

Review

Open Access



# Glycogen synthase kinase 3 $\beta$ : the nexus of chemoresistance, invasive capacity, and cancer stemness in pancreatic cancer

Masahiro Uehara<sup>1,\*</sup> , Takahiro Domoto<sup>1,#</sup> , Satoshi Takenaka<sup>1,2,3</sup> , Osamu Takeuchi<sup>4</sup> , Takeo Shimasaki<sup>1,5</sup> , Tomoharu Miyashita<sup>1,2,3</sup> , Toshinari Minamoto<sup>1</sup> 

<sup>1</sup>Division of Translational and Clinical Oncology, Cancer Research Institute, Kanazawa University, Kanazawa 920-0934, Japan.

<sup>2</sup>Department of Hepato-Biliary-Pancreatic Surgery and Transplantation, Graduate School of Medical Sciences, Kanazawa University, Kanazawa 920-8641, Japan.

<sup>3</sup>Department of Surgery, Toyama City Hospital, Toyama 939-8511, Japan.

<sup>4</sup>Biomedical Laboratory, Department of Research, Kitasato University Kitasato Institute Hospital, Tokyo 108-8642, Japan.

<sup>5</sup>Medical Research Institute, Kanazawa Medical University, Uchinada 920-0293, Japan.

\*Authors contributed equally.

**Correspondence to:** Dr. Toshinari Minamoto, Division of Translational and Clinical Oncology, Cancer Research Institute, Kanazawa University, 13-1 Takara-machi, Kanazawa 920-0934, Japan. E-mail: minamoto@staff.kanazawa-u.ac.jp

**How to cite this article:** Uehara M, Domoto T, Takenaka S, Takeuchi O, Shimasaki T, Miyashita T, Minamoto T. Glycogen synthase kinase 3 $\beta$ : the nexus of chemoresistance, invasive capacity, and cancer stemness in pancreatic cancer. *Cancer Drug Resist* 2024;7:4. <https://dx.doi.org/10.20517/cdr.2023.84>

**Received:** 28 Jul 2023 **First Decision:** 1 Dec 2023 **Revised:** 20 Dec 2023 **Accepted:** 17 Jan 2024 **Published:** 31 Jan 2024

**Academic Editor:** Godefridus J. Peters **Copy Editor:** Pei-Yun Wang **Production Editor:** Pei-Yun Wang

## Abstract

The treatment of pancreatic cancer remains a significant clinical challenge due to the limited number of patients eligible for curative (RO) surgery, failures in the clinical development of targeted and immune therapies, and the pervasive acquisition of chemotherapeutic resistance. Refractory pancreatic cancer is typified by high invasiveness and resistance to therapy, with both attributes related to tumor cell stemness. These malignant characteristics mutually enhance each other, leading to rapid cancer progression. Over the past two decades, numerous studies have produced evidence of the pivotal role of glycogen synthase kinase (GSK)3 $\beta$  in the progression of over 25 different cancer types, including pancreatic cancer. In this review, we synthesize the current knowledge on the pathological roles of aberrant GSK3 $\beta$  in supporting tumor cell proliferation and invasion, as well as its contribution to gemcitabine resistance in pancreatic cancer. Importantly, we discuss the central role of GSK3 $\beta$  as a molecular hub that mechanistically connects chemoresistance, tumor cell invasion, and stemness in pancreatic cancer. We also discuss the involvement of GSK3 $\beta$  in the formation of desmoplastic tumor stroma and in promoting anti-



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, sharing, adaptation, distribution and reproduction in any medium or format, for any purpose, even commercially, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.



cancer immune evasion, both of which constitute major obstacles to successful cancer treatment. Overall, GSK3 $\beta$  has characteristics of a promising therapeutic target to overcome chemoresistance in pancreatic cancer.

