H, Nguyen T, et al. High dose-rate brachytherapy of localized prostate cancer converts tumors from cold to hot. *J Immunother Cancer* 2020;8:e000792. [DOI](#) [PubMed](#) [PMC](#)
216. Shen Y, Xu H, Long M, et al. Screening to identify an immune landscape-based prognostic predictor and therapeutic target for prostate cancer. *Front Oncol* 2021;11:761643. [DOI](#) [PubMed](#) [PMC](#)

217. Zhang G, Luo Y. An immune-related lncRNA signature to predict the biochemical recurrence and immune landscape in prostate cancer. *Int J Gen Med* 2021;14:9031-49. [DOI](#) [PubMed](#) [PMC](#)
218. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011;144:646-74. [DOI](#) [PubMed](#)
219. Antognelli C, Mandarano M, Prosperi E, Sidoni A, Talesa VN. Glyoxalase-1-dependent methylglyoxal depletion sustains PD-L1 expression in metastatic prostate cancer cells: a novel mechanism in cancer immunosurveillance escape and a potential novel target to overcome PD-L1 blockade resistance. *Cancers* 2021;13:2965. [DOI](#) [PubMed](#) [PMC](#)
220. Datta M, Coussens LM, Nishikawa H, Hodi FS, Jain RK. Reprogramming the tumor microenvironment to improve immunotherapy: emerging strategies and combination therapies. *Am Soc Clin Oncol Educ Book* 2019;39:165-74. [DOI](#) [PubMed](#) [PMC](#)
221. Perera MPJ, Thomas PB, Risbridger GP, et al. Chimeric antigen receptor T-cell therapy in metastatic castrate-resistant prostate cancer. *Cancers* 2022;14:503. [DOI](#) [PubMed](#) [PMC](#)
222. Bander NH, Nanus DM, Milowsky MI, Kostakoglu L, Vallabahajosula S, Goldsmith SJ. Targeted systemic therapy of prostate cancer with a monoclonal antibody to prostate-specific membrane antigen. *Semin Oncol* 2003;30:667-76. [DOI](#) [PubMed](#)
223. Sardinha M, Palma Dos Reis AF, Barreira JV, Fontes Sousa M, Pacey S, Luz R. Antibody-drug conjugates in prostate cancer: a systematic review. *Cureus* 2023;15:e34490. [DOI](#) [PubMed](#) [PMC](#)
224. Kamat NV, Yu EY, Lee JK. BiTE-ing into prostate cancer with bispecific T-cell engagers. *Clin Cancer Res* 2021;27:2675-7. [DOI](#) [PubMed](#) [PMC](#)
225. Yang K, Feng S, Luo Z. Oncolytic adenovirus, a new treatment strategy for prostate cancer. *Biomedicines* 2022;10:3262. [DOI](#) [PubMed](#) [PMC](#)
226. Pieczonka CM, Telonis D, Mouraviev V, Albala D. Sipuleucel-T for the treatment of patients with metastatic castrate-resistant prostate cancer: considerations for clinical practice. *Rev Urol* 2015;17:203-10. [PubMed](#) [PMC](#)
227. Kantoff PW, Higano CS, Shore ND, et al. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *N Engl J Med* 2010;363:411-22. [DOI](#)
228. Donninger H, Li C, Eaton JW, Yaddanapudi K. Cancer vaccines: promising therapeutics or an unattainable dream. *Vaccines* 2021;9:668. [DOI](#) [PubMed](#) [PMC](#)
229. Chen S, Wainwright DA, Wu JD, et al. CD73: an emerging checkpoint for cancer immunotherapy. *Immunotherapy* 2019;11:983-97. [DOI](#) [PubMed](#) [PMC](#)
230. Mbongue JC, Nicholas DA, Torrez TW, Kim NS, Firek AF, Langridge WH. The role of indoleamine 2, 3-dioxygenase in immune suppression and autoimmunity. *Vaccines* 2015;3:703-29. [DOI](#) [PubMed](#) [PMC](#)
231. Solinas C, Migliori E, De Silva P, Willard-Gallo K. LAG3: the biological processes that motivate targeting this immune checkpoint molecule in human cancer. *Cancers* 2019;11:1213. [DOI](#) [PubMed](#) [PMC](#)
232. Miller RA, Luke JJ, Hu S, et al. Anti-CD73 antibody activates human B cells, enhances humoral responses and induces redistribution of B cells in patients with cancer. *J Immunother Cancer* 2022;10:e005802. [DOI](#) [PubMed](#) [PMC](#)
