

Review

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The role of genetic factors in HBV-related HCC: perspectives from local genetic backgrounds and clinical epidemiology

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Abstract

Familial clustering of hepatitis B surface antigen carriers (HBsAg) and hepatocellular carcinoma (HCC) has led to the evaluation of the role of genetics in hepatitis B-related diseases. Consistent reports indicate that the HLA-DP and -DQ loci are associated with persistent hepatitis B virus (HBV) infection. However, for hepatocarcinogenesis, existing studies have low power and conflicting data. Global single nucleotide polymorphism (SNP) data was collected from the 1000 Genomes Project and correlated with local epidemiological information. Southeastern Asia has a higher prevalence of HBsAg than Northeastern Asia; this was used in the evaluation of persistent HBV infection. The higher incidence of HCC in West Africa compared with East Africa was used in the evaluation of hepatocarcinogenesis. The allele frequencies for SNPs were significantly different between East Asians and Africans. Therefore, SNPs that have been identified in persistent HBV infections in East Asia may not be completely applicable in Africa. SNPs in NTCP, CTF19, and the HLA-DQ and -DP loci showed North-to-South allele frequency changes in East Asia. These findings confirm the role of genetics in persistent HBV infection. Some of the SNPs in the HLA loci show a trend of West-to-East allele frequency changes in Africa, indicating they may participate in hepatocarcinogenesis. Among the non-HLA related SNPs, rs2596542 in MICA shows a strong trend of allele frequency changes and is correlated with HCC incidence in Africa. SNPs in KIF1, IL-1A, and STAT4 also show, albeit with low statistical power, allele frequency trends compatible with HCC incidence. Taken together, there are strong correlations between background genetics in HLA-DP and -DQ loci with persistent HBV infection and hepatocarcinogenesis. The correlations were weak-positive in non-HLA loci.



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Keywords: Genetic polymorphism, genome-wide associated studies, hepatitis B virus, hepatocellular carcinoma

INTRODUCTION

Hepatitis B is a global disease that results in an increased risk of liver cirrhosis and hepatocellular carcinoma^[1,2]. A strong familial clustering of chronic hepatitis B carriers and hepatocellular carcinomas (HCC) has been well-reported^[3-6]. This could be related to the high intrafamilial spread of hepatitis B virus (HBV) infection. Infection of HBV in the early stage of life will result in chronic persistent infection^[4]. Throughout the course of this disease, intermittent relapsing liver necroinflammation will occur. This process is a defensive response to eliminate HBV replication and/or clear the virus. In general, by 80 years of age, half of chronic hepatitis B carriers will have suppressed HBV replication and cleared hepatitis B surface antigens (HBsAg)^[7].

However, some patients may develop liver cirrhosis as a result of repeated liver inflammation and fibrogenesis^[8]. Liver necroinflammation can induce chromosomal damage^[9], and HBV is able to integrate into the human genome^[10]. Such an injury to the host genome can induce chromosomal instability and promote hepatocarcinogenesis. Genetic factors associated with HCC have been considered because of its familial tendency. Many candidate genes and genome-wide associated studies (GWAS) have revealed nearly one hundred genes to be associated with chronic persistent infection or hepatocarcinogenesis^[11-14]. Previous extensive and elegant meta-analysis reports have addressed these issues. However, not all of these HBV/HCC related genes have been confirmed and replicated by subsequent studies, demonstrating the difficulties in sorting HCC-associated genes. This is partly due to the fact that HBV-host interactions are not simply a genetic problem, as well as the fact that there are differences in the genetic backgrounds of study populations.

Therefore, in this study, we shall try to understand HBV-related single nucleotide polymorphisms (SNPs) by performing correlations between genetic backgrounds and two well-known epidemiological datasets. The genetic backgrounds will be obtained from the 1000 Genomes Project (<http://www.1000genomes.org/>)^[15]. Epidemiological concerns about a higher prevalence of HBsAg in southern compared to Northeastern Asia will be used in the evaluation for persistent HBV infection^[16]. A higher prevalence of HCC in West Africa compared to East Africa will also be used for evaluation of hepatocarcinogenesis (World Health Organization, <http://gco.iarc.fr/today>). With these viewpoints, we may obtain additional information independent of the observations in reported studies about HBV-related genetic polymorphisms.

CHRONIC PERSISTENT HBV INFECTION

Hepatocarcinogenesis in chronic HBV infection is not a purely genetic disease; it depends on host and virus interactions. Persistent HBV infection is the first stage toward hepatocarcinogenesis.

HBV clearance

When humans are exposed to HBV, either acute hepatitis with spontaneous viral clearance or chronic persistent infection may develop [Figure 1]. Both the timing and transmission route of infection are important in persistent infections. Individuals who are infected in the early stages of life and via vertical transmission are more likely to develop persistent infection^[4]. In GWAS studies, HLA-DP and -DQ have been shown to consistently be associated with persistent HBV infection in East Asians^[17-23]. However, such high-risk alleles are relatively rare in Africans^[24]. Therefore, HBV infection elicited in the early stages of life remains an important mechanism of persistent HBV infection. It is independent of genetic polymorphisms in the HLA-DP and -DQ loci.