**Keywords:** Pancreatic cancer, chemoresistance, tumor invasion, cancer stemness, glycogen synthase kinase 3 $\beta$

## INTRODUCTION

Approximately one-third of patients diagnosed with pancreatic ductal adenocarcinoma (PDAC) present with locally unresectable tumors, while nearly half exhibit distant metastatic tumors. Consequently, only 10%-15% of stage I or II patients can undergo resectable (R0) surgery, a significant proportion of whom eventually suffer local disease recurrence and/or distant metastasis post-surgery<sup>[1,2]</sup>. First-line chemotherapeutic regimens for palliative treatment of locally advanced pancreatic cancer and metastatic cases include FOLFIRINOX [a combination of folate, 5-fluorouracil (FU), irinotecan, and oxaliplatin]<sup>[3]</sup>, and a combination of nanoparticle albumin-bound (nab)-paclitaxel and gemcitabine<sup>[4]</sup>. The second-line regimen is FOLFIRI<sup>[5]</sup>. These multi-agent chemotherapy regimens offer only a modestly improved efficacy compared to gemcitabine monotherapy, and are appropriate only for a small proportion of pancreatic cancer patients with a good performance status (PS 0 or 1)<sup>[3-6]</sup>. Patients with a PS of 2 or higher, who represent most pancreatic cancer cases, undergo gemcitabine monotherapy<sup>[7,8]</sup>, with many quickly developing resistance to the drug<sup>[9-11]</sup>. Unlike lung and colorectal cancer, neither preclinical studies nor clinical trials have yet to demonstrate significant efficacy against pancreatic cancer of rationally targeted agents, precision medicines based on the identification of actionable proto-oncoproteins, or immune checkpoint blockade<sup>[12-20]</sup>. Given these circumstances, the development of biology-based strategies against chemoresistance, specifically the resistance to gemcitabine by pancreatic cancer, is a pressing need in pancreatic cancer research<sup>[21]</sup>.

## MECHANISMS OF GEMCITABINE RESISTANCE IN PANCREATIC CANCER

Many reviews<sup>[9-11,22-25]</sup> have thoroughly examined the mechanisms of action of, and resistance to, gemcitabine in pancreatic cancer cells, detailing the steps of intra- and extracellular drug transportation<sup>[26-29]</sup>, metabolic activation of gemcitabine<sup>[30-35]</sup>, molecular pathways that combat drug-induced apoptosis<sup>[36-41]</sup>, pro-oncogenic pathways that enable tumor cells to survive cytotoxic effects<sup>[36,42-51]</sup>, and epithelial-mesenchymal transition (EMT), a pro-invasive tumor property<sup>[52,53]</sup>. These reviews<sup>[10,11,22-25]</sup> also discuss the role of specific micro-RNAs in altering the expression of molecules linked to gemcitabine resistance. Besides the resistance mechanisms emerging in pancreatic cancer cells, comprehensive reviews have discussed the many mechanisms for chemoresistance, including resistance to gemcitabine, targeted agents, and immune checkpoint blockades, brought on by tumor cell interactions with the tumor microenvironment (TME)<sup>[54-63]</sup> [Supplementary Table 1].

Efforts have been made to address resistance mechanisms that interfere with gemcitabine uptake and activation, with several compounds developed to enhance nucleoside transporter (NP) expression or bypass NPs and deoxycytidine kinase (dCK) through different mechanisms or chemical modifications of gemcitabine<sup>[9,11,64]</sup>. Several experimental studies have sought to reverse chemoresistance by activating multiple cell death pathways<sup>[65]</sup>, deconstructing the desmoplastic stroma, and targeting immunosuppressive pathways within the hostile TME in pancreatic cancer<sup>[66-74]</sup>. Despite a decade of extensive research efforts, there has been limited therapeutic advancement in preclinical or clinical settings. This reaffirms the urgent need to identify mechanism-based strategies and therapeutic targets to overcome gemcitabine resistance in pancreatic cancer research.

## MUTUAL DEPENDENCY OF CHEMORESISTANCE, TUMOR CELL INVASION, AND STEMNESS IN PANCREATIC CANCER

One of the fundamental challenges in addressing cancer drug resistance stems from the complex biological properties that tumor cells inherently possess (e.g., intrinsic resistance) or acquire during exposure to therapeutic agents (acquired resistance)<sup>[75]</sup>. Recent comprehensive reviews have detailed these properties, which include altered pharmacological reactions of tumor cells to drugs and their intermediate metabolites, clonal evolution of tumor cells to generate resistant clones, the latency and plasticity of cancer stem cells or tumor cells' resilience to acquiring stemness phenotype, genetic and biological heterogeneity of tumor cells, activation of pro-survival pathways, impairment in cell death pathways, and adaptation to therapeutic pressures<sup>[76,77]</sup>. The aforementioned properties synergistically lead to increased resistance to therapy in various cancer types<sup>[77]</sup>.

