

Commentary

Open Access



Understanding sarcoma drug resistance one cell at a time

Salah-Eddine Cherradi-Lamhamedi, Danh Truong, Joseph A. Ludwig

Department of Sarcoma Medical Oncology, Division of Cancer Medicine, The University of Texas, MD Anderson Cancer Center, Houston, TX 77030, USA.

Correspondence to: Dr. Joseph A. Ludwig, Department of Sarcoma Medical Oncology, Division of Cancer Medicine, The University of Texas, MD Anderson Cancer Center, 1515 Holcombe Blvd, Unit 450, Houston, TX 77030, USA.
E-mail: jaludwig@mdanderson.org

How to cite this article: Cherradi-Lamhamedi SE, Truong D, Ludwig JA. Understanding sarcoma drug resistance one cell at a time. *Cancer Drug Resist* 2022;5:90-2. <https://dx.doi.org/10.20517/cdr.2021.87>

Received: 1 Sep 2021 **First Decision:** 1 Nov 2021 **Revised:** 8 Nov 2021 **Accepted:** 2 Dec 2021 **Published:** 20 Jan 2022

Academic Editors: Godefridus J. Peters, Brian A. Van Tine **Copy Editor:** Yue-Yue Zhang **Production Editor:** Yue-Yue Zhang

As true for any cancer, intrinsic and acquired drug resistance represent perhaps the single biggest obstacle limiting our ability to cure patients with metastatic disease. Yet, tackling this challenge has been particularly difficult for sarcoma. First and foremost, sarcomas are rare and diverse, which splits a class of tumors representing less than 1 percent of all malignancies into more than fifty smaller molecularly and histologically distinct entities, each having its own etiology, prognosis, and clinical presentation. Even the most “common” sarcoma subtypes (e.g., liposarcoma, leiomyosarcoma, or gastrointestinal stromal tumors) have annual incidences in the thousands in the United States, whereas rarer subtypes like desmoplastic small round cell tumors number less than fifty per year. Due to their scarcity, it’s not uncommon for just a few cell lines, organoids, or patient-derived tumor explants to exist for each sarcoma subtype. This shortage limits their usefulness to model the wide variety of innate and adaptive mechanisms that exist in human tumors. Many times, no representative preclinical model exists. The scarcity of FDA-approved drugs aimed at sarcoma-specific targets, often fusion proteins and transcription factors, is also problematic.

Complicating the ability to understand tumor drug resistance, many sarcomas exhibit pronounced intra-tumoral heterogeneity, manifested by heterogeneous cell populations of varied lineages and states of differentiation. It is not unusual for osteosarcoma, for example, to harbor cells with varying degrees of



© The Author(s) 2022. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, sharing, adaptation, distribution and reproduction in any medium or format, for any purpose, even commercially, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.



chaotic osteogenic, adipogenic, or chondrogenic cell fates. Similarly, some liposarcomas are classified into well-differentiated and dedifferentiated variants by their predominant differentiation status but usually harbor a blend of both^[1]. Even tumors like Ewing sarcoma, which appear relatively homogeneous by light microscopy, can be subcategorized by the pathognomonic genomic translocation expressed and by the presence or absence of well-described point mutations in *STAG2*, *p53*, or *CDKN2A*^[2]. Thus, while light microscopy and careful review by an experienced sarcoma pathologist remain the cornerstone of sarcoma diagnosis, a clear trend in the research setting is to deeply interrogate each tumor and precisely catalog its cellular constituencies using a wide array of genomic, proteomic, and other “-omic” technologies. As a direct result of this unparalleled opportunity to parse out distinct groups of malignant cells, a more nuanced view of sarcoma drug resistance must consider not only the global factors linked to each sarcoma subtype but also the intratumoral features such as physical and mechanical cues (e.g., extracellular matrix, stiffness or confined adhesiveness) that allow for clonal evolution in response to the selective pressures triggered by standard chemotherapy or biologically targeted therapy.