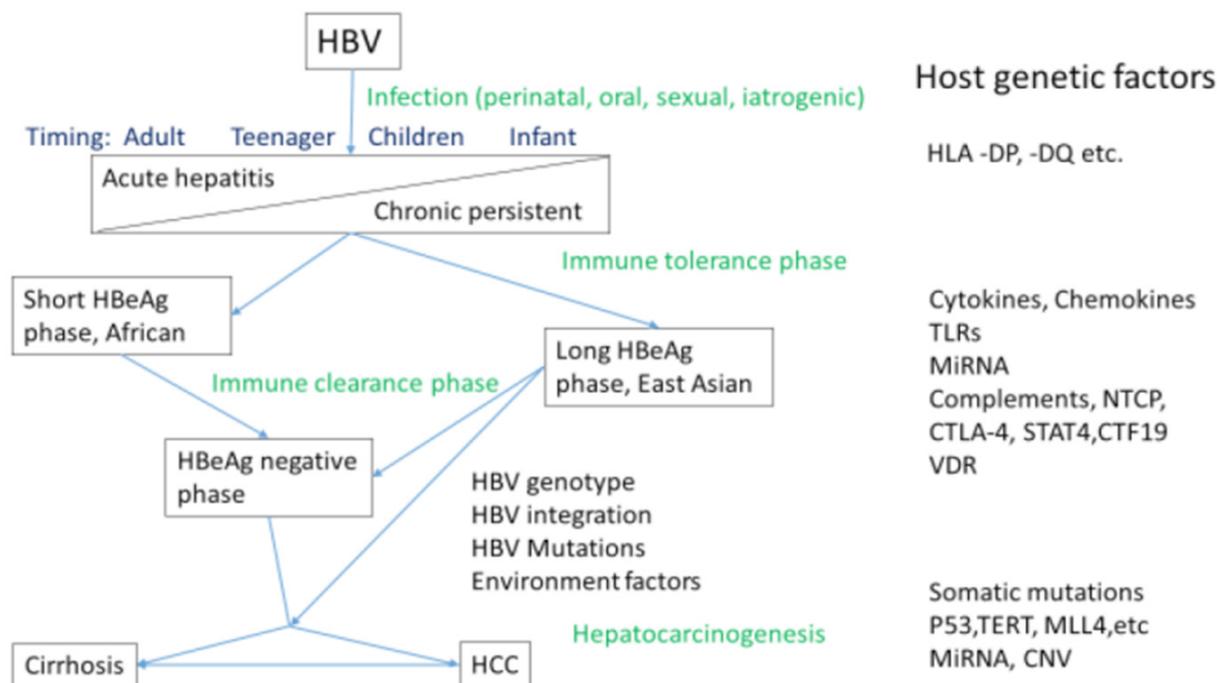


Figure 1. Summary of hepatitis B virus (HBV) and host interactions. When humans are exposed to HBV, either acute hepatitis with subsequent virus clearance or persistent infection may develop. Those who are infected in the early stages of life, and via vertical transmission, are prone to progress to persistent infection. HLA-DP and -DQ loci are associated with persistent HBV infection in East Asians. Chronic persistent HBV infection starts with the presence of hepatitis B e-antigen (HBeAg), known as the immune tolerance stage. In this phase, Africans tend to clear HBeAg before puberty while East Asians tend to clear HBeAg between the second and fourth decades of life. About 5% of HBeAg-negative patients still present with active viral replication. Many immune-related genes participate in HBV clearance. Inability to effectively clear HBV may result in liver cirrhosis and increased HBV mutations and integrations into the human genome. All of these events and some genetic polymorphisms in the host may promote hepatocarcinogenesis. HCC: hepatocellular carcinoma

Persistent HBV infection

Chronic persistent HBV infections start with the presence of the hepatitis B e-antigen (HBeAg). During the HBeAg-positive immune tolerance phase, the HBV DNA level is high with low levels of liver inflammation^[25]. Due to some unclear triggering factors, immune clearance of HBV will develop between the second and fourth decades of life in East Asians. There is a difference between Africans and East Asians regarding the duration of the HBeAg-positive period^[26-29]. A rapid clearance of HBeAg before puberty can be found in Africans but rarely occurs in East Asians^[30,31]. This difference may be associated with variations in the HLA-DP and -DQ genotypes^[24]. Certain SNPs in HLA-DP and -DQ loci may prolong the HBV replication phase in adults of East Asian descent. A high HBV DNA level in parents will increase the risk of persistent infection in their offspring (submitted for publication).

HBV genotypes C and D are associated with a more persistent liver necroinflammation^[28]. Many immune-related genes may participate in this immune clearance phase [Figure 1]. Patients either in the HBeAg-positive or -negative phase who are unable to suppress HBV replication may develop liver cirrhosis due to repeated liver inflammation and fibrogenesis.

HBV integration and host interaction

A prolonged HBV replication phase and liver cirrhosis increases the risk of HCC^[25]. The mechanism may be related to hepatitis Bx proteins^[32], increased endoplasmic reticulum stress^[9], HBV integration^[10,33] and inflammation-related chromosome damage.

Soon after HBV infection, part of the HBV genome can be integrated into the host genome. The mechanism of HBV integration is not fully understood. From the *in vitro* study done using the Na⁺-taurocholate co-transporting polypeptide (NTCP) transfected hepatoma cell line, HBV integration can be detected randomly in the host genome shortly after infection^[33]. This observation implies that HBV integration is independent of immune-related inflammation. These HBV integrations are mostly harmless but may produce genomic instability. After decades, some of the integrations may become more prominent. In the presence of additional factors, such as inflammation, HBV mutations, or environmental carcinogens, a segment of the hepatocytes carrying HBV integrations may become clonal and develop into HCC. Several integration hot sites, including TERT, KMT2B, DDX11L1, CCNE1, and CCNA2, can be found more frequently in HCC than in non-HCC tissues^[34], while FN1 is commonly found in non-tumour tissues. A prolonged HBeAg phase and high viral load carry a higher frequency of HBV integrations^[35].

HBV mutants and host interaction

The wild-type HBV genome and its proteins are not directly cytopathic. Host immune responses and inflammation are induced to clear HBV during the second to fourth decades of life in East Asians. If the HBV immune clearance process is unsuccessful and prolonged, complicated HBV mutations may develop and escape immune surveillance. Through repeat necroinflammation, several mutation hot spots in the EnhII (C1653T)/BCP(A1762T/G1764A, T1753V)/PC(G1896A) regions were found more frequently in HCC^[36]. In addition, some of the pre-S/S mutations or truncations may become directly cytopathic and/or carcinogenic^[37].

HLA SNPS IN RELATION TO HBV INFECTION AND HEPATOCARCINOGENESIS

Global allele frequency of HBV-related SNPs in HLA loci

Based on GWAS studies, HLA-DP and -DQ loci are associated with persistent HBV infection. These SNPs have been reported quite consistently from different centres in East Asia.

To understand the global allele frequency of the HBV-related SNPs, we collected data from the 1000 Genomes Project, with the results listed in Table 1. Although HBsAg prevalence in East Asians is as high as in Africans, the two populations did not show similar allele frequencies on these SNPs. In general, the allele frequencies of these SNPs are significantly different between East Asians and other global populations. Only 5/19 (26.3%) SNPs showed similar allele frequencies between populations from East Asia and Africa [Table 1]. Therefore, these HBV-related SNPs in the HLA-DP and -DQ loci may not completely explain the high prevalence of HBsAg in Africa.

We suspect that the evolution of these SNPs may be related to human migration^[24]. The Indo-China peninsula and southern China are mountainous and forested areas. Such geographic environments are associated with a great diversity of microorganisms, insects, plants, and animals. People who can survive in this milieu may need some adjustments to their immunity, lest they succumb to a cytokine storm after exposure to multiple unfamiliar microorganisms. Because of modified antigen presentation resulting from HLA-DP and -DQ loci, immunity may be decreased or separated into several stages to avoid the development of cytokine storm. Unfortunately, such immunity may also allow HBV infection to become chronic and persistent. HBV clearance is delayed, but clearance may finally occur several decades later. As a matter of fact, only a minority of HBsAg carriers die of acute or chronic liver disease, and around half of chronic HBsAg carriers clear HBsAg by 80 years of age^[7].