The concept of cancer stem cells (CSCs) has been proposed based on the observation that the uncontrolled proliferation of cancer is driven by a biologically distinct subset of tumor cells that are present in a relatively small proportion within the whole tumor<sup>[78,79]</sup>. CSCs are characterized by their ability to self-renew, sustain tumor propagation, express specific cell surface markers, and use multidrug efflux pumps. The clinical implication of CSCs lies in their potential to drive tumor regrowth after seemingly successful treatment with chemotherapeutics, radiation, or targeted agents because of their inherent resistance to therapy<sup>[80-83]</sup>. This has positioned CSCs as a critical therapeutic target, albeit one that is currently elusive due to their complex biological nature in intractable cancer types, including pancreatic cancer<sup>[84-90]</sup>. Several studies have shown that cancer therapy spares not only CSCs but also some residual cancer cells that acquire a CSC-like phenotype without mutation-based clonal selection, thus becoming resistant to therapy. This phenotypic switch is often associated with the activation of pro-oncogenic pathways such as Wnt- and Notch-mediated pathways<sup>[91]</sup>. Consistently, gemcitabine treatment can induce a shift towards a cancer stemness phenotype, primarily in gemcitabine-naïve pancreatic cancer cells<sup>[53,92-95]</sup> [Table 1, Figure 1].

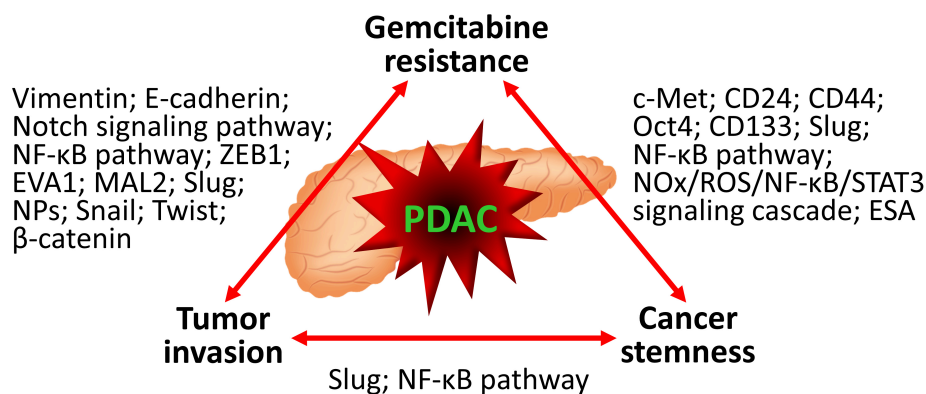
Highly invasive capacity and therapy resistance are the defining biological and clinical characteristics of pancreatic cancer, often resulting in treatment failure and poor patient outcomes<sup>[1,2]</sup>. Although not sufficient on its own, tumor invasion serves as an initial step in the complex process leading to cancer progression and metastasis<sup>[96]</sup>. EMT, which modifies the morphological and functional behaviors of cancer cells to resemble mesenchymal cell types, is a key prerequisite for the invasion seen in many cancer types<sup>[97,98]</sup>, including pancreatic cancer<sup>[99]</sup>. A growing body of research has demonstrated the causal link between chemotherapeutic stimuli and EMT in various cancer types, such as colon<sup>[100,101]</sup>, ovarian<sup>[102]</sup>, breast<sup>[103-107]</sup>, and skin squamous cell carcinoma<sup>[108]</sup>. These studies have given rise to a concept that chemoresistance and the tumor invasion-metastasis cascade are interrelated processes that accelerate cancer progression<sup>[109-112]</sup>. In line with this concept and the paradox of treatment-induced metastasis, suggesting that nearly all cancer treatments can inadvertently trigger and facilitate metastatic spread<sup>[113,114]</sup>, several studies have reported an association of resistance to gemcitabine with EMT and invasion in pancreatic cancer<sup>[49,52,53,92,93,95,115,116]</sup> [Table 1, Figure 1].