233. Fan X, Quezada SA, Sepulveda MA, Sharma P, Allison JP. Engagement of the ICOS pathway markedly enhances efficacy of CTLA-4 blockade in cancer immunotherapy. *J Exp Med* 2014;211:715-25. [DOI](#) [PubMed](#) [PMC](#)
234. Fu Y, Lin Q, Zhang Z, Zhang L. Therapeutic strategies for the costimulatory molecule OX40 in T-cell-mediated immunity. *Acta Pharm Sin B* 2020;10:414-33. [DOI](#) [PubMed](#) [PMC](#)
235. Ye L, Jia K, Wang L, et al. CD137, an attractive candidate for the immunotherapy of lung cancer. *Cancer Sci* 2020;111:1461-7. [DOI](#) [PubMed](#) [PMC](#)
236. Bayer AL, Pugliese A, Malek TR. The IL-2/IL-2R system: from basic science to therapeutic applications to enhance immune regulation. *Immunol Res* 2013;57:197-209. [DOI](#) [PubMed](#) [PMC](#)
237. Schepisi G, Cursano MC, Casadei C, et al. CAR-T cell therapy: a potential new strategy against prostate cancer. *J Immunother Cancer* 2019;7:258. [DOI](#) [PubMed](#) [PMC](#)
238. Patel S, Burga RA, Powell AB, et al. Beyond CAR T cells: other cell-based immunotherapeutic strategies against cancer. *front oncol* 2019;9:196. [DOI](#) [PubMed](#) [PMC](#)
239. Mills JK, Henderson MA, Giuffrida L, et al. Generating CAR T cells from tumor-infiltrating lymphocytes. *Ther Adv Vaccines Immunother* 2021;9:25151355211017119. [DOI](#) [PubMed](#) [PMC](#)
240. Zhou B, Lin W, Long Y, et al. Notch signaling pathway: architecture, disease, and therapeutics. *Signal Transduct Target Ther* 2022;7:95. [DOI](#) [PubMed](#) [PMC](#)
241. Day KC, Lorenzatti Hiles G, Kozminsky M, et al. HER2 and EGFR overexpression support metastatic progression of prostate cancer to bone. *Cancer Res* 2017;77:74-85. [DOI](#) [PubMed](#) [PMC](#)
242. Hsu EC, Rice MA, Bermudez A, et al. Trop2 is a driver of metastatic prostate cancer with neuroendocrine phenotype via PARP1. *Proc Natl Acad Sci USA* 2020;117:2032-42. [DOI](#) [PubMed](#) [PMC](#)
243. Rosellini M, Santoni M, Mollica V, et al. Treating prostate cancer by antibody-drug conjugates. *Int J Mol Sci* 2021;22:1551. [DOI](#) [PubMed](#) [PMC](#)
244. Tian Z, Liu M, Zhang Y, Wang X. Bispecific T cell engagers: an emerging therapy for management of hematologic malignancies. *J Hematol Oncol* 2021;14:75. [DOI](#) [PubMed](#) [PMC](#)
245. Lin D, Shen Y, Liang T. Oncolytic virotherapy: basic principles, recent advances and future directions. *Signal Transduct Target Ther* 2023;8:156. [DOI](#) [PubMed](#) [PMC](#)
246. Lee J, Fassnacht M, Nair S, Boczkowski D, Gilboa E. Tumor immunotherapy targeting fibroblast activation protein, a product

- expressed in tumor-associated fibroblasts. *Cancer Res* 2005;65:11156-63. DOI PubMed
247. Wen Y, Wang CT, Ma TT, et al. Immunotherapy targeting fibroblast activation protein inhibits tumor growth and increases survival in a murine colon cancer model. *Cancer Sci* 2010;101:2325-32. DOI PubMed PMC
248. Yu F, Wang X, Guo ZS, Bartlett DL, Gottschalk SM, Song XT. T-cell engager-armed oncolytic vaccinia virus significantly enhances antitumor therapy. *Mol Ther* 2014;22:102-11. DOI PubMed PMC
249. Freedman JD, Duffy MR, Lei-Rossmann J, et al. An oncolytic virus expressing a T-cell engager simultaneously targets cancer and immunosuppressive stromal cells. *Cancer Res* 2018;78:6852-65. DOI
250. Chaudary N, Pintilie M, Jelveh S, Lindsay P, Hill RP, Milosevic M. Plerixafor improves primary tumor response and reduces metastases in cervical cancer treated with radio-chemotherapy. *Clin Cancer Res* 2017;23:1242-9. DOI PubMed