Mechanism of persistent HBV infection in HLA-DP and- DQ SNPs

Both rs3077 and rs9277535 were identified by GWAS to be associated with persistent HBV infection in Japanese patients^[18]. Allele A of rs3077 and rs9277535 are associated with a higher mRNA expression than allele G^[38]. The prevalence of the A allele is lower in East Asia compared to other geographic areas [Table 1].

Table 1. Allele frequency differences between African and East Asian regions on HBV-related SNPs at HLA regions

Gene	SNP	Variant	Allele	Allele frequency					P value AFR vs. EAS
				AFR n = 1,322	AMR n = 694	EUR n = 1,008	SAS n = 978	EAS n = 1,008	
<i>LOC107987449/ LOC107987459</i>	rs9272105	Intron	G	0.417	0.467	0.535	0.447	0.575	NS
<i>HLADQA2-DQB1</i>	rs9275319	Intergenic	G	0.113	0.290	0.163	0.091	0.135	NS
<i>HLADQA2-DQB1</i>	rs2856718	Intergenic	T	0.359	0.38	0.351	0.508	0.528	< 0.001
<i>HLADQA2-DQB1</i>	rs9275572	Intergenic	A	0.399	0.314	0.4	0.281	0.254	< 0.001
<i>HLA-DQA2</i>	rs9276370	2KB Upstream	G	0.711	0.367	0.408	0.215	0.159	< 0.001
<i>HLA-DQB2</i>	rs7756516	3 Prime UTR	C	0.631	0.432	0.468	0.335	0.194	< 0.001
<i>HLA-DQB2</i>	rs7453920	Intron	A	0.305	0.274	0.384	0.192	0.127	< 0.001
<i>HLA-DPA1</i>	rs3077	3 Prime UTR	A	0.419	0.716	0.811	0.633	0.320	< 0.001
<i>HLA-DPA1</i>	rs9277341	Intron	T	0.257	0.597	0.682	0.41	0.165	< 0.001
<i>HLA-DPA1/HLA-DPB1</i>	rs3135021	Intron	A	0.356	0.421	0.279	0.435	0.255	< 0.001
<i>HLA-DPB1</i>	rs9277535	3 Prime UTR	G	0.191	0.288	0.271	0.297	0.612	< 0.001
<i>HLA-DPB1</i>	rs9277542	3 Prime UTR	T	0.402	0.669	0.686	0.634	0.38	NS
<i>HLA-DPB1</i>	rs10484569	Downstream	A	0.048	0.036	0.04	0.025	0.39	< 0.001
<i>HLA-DPA2 (Pseudogene)</i>	rs3128917	Downstream HLADPB1	G	0.511	0.265	0.271	0.281	0.535	NS
<i>HLA-DPA2 (Pseudogene)</i>	rs2281388	Downstream HLADPB1	A	0.002	0.017	0.024	0.024	0.378	< 0.001
<i>HLA-DPA2 (Pseudogene)</i>	rs3117222	Downstream HLADPB1	T	0.509	0.264	0.271	0.28	0.537	NS
<i>HLA-DPB2 (Pseudogene)</i>	rs9380343	2KB Upstream	T	0.048	0.032	0.043	0.028	0.394	< 0.001
<i>LOC105375021</i>	rs9366816	Intron	C	0.157	0.375	0.231	0.177	0.46	< 0.001

AFR: African; AMR: American; EUR: European; SAS: South Asian; EAS: East Asian; UTR: un-transcript region; SNP: single nucleotide polymorphism; HLA: human leukocyte antigen; NS: no significance

This may suggest that the antigen presentation and immune response of Allele G are weaker than those of Allele A. Such behaviours may favour a persistent HBV infection.

The SNP rs7756516 in HLA-DQB2 was associated with persistent HBV infection^[23]. We checked potential mRNA binding using the SegalLab tool (http://genie.weizmann.ac.il/pubs/mir07/mir07_prediction.html) and found that microRNA-550 may bind to the G allele of this SNP. The binding of miR-RNA-550 decreased mRNA stability, a potential reason why the mRNA level is low and weak function of antigen presentation. The allele frequency of the G allele is 0.806 in East Asians [Table 1], which is much higher than in other areas worldwide (0.369-0.665).

HBV-related SNPs in the HLA region among East Asians

The human migration theory was based on a geographic block on the Indo-China peninsula. After crossing this region, the ancestors of East Asians spread to Northern China, Korea, and Japan.

Northern China is generally a grassland and is associated with a lower prevalence of HBsAg than southern China^[16]. With this in mind, we examined the allele frequencies in East Asian populations. We proposed that a lower HBsAg prevalence in northeast Asia could be related to genetic polymorphisms.

The allele frequencies of HBV-related SNPs in East Asia were obtained from the 1000 Genomes Project [Table 2]. A zone in the HLA-DP and -DQ regions showed a trend of allele frequency changes according to HBsAg prevalence and geographic location. On the other hand, background genetics may explain a lower prevalence of HBsAg in North versus Southeast Asians. This observation may be due to the race differences between North- and Southeast Asia. While plausible, such trends were not found in pseudogene regions [Table 2]. We therefore suggest that only active genes participated in the environmental evolution or adaptation. This is additional evidence that supports the role of geographic blocks in the evolution of HBV-related SNPs in HLA regions.

Table 2. Allele frequency trends among east Asian regions according to geographic location on HBV-related SNPs in HLA regions

Gene	SNP	Allele	JPT n = 208	CHB n = 206	CHS n = 210	KHV n = 198	CDX n = 186	P value X ² for trend
HLADQA1-DRB1	rs9272105	A	0.404	0.471	0.49	0.586	0.371	NS
HLADQA2-DQB1	rs9275319	G	0.269	0.141	0.105	0.076	0.075	< 0.0000001
HLA-DQB1	rs2856718	T	0.452	0.519	0.605	0.424	0.645	0.01035
HLA-DQA2/HLA-DQB1	rs9275572	A	0.341	0.311	0.214	0.232	0.161	6.133E-06
HLA-DQA2	rs9276370	G	0.221	0.228	0.09	0.152	0.097	0.00005987
HLA-DQB2	rs7756516	C	0.346	0.248	0.09	0.172	0.108	0.001622
HLA-DQB2	rs7453920	A	0.207	0.189	0.071	0.101	0.059	2.09E-07
HLA-DPA1	rs3077	A	0.413	0.379	0.271	0.303	0.226	0.00001509
HLA-DPA1	rs9277341	T	0.111	0.228	0.129	0.222	0.134	NS
HLA-DPA1/HLA-DPB1	rs3135021	A	0.409	0.301	0.214	0.182	0.156	< 0.0000001
HLA-DPB1	rs9277535	G	0.558	0.539	0.614	0.722	0.634	0.001622
HLA-DPB1	rs9277542	T	0.442	0.461	0.39	0.242	0.355	0.0002659
HLA-DPB1	rs10484569	A	0.375	0.374	0.386	0.394	0.425	0.000229
HLA-DPA2 (Pseudogene)	rs3128917	G	0.553	0.461	0.519	0.606	0.538	NS
HLA-DPA2 (Pseudogene)	rs2281388	A	0.365	0.335	0.371	0.399	0.425	NS
HLA-DPA2 (Pseudogene)	rs3117222	T	0.558	0.461	0.519	0.606	0.543	NS
HLA-DPB2 (Pseudogene)	rs9380343	T	0.375	0.379	0.39	0.394	0.435	NS
LOC105375021	rs9366816	C	0.466	0.481	0.429	0.439	0.489	NS