Regardless of therapy resistance, mounting evidence has shown a strong association between the pro-invasive phenotype (represented by EMT) and CSCs in many cancer types<sup>[117-120]</sup>, including pancreatic cancer<sup>[121]</sup>, because of shared distinct biological mechanisms. Consistent with this connection, many of the studies referenced in Table 1 have reported a simultaneous association of resistance to gemcitabine with both pro-invasive and cancer stemness phenotypes<sup>[52,53,92,93,95]</sup>. This highlights a three-way (or triangular) interaction among chemoresistance, tumor invasion, and cancer stemness in pancreatic cancer [Figure 1], as previously suggested<sup>[122,123]</sup>. We have established gemcitabine-sensitive pancreatic cancer BxPC-3 derivative

**Table 1. Tumor stemness and pro-invasive properties of PDAC cells surviving the insult from continuous or repeated exposure to gemcitabine**

	<b>Mechanisms for gemcitabine-induced phenotypes</b>	<b>Ref.</b>
<b>Cancer stemness phenotype</b>	Acquisition of resistance to gemcitabine imparts stemness phenotype to the PDAC cells via the phosphorylation-mediated activation of c-Met receptor protein tyrosine kinase and the increased expression of CSC markers CD24 and CD44	[53]
	PDAC cells surviving gemcitabine treatment express the CSC markers: CD44, CD24, Oct4, and CD133, and an EMT marker slug	[92]
	Gemcitabine-resistant PDAC cells acquire cancer stemness and EMT phenotypes mediated by the NF-κB pathway	[93]
	Gemcitabine treatment promotes chemoresistance and cancer stemness through the NOx/ROS/NF-κB/STAT3 signaling cascade	[94]
	Gemcitabine-resistant PDAC cells show the activation of c-Met receptor protein tyrosine kinase and the increased expression of CSC markers CD24, CD44, and ESA	[95]
<b>Pro-invasive phenotype</b>	Gemcitabine-resistant PDAC cells show spindle shape with pseudopodia, increased expression of vimentin, and decreased expression of E-cadherin	[53]
	Gemcitabine-resistant PDAC cells acquire EMT phenotype by activation of the Notch signaling pathway	[49]
	Gemcitabine-resistant PDAC cells show the gene expression profile responsible for EMT phenotype	[115]
	PDAC cells surviving gemcitabine treatment express the CSC markers: CD44, CD24, Oct4, and CD133, and an EMT marker slug	[92]
	Gemcitabine-resistant PDAC cells acquire cancer stemness and EMT phenotypes mediated by the NF-κB pathway	[93]
	Gemcitabine-resistant PDAC cells showed increased expression of ZEB1	[52]
	Suppression of EMT leads to an increase in PDAC cell proliferation with enhanced expression of NPs in tumors, contributing to enhanced sensitivity to gemcitabine and increased survival of mouse models of PDAC with deletion of snail or twist	[11]
	Gemcitabine-resistant PDAC cells show spindle shape with pseudopodia, migratory and invasive capacity, increased vimentin and decreased E-cadherin expression, and nuclear localization of β-catenin	[95]

PDAC: Pancreatic ductal adenocarcinoma; CSC: cancer stem cell; CD: cluster of differentiation; Oct4: octamer-binding transcription factor 4; EMT: epithelial-mesenchymal transition; NF-κB: nuclear factor-κB; NOx: nitrogen oxides; ROS: reactive oxygen species; STAT3: signal transducer and activator of transcription 3; ESA: epithelial-specific antigen; ZEB1: zinc-finger-enhancer binding protein 1; NPs: nucleoside transporters.



**Figure 1.** The mechanistic interconnection of chemoresistance, tumor invasion, and cancer stemness presents in chemoresistant pancreatic cancer. Mechanisms responsible for the respective connections are described in Table 1. NF-κB: Nuclear factor-κB; ZEB1: zinc-finger-enhancer binding protein 1; EVA1: epithelial V-like antigen 1; MAL2: myelin and lymphocyte protein 2; NPs: nucleoside transporters; CD: cluster of differentiation; Oct4: octamer-binding transcription factor 4; NOx: nitrogen oxides; ROS: reactive oxygen species; STAT3: signal transducer and activator of transcription 3; ESA: epithelial-specific antigen; PDAC: pancreatic ductal adenocarcinoma.

cell clones that gained stepwise resistance to gemcitabine (BxG30, BxG140, and BxG400 in increasing order of resistance)<sup>[124]</sup>. We have recently observed an increased invasive capacity with the formation of characteristic cellular surface microstructures (lamellipodia<sup>[125]</sup> and invadopodia<sup>[126]</sup>), and sphere-forming

ability in the resistant cell clones compared to their parental BxPC-3 cells [Figure 2A]. Moreover, we have observed severe local tumor invasion and metastasis to the liver and peritoneum in mice orthotopically transplanted with the most resistant BxG400 cells [Figure 2B], which models the refractory pancreatic cancer patients developing resistance to gemcitabine. Our preliminary observations reaffirm the interdependency between these malignant properties in pancreatic cancer cells acquiring resistance to gemcitabine.