This has been achieved in other cancer types by engaging powerful single-cell technologies - before and after drug exposure - to determine a tumor’s baseline transcriptomic state and nascent cellular changes associated with drug resistance^[3-6]. Single-cell and single-nucleus RNA-sequencing (sc/snRNA-seq) are the most common methods used currently and, when paired with freely available custom-made bioinformatic tools written in R or Python, provide the revolutionary ability to deeply interrogate intra-tumoral complexity^[7-9]. Furthermore, while still in their infancy, single-cell proteomic technologies have introduced promising technologies that enable high-resolution spatial image - omic (SIO) analyses of human primary tumors, which allows one to learn not only *what* cell types exist but also *how* they interact in 2D or 3D space^[10]. Though the clinical value of checkpoint blockade and cellular immunotherapies remains to be proven across the range of sarcoma subtypes, the advent of SIO, image mass cytometry, multi-color fluorescent microscopy, and other techniques should provide a fresh opportunity to examine whether such therapies can invoke an anti-cancer response. Complementing the trained eye of an experienced sarcoma pathologist, deep-learning and artificial intelligence, while still under-utilized, are primed to aid pathologists to make sense of the torrent of data generated from each slide and may allow them to perceive subtle cell-cell interactions that contribute to drug resistance and the feedback mechanisms that sustain them^[11].

Though classic measures of drug efficacy that rely on tumor size or patient survival will persist as important clinical study endpoints, a deeper appreciation of drug sensitivity/resistance must inevitably decipher how each patient’s tumor - and the cells within them - evade therapy. Like no time in history, the tools exist today to help cure cancer one cell at a time.

DECLARATIONS

Authors’ contributions

Contributed to the written content and review of this manuscript: Cherradi-Lamhamedi SE, Truong D, Ludwig JA

Availability of data and materials

Not applicable.

Financial support and sponsorship

None.

Conflicts of interest

All authors declared that there are no conflicts of interest.

Ethical approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Copyright

© The Author(s) 2022.

REFERENCES

1. Thway K, Jones RL, Noujaim J, Zaidi S, Miah AB, Fisher C. Dedifferentiated Liposarcoma: updates on morphology, genetics, and therapeutic strategies. *Adv Anat Pathol* 2016;23:30-40. DOI PubMed
2. Crompton BD, Stewart C, Taylor-Weiner A, et al. The genomic landscape of pediatric Ewing sarcoma. *Cancer Discov* 2014;4:1326-41. DOI PubMed
3. Navin N, Kendall J, Troge J, et al. Tumour evolution inferred by single-cell sequencing. *Nature* 2011;472:90-4. DOI PubMed PMC
4. Wang Y, Waters J, Leung ML, et al. Clonal evolution in breast cancer revealed by single nucleus genome sequencing. *Nature* 2014;512:155-60. DOI PubMed PMC
5. Alizadeh AA, Aranda V, Bardelli A, et al. Toward understanding and exploiting tumor heterogeneity. *Nat Med* 2015;21:846-53. DOI PubMed PMC
6. Method of the Year 2019: single-cell multimodal omics. *Nat Methods* 2020;17:1. DOI PubMed
7. Kim C, Gao R, Sei E, et al. Chemoresistance evolution in triple-negative breast cancer delineated by single-cell sequencing. *Cell* 2018;173:879-93.e13. DOI PubMed PMC
8. Moco3: an analysis toolkit for single-cell RNA-seq. 2020. Available from: <https://cole-trapnell-lab.github.io/monocle3/> [Last accessed on 4 Jan 2022].
9. SEURAT: an R toolkit for single-cell genomics. 2020. Available from: <https://satijalab.org/seurat/> [Last accessed on 4 Jan 2022].
10. Schapiro D, Jackson HW, Raghuraman S, et al. histoCAT: analysis of cell phenotypes and interactions in multiplex image cytometry data. *Nat Methods* 2017;14:873-6. DOI PubMed PMC
11. Weiskittel TM, Correia C, Yu GT, et al. The trifecta of single-cell, systems-biology, and machine-learning approaches. *Genes (Basel)* 2021;12:1098. DOI PubMed PMC