JPT: Japanese in Tokyo, Japan; CHB: Han Chinese in Beijing, China; CHS: Southern Han Chinese; KHV: Kinh in Ho Chi Minh City, Vietnam; CDX: Chinese Dai in Xishuangbanna, China; SNP: single nucleotide polymorphism; HLA: human leukocyte antigen; HBV: hepatitis B virus; NS: no significance

Hepatocarcinogenesis in HLA loci

Hepatocarcinogenesis is a multifactorial process. There is strong evidence for the role of genetics in persistent HBV infections. However, controversy exists over genetic reports on HBV-related hepatocarcinogenesis. When we examined the global incidence of HBV-related HCC, a higher incidence of HCC could be found in West Africa versus East Africa. Based on this trend, we examined the allele frequency distribution of the reported SNPs and correlated these with HCC incidence in different geographic regions in Africa. One should be notice that the mechanism of hepatocarcinogenesis can be diverse among regions. For example, aflatoxin or other environmental factors may be important in Africans^[39]. On the other hand, a long active HBV replication phase is the key factor in East Asians^[26-29].

In HLA HBV-related SNPs, five SNPs (rs2856718, rs9275572, rs3077 and rs9277341) showed a trend of West-to-East allele frequencies change in Africans ($P < 0.00001$; Table 3). These SNPs were mainly located in HLA-DQ and HLA DPA1 regions; the distribution of these HCC-related SNPs in HLA regions was similar to that observed in persistent HBV infection [Table 2]. All of these SNPs, except rs9277341, were reported to be associated with a greater risk of HCC. The rs9277341 allele had a significant difference in frequency between West and East ($P < 10^{-7}$), but no study had examined its effect on the risk of developing HCC. Further studies regarding this may be needed. The mechanism of hepatocarcinogenesis is probably related to persistent HBV replication and repeated liver necroinflammation.

Non-HLA SNPs in relation to HBV infection and hepatocarcinogenesis

Many SNPs in non-HLA loci were also reported to be associated with HBV infection and/or hepatocarcinogenesis. The associations with HBV infection reported in non-HLA SNPs were generally weaker than those in HLA regions. However, many SNPs related to HBV persistence or carcinogenesis could not be replicated in other studies. This is at least in part due to the different genetic backgrounds across study populations. We saw significant allele frequency differences between Africans and East Asians. Only 2/20 (10%) SNPs showed a similar allele frequency between the two populations [Table 4].

Table 3. Allele frequency trends among African regions according to geographic location on HBV-related SNPs in HLA regions

Gene	SNP	Allele	GWD n = 226	MSL n = 170	YRI n = 216	ESN n = 198	LWK n = 198	P value X ² for trend
HLADQA1-DRB1	rs9272105	G	0.367	0.329	0.394	0.535	0.384	NS
HLADQA2-DQB1	rs9275319	G	0.111	0.235	0.106	0.061	0.091	0.008894
HLA-DQB1	rs2856718	T	0.482	0.371	0.343	0.354	0.288	8.64E-05
HLA-DQA2/HLA-DQB1	rs9275572	A	0.204	0.412	0.472	0.449	0.449	1.95E-07
HLA-DQA2	rs9276370	G	0.743	0.812	0.722	0.742	0.631	0.004733
HLA-DQB2	rs7756516	C	0.588	0.659	0.713	0.662	0.530	NS
HLA-DQB2	rs7453920	A	0.296	0.324	0.282	0.323	0.283	NS
HLA-DPA1	rs3077	A	0.358	0.318	0.264	0.46	0.561	1.26E-06
HLA-DPA1	rs9277341	T	0.15	0.182	0.185	0.268	0.399	< 0.0000001
HLA-DPA1/HLA-DPB1	rs3135021	A	0.403	0.353	0.25	0.328	0.434	NS
HLA-DPB1	rs9277535	G	0.173	0.188	0.116	0.212	0.212	NS
HLA-DPB1	rs9277542	T	0.403	0.312	0.292	0.414	0.48	NS
HLA-DPB1	rs10484569	A	0.018	0.018	0.037	0.086	0.061	0.000635
HLA-DPA2 (Pseudogene)	rs3128917	G	0.491	0.571	0.648	0.505	0.46	NS
HLA-DPA2 (Pseudogene)	rs2281388	A	0	0	0	0	0	NS
HLA-DPA2 (Pseudogene)	rs3117222	T	0.491	0.571	0.648	0.495	0.46	NS
HLA-DPB2 (Pseudogene)	rs9380343	T	0.08	0.082	0.009	0.03	0.02	0.000223
LOC105375021	rs9366816	C	0.159	0.135	0.088	0.197	0.192	NS

GWD: Gambian in Western Divisions in the Gambia; MSL: Mende in Sierra Leone; YRI: Yoruba in Ibadan, Nigeria; ESN: Esan in Nigeria; LWK: Luhya in Webuye, Kenya; SNP: single nucleotide polymorphism; HLA: human leukocyte antigen; HBV: hepatitis B virus; NS: no significance

Table 4. Allele frequency differences between African and East Asian regions in HBV-related SNPs at non-HLA regions