## OUTLINE OF GLYCOGEN SYNTHASE KINASE 3 $\beta$ BIOLOGY AND ITS INVOLVEMENT IN DISEASES

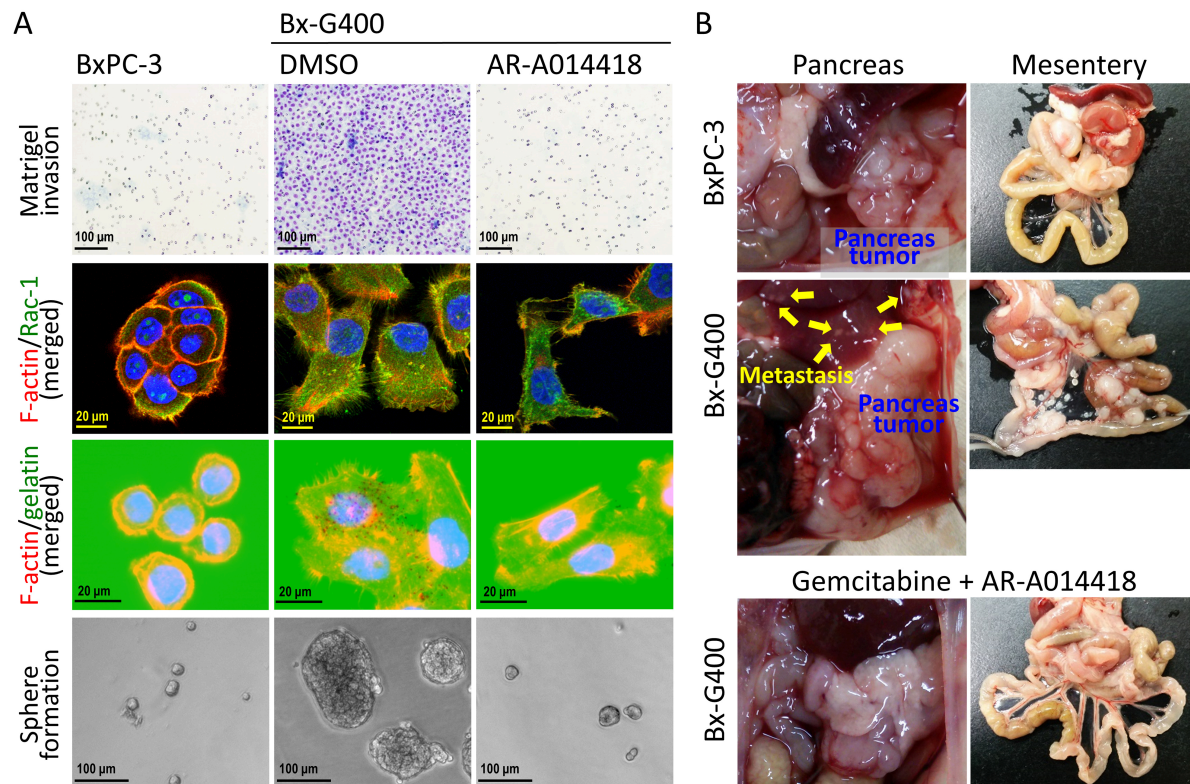
Glycogen synthase kinase (GSK)3 $\beta$  is an isoform of the GSK3 family of serine (S)/threonine (T) protein kinases. GSK3 $\beta$  is involved in a multitude of biological processes and pathways in the complex molecular networks of cells, where it interacts with nearly 100 or more structural and functional proteins via phosphorylation<sup>[127,128]</sup>. The enzymatic activity of GSK3 $\beta$  is regulated by the differential phosphorylation of its S9 (inactive form) and tyrosine (Y)216 (active form) residues. Even though GSK3 $\beta$  is typically active in cells, inhibitory regulation of its activity primarily benefits normal cells in maintaining their vital activity and homeostasis in response to various extracellular and intracellular stimuli<sup>[127-129]</sup>. Due to the redundancy in cellular expression and functions between GSK3 $\beta$  and its isoform GSK3 $\alpha$ , the pathological roles of GSK3 $\beta$  have garnered increased attention. Aberrant activity and expression of GSK3 $\beta$ , as well as defects in its inhibitory regulation, contribute to the pathogenesis and progression of common diseases, including type 2 diabetes mellitus, neurodegenerative diseases (e.g., Alzheimer's disease), inflammatory and immunological diseases, and cancer<sup>[130,131]</sup>. Given its counteracting functions in normal cells and diseases, GSK3 $\beta$  is considered a potential therapeutic target in major health disorders, thereby driving the identification and development of pharmacological GSK3 $\beta$  inhibitors<sup>[132-136]</sup>.