251. Kim ST, Hong JY, Park SH, et al. First-in-human phase I trial of anti-hepatocyte growth factor antibody (YYB101) in refractory solid tumor patients. *Ther Adv Med Oncol* 2020;12:1758835920926796. DOI PubMed PMC
252. Zhang X, Luo H. Effects of thalidomide on growth and VEGF-A expression in SW480 colon cancer cells. *Oncol Lett* 2018;15:3313-20. DOI PubMed PMC
253. Pulte ED, Dmytrijuk A, Nie L, et al. FDA approval summary: lenalidomide as maintenance therapy after autologous stem cell transplant in newly diagnosed multiple myeloma. *Oncologist* 2018;23:734-9. DOI PubMed PMC
254. Summers J, Cohen MH, Keegan P, Pazdur R. FDA drug approval summary: bevacizumab plus interferon for advanced renal cell carcinoma. *Oncologist* 2010;15:104-11. DOI PubMed PMC
255. Singh AD, Parmar S. Ramucirumab (Cyramza): a breakthrough treatment for gastric cancer. *P T* 2015;40:430-68. PubMed PMC
256. Chiorean EG, Sweeney C, Youssoufian H, et al. A phase I study of olaratumab, an anti-platelet-derived growth factor receptor alpha (PDGFR α) monoclonal antibody, in patients with advanced solid tumors. *Cancer Chemother Pharmacol* 2014;73:595-604. DOI
257. Kwon MJ. Matrix metalloproteinases as therapeutic targets in breast cancer. *Front Oncol* 2022;12:1108695. DOI PubMed PMC
258. Nadal L, Corbellari R, Villa A, et al. Novel human monoclonal antibodies specific to the alternatively spliced domain D of Tenascin C efficiently target tumors in vivo. *MAbs* 2020;12:1836713. DOI PubMed PMC
259. Neesse A, Frese KK, Bapiro TE, et al. CTGF antagonism with mAb FG-3019 enhances chemotherapy response without increasing drug delivery in murine ductal pancreas cancer. *Proc Natl Acad Sci USA* 2013;110:12325-30. DOI PubMed PMC
260. Ke FY, Chen WY, Lin MC, Hwang YC, Kuo KT, Wu HC. Novel monoclonal antibody against integrin $\alpha 3$ shows therapeutic potential for ovarian cancer. *Cancer Sci* 2020;111:3478-92. DOI PubMed PMC
261. Sun Y, Zhao C, Ye Y, et al. High expression of fibronectin 1 indicates poor prognosis in gastric cancer. *Oncol Lett* 2020;19:93-102. DOI PubMed PMC
262. StreeL G, Lucas S. Targeting immunosuppression by TGF- $\beta 1$ for cancer immunotherapy. *Biochem Pharmacol* 2021;192:114697. DOI PubMed PMC
263. Ma C, Xi S, Sun H, Zhang M, Pei Y. Identifying the oncogenic roles of FAP in human cancers based on systematic analysis. *Aging* 2023;15:7056-83. DOI PubMed PMC
264. Martori C, Sanchez-Moral L, Paul T, et al. Macrophages as a therapeutic target in metastatic prostate cancer: a way to overcome immunotherapy resistance? *Cancers* 2022;14:440. DOI PubMed PMC
265. Gordon S, Plüddemann A. The mononuclear phagocytic system. generation of diversity. *Front Immunol* 2019;10:1893. DOI PubMed PMC
266. Martinez FO, Gordon S. The M1 and M2 paradigm of macrophage activation: time for reassessment. *F1000Prime Rep* 2014;6:13. DOI PubMed PMC
267. Das A, Sinha M, Datta S, et al. Monocyte and macrophage plasticity in tissue repair and regeneration. *Am J Pathol* 2015;185:2596-606. DOI PubMed PMC
268. Gordon S, Plüddemann A. Tissue macrophages: heterogeneity and functions. *BMC Biol* 2017;15:53. DOI PubMed PMC
269. He MX, Cuoco MS, Crowdis J, et al. Transcriptional mediators of treatment resistance in lethal prostate cancer. *Nat Med* 2021;27:426-33. DOI PubMed PMC
270. Stout RD, Jiang C, Matta B, Tietzel I, Watkins SK, Suttles J. Macrophages sequentially change their functional phenotype in response to changes in microenvironmental influences. *J Immunol* 2005;175:342-9. DOI PubMed