Position	Gene	SNP	Variant	Allele	AFR	AMR	EUR	SAS	EAS	AFR vs. EAS
1:10325413	<i>KIF1</i>	rs17401966	Intron	G	0.057	0.32	0.309	0.272	0.288	< 0.001
2:112774138	<i>IL-1A/MIR-122; DELINS</i>	rs16347	Indel 3 prime UTR	TGAA	0.195	0.477	0.322	0.286	0.704	< 0.001
2:112836810	<i>IL-10</i>	rs1800872	5 prime UTR	G	0.564	0.667	0.76	0.542	0.324	< 0.001
2:191099907	<i>STAT4</i>	rs7574865	Intron	T	0.116	0.362	0.23	0.30	0.347	< 0.001
2:203867624	<i>CTLA4</i>	rs5742909	Upstream	T	0.003	0.063	0.084	0.029	0.097	< 0.001
2:203867991	<i>CTLA4</i>	rs231775	Missense	A	0.612	0.537	0.641	0.69	0.363	< 0.001
2:211380366-7	<i>ERBB4</i>	rs6147150	Indel 3 prime UTR	TG	0.591	0.365	0.399	0.244	0.284	< 0.001
3:157429779	<i>VEPH1</i>	rs2120243	intron	A	0.332	0.308	0.43	0.327	0.348	NS
4:186082920	<i>TLR-3</i>	rs3775291	Missense	T	0.026	0.305	0.324	0.263	0.328	< 0.001
6:31162816	<i>CTF19</i>	rs1419881	3 Prime UTR	G	0.363	0.496	0.547	0.55	0.433	< 0.001
6:31398818	<i>MICA-AS1/MICA</i>	rs2596542	Intron/Upstream	T	0.541	0.519	0.396	0.369	0.275	< 0.001
6:31575254	<i>TNFA</i>	rs1800629	Upstream	A	0.12	0.069	0.134	0.053	0.059	< 0.001
6:31575324	<i>TNFA</i>	rs361525	Upstream	A	0.038	0.082	0.064	0.105	0.031	NS
7:100103553	<i>AP4M1/MCM7/MIR-106b</i>	rs999885	Intron/Upstream	G	0.78	0.411	0.472	0.34	0.192	< 0.001
11:86921775	<i>PRSS23/LOC107984428</i>	rs1048338	intron/NCT	C	0.3	0.254	0.14	0.303	0.397	< 0.001
11:112164265	<i>IL-18</i>	rs187238	2KB Upstream	G	0.20	0.314	0.278	0.184	0.121	< 0.001
13:32014688	<i>GRIK1</i>	rs455804	Intron	A	0.416	0.228	0.236	0.195	0.313	< 0.001
14:69778476	<i>NTCP</i>	rs2296651	Missense	A	0.000	0.000	0.000	0.000	0.071	< 0.001
14:69796098	<i>NTCP</i>	rs4646287	Intron	T	0.001	0	0	0.018	0.101	< 0.001
18:5900774	<i>TMEM200C</i>	rs2212522	2KB Upstream	T	0.586	0.344	0.232	0.41	0.502	< 0.001

AFR: African; AMR: American; EUR: European; SAS: South Asian; EAS: East Asian; UTR: un-transcript region; NCT: non-coding transcript; SNP: single nucleotide polymorphism; HLA: human leukocyte antigen; HBV: hepatitis B virus; NS: no significance

HBV-related SNPs in the non-HLA region among East Asians

Most of the persistent HBV infection-related SNPs in non-HLA regions were reported in East Asia. We examined whether these SNPs also showed geographical differences in allele frequencies between Northern and Southern regions. We found that only 2 of 20 (10%) SNPs showed significant North-to-South allele frequency trend in East Asians [Table 5]. Among these, NTCP, a functional receptor of hepatitis B^[40], showed the highest trend ($P = 2.23 \times 10^{-6}$).

Table 5. Allele frequency trends among east Asian regions according to geographic location on HBV-related SNPs in non-HLA regions

Gene	SNP	Allele	JPT	CHB	CHS	KHV	CDX	P value χ^2 for trend
<i>KIF1</i>	rs17401966	G	0.293	0.286	0.367	0.273	0.21	NS
<i>IL-1A/MIR-122; DELINS</i>	rs16347	TGAA	0.726	0.631	0.729	0.747	0.688	NS
<i>IL-10</i>	rs1800872	G	0.361	0.257	0.314	0.343	0.349	NS
<i>STAT4</i>	rs7574865	T	0.327	0.354	0.352	0.354	0.349	NS
<i>CTLA4</i>	rs5742909	T	0.096	0.117	0.114	0.086	0.07	NS
<i>CTLA4</i>	rs231775	A	0.375	0.311	0.343	0.338	0.457	NS
<i>ERBB4</i>	rs6147150	TG	0.226	0.252	0.286	0.323	0.339	0.003133
<i>VEPH1</i>	rs2120243	A	0.274	0.335	0.376	0.354	0.409	0.007567
<i>TLR-3</i>	rs3775291	T	0.293	0.291	0.329	0.389	0.344	0.05213
<i>CTF19</i>	rs1419881	G	0.5	0.495	0.448	0.414	0.29	1.08E-05
<i>MICA-AS1/MICA</i>	rs2596542	C	0.332	0.272	0.229	0.308	0.231	NS
<i>TNFA</i>	rs1800629	A	0.019	0.092	0.057	0.056	0.07	NS
<i>TNFA</i>	rs361525	A	0.014	0.034	0.038	0.056	0.011	NS
<i>AP4M1/MCM7/MIR-106b</i>	rs999885	G	0.168	0.184	0.19	0.197	0.226	NS
<i>RSS23/LOC107984428</i>	rs1048338	C	0.399	0.456	0.333	0.389	0.409	NS
<i>IL-18</i>	rs187238	G	0.159	0.083	0.114	0.116	0.134	NS
<i>GRIK1</i>	rs455804	A	0.216	0.345	0.343	0.298	0.371	0.01208
<i>NTCP</i>	rs2296651	A	0.024	0.029	0.081	0.111	0.118	2.23E-06
<i>NTCP</i>	rs4646287	T	0.154	0.102	0.071	0.086	0.091	0.035
<i>TMEM200C</i>	rs2212522	T	0.433	0.519	0.5	0.515	0.548	0.04036

JPT: Japanese in Tokyo, Japan; CHB: Han Chinese in Beijing, China; CHS: Southern Han Chinese; KHV: Kinh in Ho Chi Minh City, Vietnam; CDX: Chinese Dai in Xishuangbanna, China; SNP: single nucleotide polymorphism; HLA: human leukocyte antigen; HBV: hepatitis B virus; NS: no significance

The T allele of rs2296651 is a missense mutation^[41] and is found only in East Asians [Table 4]. There is a higher T-allele frequency in Southern East Asia (0.111-0.118) than in Northern East Asia (0.024-0.029) [Table 5]. However, the T allele is known to protect against persistent HBV infection. A higher T-allele frequency in a region with a high prevalence of HBsAg requires an explanation. We propose that the T allele is an evolutionary mechanism to defend against persistent HBV infection in the presence of a weakened antigen presentation system.

The rs1419881 in transcription factor 19 (CTF19) shows significant allele frequency differences between the Northern and Southern regions ($P = 1.08 \times 10^{-5}$ Table 5). This GWAS-identified SNP was found to be associated with persistent HBV infection in Korea^[21]. This SNP was validated in China but was not associated with persistent HBV infection in the Thai population^[42]. The G allele is the risk-associated allele, which showed a higher frequency in Japanese people in Tokyo, Japan (JPT; 0.5) than in Chinese Dai people in Xishuangbanna, China (CDX; 0.29). This association was inversely related to HBsAg prevalence [Table 5]. CTF19 mainly plays a role in the transcription of genes required in the later stages of cell cycle progression. Its mechanism in persistent HBV infection is unclear. Whether it is also similar to NTCP, which is associated with an increased defensive response in people living in regions with a high HBsAg prevalence, will require future studies.