The biochemistry, biology, and functions of GSK3 family kinases (GSK3 $\alpha$  and GSK3 $\beta$ ) in normal cells, as well as their involvement in a wide range of common diseases, have increasingly attracted scientific attention in biomedical and pharmacological fields. This topic has been extensively reviewed in previous literature<sup>[127-136]</sup>, and is therefore only briefly outlined here.

## TUMOR-PROMOTING ROLES OF GSK3 $\beta$ IN PANCREATIC CANCER

Contrary to its pathological roles in diseases other than cancer, activated GSK3 $\beta$  counters pro-oncogenic pathways such as those mediated by Wnt/ $\beta$ -catenin, Hedgehog, and Notch signaling and transcription factors (e.g., snail) that induce EMT in normal cells. Given its functions in non-transformed cells, activation of GSK3 $\beta$  has long been hypothesized to suppress the development and progression of cancer<sup>[137-139]</sup>, thereby posing a challenge to pharmaceutical industries and clinical oncologists aiming to develop and apply GSK3 $\beta$  inhibitors for cancer treatment. However, no direct evidence supports the tumor-suppressor role of this kinase, nor the effect of GSK3 $\beta$  inhibition on promoting cancer development and progression. Contrary to this hypothesis, a significant amount of research conducted by our group and others over the last two decades has provided solid evidence demonstrating the tumor-promoting roles of active GSK3 $\beta$  as well as therapeutic effects related to its inhibition in more than 25 different cancer types (reviewed in<sup>[140-146]</sup>), including pancreatic cancer (reviewed in<sup>[147-152]</sup>). As a result, GSK3 $\beta$  has emerged as a potential therapeutic target in cancer, encouraging the development of GSK3 $\beta$  inhibitors for cancer treatment<sup>[153-155]</sup>. Similar to other cancer types, accumulating evidence for pancreatic cancer<sup>[156-182]</sup> shows that deregulated GSK3 $\beta$  supports tumor cell survival, immortality, and proliferation by mediating distinct pathways. As discussed below, GSK3 $\beta$  also facilitates the invasion of tumor cells and makes them unresponsive or resistant to chemotherapy and ionizing radiation [Figure 3].