271. Li XF, Selli C, Zhou HL, et al. Macrophages promote anti-androgen resistance in prostate cancer bone disease. *J Exp Med* 2023;220. DOI PubMed PMC
272. Mantovani A, Allavena P, Marchesi F, Garlanda C. Macrophages as tools and targets in cancer therapy. *Nat Rev Drug Discov* 2022;21:799-820. DOI PubMed PMC
273. Abram CL, Lowell CA. The diverse functions of Src family kinases in macrophages. *Front Biosci* 2008;13:4426-50. DOI PubMed PMC
274. Loi M, Salvatore G, Sottili M, et al. Tumor-associated macrophages (TAMs) modulate response to HER2-targeted agents in a humanized mouse model of breast cancer. *Clin Transl Oncol* 2022;24:1395-402. DOI
275. Watanabe H, Ohashi K, Nishii K, et al. A long-term response to nivolumab in a case of PD-L1-negative lung adenocarcinoma with an EGFR mutation and surrounding PD-L1-positive tumor-associated macrophages. *Intern Med* 2019;58:3033-7. DOI PubMed PMC
276. Su S, Lei A, Wang X, et al. Induced CAR-macrophages as a novel therapeutic cell type for cancer immune cell therapies. *Cells* 2022;11:1652. DOI PubMed PMC
277. Liu M, Liu J, Liang Z, et al. CAR-macrophages and CAR-T cells synergistically kill tumor cells in vitro. *Cells* 2022;11:3692. DOI PubMed PMC

278. Huber ML, Haynes L, Parker C, Iversen P. Interdisciplinary critique of sipuleucel-T as immunotherapy in castration-resistant prostate cancer. *J Natl Cancer Inst* 2012;104:273-9. [DOI](#) [PubMed](#) [PMC](#)
279. Murphy G, Tjoa B, Ragde H, Kenny G, Boynton A. Phase I clinical trial: T-cell therapy for prostate cancer using autologous dendritic cells pulsed with HLA-A0201-specific peptides from prostate-specific membrane antigen. *Prostate* 1996;29:371-80. [DOI](#) [PubMed](#)
280. Tjoa B, Erickson S, Bowes V, et al. Follow-up evaluation of prostate cancer patients infused with autologous dendritic cells pulsed with PSMA peptides. *Prostate* 1997;32:272-8. [DOI](#)
281. Jähnisch H, Füssel S, Kiessling A, et al. Dendritic cell-based immunotherapy for prostate cancer. *Clin Dev Immunol* 2010;2010:517493. [DOI](#) [PubMed](#) [PMC](#)
282. Kiessling A, Wehner R, Füssel S, Bachmann M, Wirth MP, Schmitz M. Tumor-associated antigens for specific immunotherapy of prostate cancer. *Cancers* 2012;4:193-217. [DOI](#) [PubMed](#) [PMC](#)
283. Kitamura H, Torigoe T, Asanuma H, Honma I, Sato N, Tsukamoto T. Down-regulation of HLA class I antigens in prostate cancer tissues and up-regulation by histone deacetylase inhibition. *J Urol* 2007;178:692-6. [DOI](#) [PubMed](#)
284. Lozano M, Cid J, Benitez-Ribas D, Otero MJ. Technical challenges in the manufacture of dendritic cell cancer therapies. *Eur Oncol Haematol* 2019;15:22-8. [DOI](#)
285. Kongsted P, Borch TH, Ellebaek E, et al. Dendritic cell vaccination in combination with docetaxel for patients with metastatic castration-resistant prostate cancer: a randomized phase II study. *Cytotherapy* 2017;19:500-13. [DOI](#)
286. Glabman RA, Choyke PL, Sato N. Cancer-associated fibroblasts: tumorigenicity and targeting for cancer therapy. *Cancers* 2022;14:3906. [DOI](#) [PubMed](#) [PMC](#)
287. Boudewijns S, Westdorp H, Koornstra RH, et al. Immune-related adverse events of dendritic cell vaccination correlate with immunologic and clinical outcome in stage III and IV melanoma patients. *J Immunother* 2016;39:241-8. [DOI](#) [PubMed](#) [PMC](#)
288. Agrawal A, Sridharan A, Prakash S, Agrawal H. Dendritic cells and aging: consequences for autoimmunity. *Expert Rev Clin Immunol* 2012;8:73-80. [DOI](#) [PubMed](#) [PMC](#)