Hepatocarcinogenesis in non-HLA loci

The major histocompatibility complex class I-related chain A (MICA) was reported to be associated with HCV-related HCC^[43]. In non-HLA HBV-related SNPs, only rs2596542 in MICA showed a significant trend ($P = 0.00011$; Table 6). Its C allele frequency is lower in West Africa than in East Africa. The C allele is protective against hepatocarcinogenesis, whereas the T allele is a risk factor^[44]. These findings correlate with a higher incidence of HCC in West than in East Africa. The MICA molecule is a ligand of the natural killing group 2 member D molecule, which is involved in nature killer cell function. Some of the tumour cell may relieve soluble MICA molecules to block immune surveillance^[45,46].

Table 6. Allele frequency trends among African regions according to geographic location on HBV-related SNPs in non-HLA regions

Gene	SNP	Allele	GWD	MSL	YRI	ESN	LWK	P value X ² for trend
<i>KIF1</i>	rs17401966	G	0.04	0.035	0.037	0.051	0.096	0.009852
<i>IL-1A/MIR-122; DELINS</i>	rs16347	TGAA	0.15	0.159	0.204	0.202	0.263	0.002744
<i>IL-10</i>	rs1800872	G	0.527	0.524	0.532	0.551	0.606	NS
<i>STAT4</i>	rs7574865	T	0.071	0.129	0.125	0.131	0.131	0.05661
<i>CTLA4</i>	rs231775	A	0.593	0.706	0.648	0.616	0.52	0.05029
<i>ERBB4</i>	rs6147150	TG	0.659	0.594	0.588	0.505	0.641	NS
<i>VEPH1</i>	rs2120243	A	0.296	0.335	0.343	0.364	0.293	NS
<i>TLR-3</i>	rs3775291	T	0.018	0.012	0.009	0.005	0.035	NS
<i>CTF19</i>	rs1419881	G	0.429	0.347	0.361	0.288	0.379	NS
<i>MICA-AS1/MICA</i>	rs2596542	C	0.429	0.565	0.491	0.657	0.586	0.00011
<i>TNFA</i>	rs1800629	A	0.142	0.159	0.102	0.126	0.086	0.05056
<i>TNFA</i>	rs361525	A	0.084	0.047	0.005	0.01	0.061	0.04351
<i>HLADQA1-DRB1</i>	rs9272105	G	0.062	0.129	0.199	0.253	0.242	< 0.0000001
<i>HLADQA2-DQB1</i>	rs9275319	G	0.111	0.235	0.106	0.061	0.091	0.008894
<i>AP4M1/MCM7/MIR-106b</i>	rs999885	G	0.752	0.835	0.755	0.813	0.838	0.07368
<i>PRSS23/LOC107984428</i>	rs1048338	C	0.412	0.3	0.259	0.273	0.273	0.001384
<i>IL-18</i>	rs187238	G	0.248	0.166	0.204	0.207	0.162	0.006079
<i>GRIK1</i>	rs455805	A	0.358	0.382	0.472	0.49	0.399	0.08
<i>NTCP</i>	rs4646287	T	0.0	0.0	0.0	0.0	0.005	NS
<i>TMEM200C</i>	rs2212522	T	0.615	0.671	0.528	0.581	0.581	NS

GWD: Gambian in Western Divisions in the Gambia; MSL: Mende in Sierra Leone; YRI: Yoruba in Ibadan, Nigeria; ESN: Esan in Nigeria; LWK: Luhya in Webuye, Kenya; SNP: single nucleotide polymorphism; HLA: human leukocyte antigen; HBV: hepatitis B virus; NS: no significance

The rest of non-HLA-related SNPs show weak or absent West-to-East allele frequency trend. This also confirms a low power of hepatocarcinogenesis in each HBV-related SNP.

The rs17401966 SNP in Kinesin Family Member 1B is a tumour suppressor gene. It has been identified by GWAS to be associated with HCC^[47], although there is some controversy in the subsequent validation studies. A meta-analysis has revealed that the G allele is a protective allele in the Chinese population^[48]. This G allele shows a higher prevalence in East Asia than in Africa (0.288 vs. 0.057, $P < 0.001$; Table 4). It is interesting to find that there is a trend of a higher G allele frequency in the Luhya people in Webuye, Kenya (LWK), east Africa, than in the Gambian people in Western Divisions in the Gambia (GWD), West Africa (0.096 vs. 0.04, $P = 0.009852$; Table 6). This is compatible with a lower incidence of HCC in East Africa versus in West Africa.

STATs pathway and associated cytokines play roles on hepatitis clearance and fibrogenesis^[49,50]. The rs7574865 SNP in signal transducer and activator of transcription 4 (STAT4) is related to persistent HBV infection and HCC. The T allele shows a lower prevalence in HCC than in chronic hepatitis B^[51]. There is a weak East-to-West T-allele frequency trend in Africa (0.131 in LWK and 0.071 in GWD; $P = 0.05561$; Table 6). However, the higher incidence of HCC in lower T-allele frequency areas supports the conclusion made by the meta-analysis.

The SNP rs16347 in the 3'-untranslated regions of interleukin-1alpha (IL-1A) carries a miRNA-122 binding site. A variant with a TGAA insertion decreases miRNA-122 binding and increases IL-1A mRNA expression^[52]. The prevalence of this insertion variant is low in Southern Chinese patients with HCC. However, another study from China has not shown this result but instead associates the insertion variant with HBV genome mutants^[53]. When we looked at the allele frequency in Africa, the TGAA insertion variant was lower in West Africa than in East Africa. This allele frequency correlated with the lower

incidence of HCC in East Africa than in West Africa [Table 6]. It should be noted that the function of miRNA-122 is complicated and that the frequency of this insertion variant in East Asians (0.704) is much higher than in Africans (0.199; Table 4). If this SNP is associated with HCC, then it will have a significant impact in East Asians.

Rs187238 is located upstream of interleukin-18 (IL-18) (-148 G>C). The G allele induces increased IL-18 mRNA expression compared to the C allele^[54]. The frequency of the G allele is lower in HCC cases than in non-HCC cases. This implies that those with a stronger immunity may be able to control HBV and hepatocarcinogenesis. However, a follow-up study did not validate the allele differences between HCC and non-HCC5 cases^[55]. In this review, the G-allele frequency was higher in West than in East Africa (0.248 to 0.152; Table 6), contradicting the higher incidence of HCC in West Africa. The incident epidemiology does not support rs187238 playing a role in hepatocarcinogenesis in Africans. This does not necessarily provide evidence against the association of this SNP with hepatocarcinogenesis in East Asians. Termination of HBV replication is more important in East Asians than in Africans.

Rs1048338 in PRSS23 has been identified by GWAS to be associated with HCC in China^[56]. However, no other report has validated this observation. A trend of a higher C allele frequency in West compared to East Africa (0.412 to 0.273, $P = 0.001384$; Table 6) has also been observed. This finding is not against the association rs1048338 with hepatocarcinogenesis. However, more studies are needed to confirm its association with HBV-related HCC.

CONCLUSION

We confirm that there are significant differences in genetic background between Africa and East Asia. By correlating genetic backgrounds with clinical epidemiology, we have found that the allele frequency of HLA-DQ and -DP loci do explain a higher prevalence of HBsAg in Southeastern compared to Northeastern Asia. Some of these SNPs also showed West-to-East changes in allele frequency in Africa and are correlated with HCC incidence. For the non-HLA loci, SNPs in NTCP and CTF19 showed allele frequency trends from North-to-South in East Asians, supporting their association with fighting in persistent HBV infection. There is a strong correlation between allele frequency and HCC incidence on SNPs located in MICA and weak positive correlations in KIF1, STAT4, and IL1A. The studies concerning genetic factors and hepatocarcinogenesis are difficult since multiple factors are involved and different genetic backgrounds exist among the study populations.

DECLARATIONS

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Authors' contributions

Designed the study and wrote the manuscript: Tai DI

Collected and organized data: Tai J

Availability of data and materials

The data source is from the 1000 Genomes Project (<http://www.1000genomes.org/>).

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Conflicts of interest

Both authors declared that there are no conflicts of interest.

Ethical approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

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REFERENCES

1. Schweitzer A, Horn J, Mikolajczyk RT, Krause G, Ott JJ. Estimations of worldwide prevalence of chronic hepatitis B virus infection: a systematic review of data published between 1965 and 2013. *Lancet* 2015;386:1546-55.
2. Chen CJ, Tai J, Tai DI. Hepatocellular carcinoma occurred in a Hepatitis B carrier clinic cohort during a mean follow up of 10 years. *Hepatoma Res* 2019;5:25.
3. Yu MW, Chang HC, Liaw YF, et al. Familial risk of hepatocellular carcinoma among chronic hepatitis B carriers and their relatives. *J Natl Cancer Inst* 2000;92:1159-64.
4. Burk RD, Hwang LY, Ho GY, Shafritz DA, Beasley RP. Outcome of perinatal hepatitis B virus exposure is dependent on maternal virus load. *J Infect Dis* 1994;170:1418-23.
5. Tai DI, Chen CH, Chang TT, et al. Eight-year nationwide survival analysis in relatives of patients with hepatocellular carcinoma: role of viral infection. *J Gastroenterol Hepatol* 2002;17:682-9.
6. Chen CH, Chen YY, Chen GH, et al. Hepatitis B virus transmission and hepatocarcinogenesis: a 9 year retrospective cohort of 13676 relatives with hepatocellular carcinoma. *J Hepatol* 2004;40:653-9.
7. Tai DI, Tsay PK, Chen WT, Chu CM, Liaw YF. Relative roles of HBsAg seroclearance and mortality in the decline of HBsAg prevalence with increasing age. *Am J Gastroenterol* 2010;105:1102-9.
8. Liaw YF, Tai DI, Chu CM, Chen TJ. The development of cirrhosis in patients with chronic type B hepatitis: a prospective study. *Hepatology* 1988;8:493-6.
9. Cardin R, Picocchi M, Bortolami M, et al. Oxidative damage in the progression of chronic liver disease to hepatocellular carcinoma: an intricate pathway. *World J Gastroenterol* 2014;20:3078-86.
10. Liu WC, Wu IC, Lee YC, et al. Hepatocellular carcinoma-associated single-nucleotide variants and deletions identified by the use of genome-wide high-throughput analysis of hepatitis B virus. *J Pathol* 2017;243:176-92.
11. Zhang Z, Wang C, Liu Z, et al. Host genetic determinants of hepatitis B virus infection. *Front Genet* 2019;10:696.
12. Akcay IM, Katrinli S, Ozdil K, Doganay GD, Doganay L. Host genetic factors affecting hepatitis B infection outcomes: insights from genome-wide association studies. *World J Gastroenterol* 2018;24:3347-60.
13. Zhu H, Wu J, Shen X. Genome-wide association study: new genetic insights into HBV/HCV-related hepatocellular carcinoma genomes. *Scand J Gastroenterol* 2017;52:209-15.
14. Hai H, Tamori A, Kawada N. Role of hepatitis B virus DNA integration in human hepatocarcinogenesis. *World J Gastroenterol* 2014;20:6236-43.
15. 1000 Genomes Project Consortium, Auton A, Brooks LD, et al. A global reference for human genetic variation. *Nature* 2015;526:68-74.
16. Xia GL, Liu CB, Cao HL, et al. Prevalence of hepatitis B and C virus infections in the general Chinese population. Results from a nationwide cross-sectional seroepidemiologic study of hepatitis A, B, C, D, and E virus infections in China, 1992. *Int Hepatol Comm* 1996;5:62-73.
17. Kamatani Y, Watanapokayakit S, Ochi H, et al. A genome-wide association study identifies variants in the HLA-DP locus associated with chronic hepatitis B in Asians. *Nat Genet* 2009;41:591-5.
18. Mbarek H, Ochi H, Urabe Y, et al. A genome-wide association study of chronic hepatitis B identified novel risk locus in a Japanese population. *Hum Mol Genet* 2011;20:3884-92.
19. Hu Z, Liu Y, Zhai X, et al. New loci associated with chronic hepatitis B virus infection in Han Chinese. *Nat Genet* 2013;45:1499-503.
20. Nishida N, Sawai H, Matsuura K, et al. Genome-wide association study confirming association of HLA-DP with protection against chronic hepatitis B and viral clearance in Japanese and Korean. *PLoS One* 2012;7:e39175.
21. Kim YJ, Kim HY, Lee JH, et al. A genome-wide association study identified new variants associated with the risk of chronic hepatitis B. *Hum Mol Genet* 2013;22:4233-8.
22. Al-Qahtani AA, Al-Anazi MR, Abdo AA, Sanai FM, Al-Hamoudi W, et al. Association between HLA variations and chronic hepatitis B virus infection in Saudi Arabian patients. *PLoS One* 2014;9:e80445.
23. Chang SW, Fann CS, Su WH, et al. A genome-wide association study on chronic HBV infection and its clinical progression in male Han-Taiwanese. *PLoS One* 2014;9:e99724.
24. Tai DI, Jeng WJ, Lin CY. A global perspective on HBV-related SNPs and evolution during human migration. *Hepatol Commun* 2017;1:1005-13.
25. Liaw YF, Chu CM. Hepatitis B virus infection. *Lancet* 2009;373:582-92.
26. Chang MH, Hsu HY, Hsu HC, et al. The significance of spontaneous hepatitis B e antigen seroconversion in childhood: with special

- emphasis on the clearance of hepatitis B e antigen before 3 years of age. *Hepatology* 1995;22:1387-92.
27. Hsu YS, Chien RN, Yeh CT, et al. Long-term outcome after spontaneous HBsAg seroconversion in patients with chronic hepatitis B. *Hepatology* 2002;35:1522-7.
 28. Lin CL, Kao JH. Natural history of acute and chronic hepatitis B: The role of HBV genotypes and mutants. *Best Pract Res Clin Gastroenterol* 2017;31:249-55.
 29. Chang MH, Sung JL, Lee CY, et al. Factors affecting clearance of hepatitis B e antigen in hepatitis B surface antigen carrier children. *J Pediatr* 1989;115:386-90.
 30. Hadziyannis SJ. Natural history of chronic hepatitis B in Euro-Mediterranean and African countries. *J Hepatol* 2011;55:183-91.
 31. Iorio R, Giannattasio A, Cirillo F, D' Alessandro L, Vegnente A. Long-term outcome in children with chronic hepatitis B: a 24-year observation period. *Clin Infect Dis* 2007;45:943-9.
 32. Liu S, Koh SS, Lee CG. Hepatitis B virus X protein and hepatocarcinogenesis. *Int J Mol Sci* 2016;17:E940.
 33. Tu T, Budzinska MA, Vondran FWR, Shackel NA, Urban S. Hepatitis B virus DNA integration occurs early in the viral life cycle in an in vitro infection model via sodium taurocholate cotransporting polypeptide-dependent uptake of enveloped virus particles. *J Virol* 2018;92:e02007-17
 34. Budzinska MA, Shackel NA, Urban S, Tu T. Cellular genomic sites of hepatitis B virus DNA integration. *Genes (Basel)* 2018;9:E365.
 35. Yang L, Ye S, Zhao X, et al. Molecular characterization of HBV DNA integration in patients with hepatitis and hepatocellular carcinoma. *J Cancer* 2018;9:3225-35.
 36. An P, Xu J, Yu Y, Winkler CA. Host and viral variation in HBV-related hepatocellular carcinoma. *Front Genet* 2018;9:261
 37. Lazarevic I. Clinical implications of hepatitis B virus mutations: recent advances. *World J Gastroenterol* 2014;20:7653-64.
 38. O'Brien TR, Kohaar I, Pfeiffer RM, et al. Risk alleles for chronic hepatitis B are associated with decreased mRNA expression of HLA-DPA1 and HLA-DPB1 in normal human liver. *Genes Immun* 2011;12:428-33.
 39. Yang JD, Hainaut P, Gores GJ, et al. A global view of hepatocellular carcinoma: trends, risk, prevention and management. *Nat Rev Gastroenterol Hepatol* 2019;16:589-604.
 40. Yan H, Zhong G, Xu G, et al. Sodium taurocholate cotransporting polypeptide is a functional receptor for human hepatitis B and D virus. *Elife* 2012;1:e00049.
 41. Li N, Zhang P, Yang C, et al. Association of genetic variation of sodium taurocholate cotransporting polypeptide with chronic hepatitis B virus infection. *Genet Test Mol Biomarkers* 2014;18:425-9.
 42. Posuwan N, Payungporn S, Tangkijvanich P, et al. Genetic association of human leukocyte antigens with chronicity or resolution of hepatitis B infection in Thai population. *PLoS One* 2014;9:e86007.
 43. Kumar V, Kato N, Urabe Y, et al. Genome-wide association study identifies a susceptibility locus for HCV-induced hepatocellular carcinoma. *Nat Genet* 2011;43:455
 44. Luo X, Wang Y, Shen A, Deng H, Ye M. Relationship between the rs2596542 polymorphism in the MICA gene promoter and HBV/HCV infection-induced hepatocellular carcinoma: a meta-analysis. *BMC Med Genet* 2019;20:142.
 45. Diefenbach A, Jensen ER, Jamieson AM, Raulet DH. Rae1 and H60 ligands of the NKG2D receptor stimulate tumour immunity. *Nature* 2001;413:165-71.
 46. Groh V, Wu J, Yee C, Spies T. Tumour-derived soluble MIC ligands impair expression of NKG2D and T-cell activation. *Nature* 2002;419:734-8.
 47. Zhang H, Zhai Y, Hu Z, et al. Genome-wide association study identifies 1p36.22 as a new susceptibility locus for hepatocellular carcinoma in chronic hepatitis B virus carriers. *Nat Genet* 2010;42:755-8.
 48. Luo YY, Zhang HP, Huang AL, Hu JL. Association between KIF1B rs17401966 genetic polymorphism and hepatocellular carcinoma susceptibility: an updated meta-analysis. *BMC Med Genet* 2019;20:59.
 49. Kong XN, Horiguchi N, Mori M, Guo B. Cytokines and STATs in liver fibrosis. *Front Physiol* 2012;3:69.
 50. Gao B. Cytokines, STATs and liver disease. *Cell Mol Immunol* 2005;2:92-100.
 51. Shi H, He H, Ojha SC, et al. Association of STAT3 and STAT4 polymorphisms with susceptibility to chronic hepatitis B virus infection and risk of hepatocellular carcinoma: a meta-analysis. *Biosci Rep* 2019;39:BSR20190783
 52. Gao Y, He Y, Ding J, et al. An insertion/deletion polymorphism at miRNA-122-binding site in the interleukin-1alpha 3' untranslated region confers risk for hepatocellular carcinoma. *Carcinogenesis* 2009;30:2064-9.
 53. Du Y, Han X, Pu R, et al. Association of miRNA-122-binding site polymorphism at the interleukin-1 alpha gene and its interaction with hepatitis B virus mutations with hepatocellular carcinoma risk. *Front Med* 2014;8:217-26.
 54. Kim YS, Cheong JY, Cho SW, et al. A functional SNP of the Interleukin-18 gene is associated with the presence of hepatocellular carcinoma in hepatitis B virus-infected patients. *Dig Dis Sci* 2009;54:2722-8.
 55. Zhu SL, Zhao Y, Hu XY, et al. Genetic polymorphisms -137 (rs187238) and -607 (rs1946518) in the interleukin-18 promoter may not be associated with development of hepatocellular carcinoma. *Sci Rep* 2016;6:39404.
 56. Qu LS, Jin F, Guo YM, et al. Nine susceptibility loci for hepatitis B virus-related hepatocellular carcinoma identified by a pilot two-stage genome-wide association study. *Oncol Lett* 2016;11:624-32.