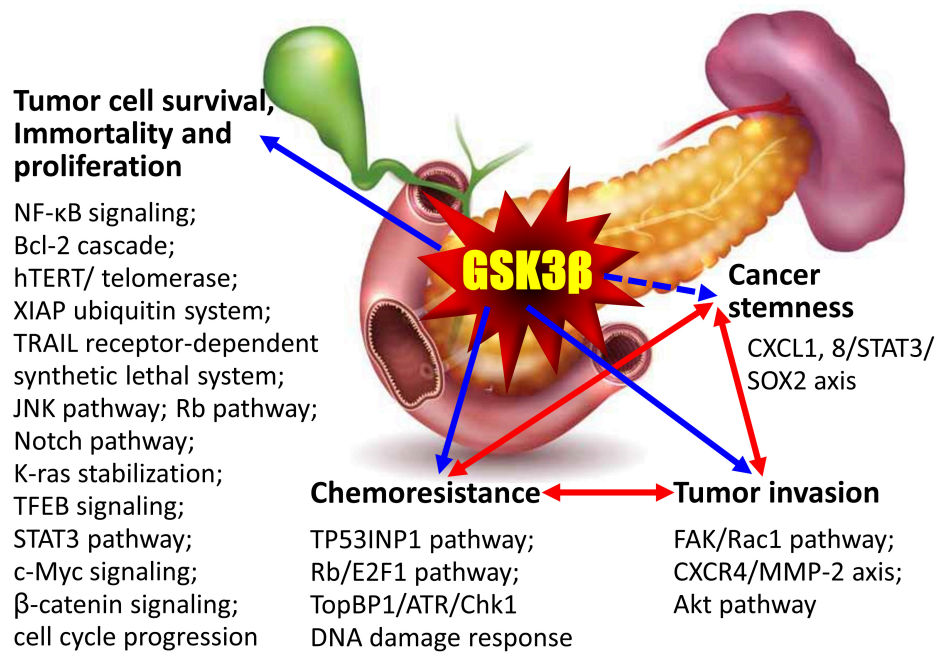




**Figure 2.** (A) Representative findings of matrigel invasion (upper panels), formation of lamellipodia and invadopodia [middle six panels: nuclei were counterstained with DAPI (blue fluorescence), and sphere formation (lower panels)] of BxPC-3 cells and BxPC-3 cell-derived gemcitabine-resistant Bx-G400 cells that were treated with DMSO (diluent of the inhibitor) and GSK3 $\beta$  inhibitor, AR-A014418; (B) Laparotomy findings of the mice with intrapancreatic transplantation of BxPC-3 and Bx-G400 cells, respectively. The bottom panels show the mice treated with gemcitabine and AR-A014418 in combination for 8 weeks after transplantation. DAPI: 4',6-diamidino-2-phenylindole; DMSO: dimethyl sulfoxide; GSK3 $\beta$ : glycogen synthase kinase 3 $\beta$ .

## GSK3 $\beta$ AS A MOLECULAR HUB IN MECHANISTICALLY WIRING THE CHEMORESISTANCE, TUMOR INVASION AND CANCER STEMNESS

As depicted in Figure 3, most studies on pancreatic cancer have demonstrated that the deregulated GSK3 $\beta$  sustains tumor cell survival, immortalization, and proliferation - common and fundamental features that engender therapy resistance in nearly all cancer types. This is achieved by enhancing cell immortality and several pro-oncogenic pathways [e.g., Nuclear factor- $\kappa$ B (NF- $\kappa$ B), Notch, K-ras, c-Myc], and by abrogating distinct tumor suppressor pathways (e.g., Rb) [Figure 3]. Our group and others have previously reported that GSK3 $\beta$  contributes to the unresponsiveness of pancreatic cancer cells to gemcitabine by impairing the DNA damage response mediated by tumor protein 53-inducible nuclear protein 1 (TP53INP1) and topoisomerase II $\beta$  binding protein 1 (TopBP1)/ataxia telangiectasia and Rad3-related (ATR)/checkpoint kinase 1 (Chk1)<sup>[165,169,178]</sup>. We have also shown that GSK3 $\beta$  plays a role in the acquisition of resistance to gemcitabine in resistant pancreatic cancer cell clones derived from BxPC-3<sup>[124]</sup> [Figure 2], via impairing the functional interaction between the Rb tumor suppressor protein and pro-oncogenic E2 transcription factor (E2F)<sub>1</sub><sup>[179]</sup>. Our previous studies on gemcitabine-unresponsive pancreatic cancer and temozolomide-resistant glioblastoma cells have indicated that GSK3 $\beta$  enhances tumor cell migration and invasion via the focal adhesion kinase (FAK), Rac1, and c-Jun N-terminal kinase (JNK)-mediated pathway<sup>[169,183]</sup>. It is conceivable that the respective mechanisms responsible for chemoresistance and tumor invasion share GSK3 $\beta$  as a common trigger for both malignant properties in gemcitabine-resistant pancreatic cancer.



**Figure 3.** Tumor-promoting properties of GSK3β and underlying mechanisms reported in pancreatic cancer<sup>[156-182]</sup>. Dotted arrow: preliminary findings (by Domoto *et al.*). The mechanisms responsible for the respective interconnections indicated by red bidirectional arrows are shown in Table 1 and Figure 1. GSK3β: Glycogen synthase kinase 3β; NF-κB: nuclear factor κB; Bcl-2: B-cell/chronic lymphocytic leukemia lymphoma 2; hTERT: human telomerase reverse transcriptase; XIAP: X-linked inhibitor of apoptosis protein; TRAIL: tumor necrosis factor-related apoptosis-inducing ligand; JNK: c-Jun N-terminal kinase; TFEB: transcription factor EB; STAT3: signal transducer and activator of transcription 3; TP53INP1: tumor protein 53-inducible nuclear protein 1; E2F1: E2 transcription factor 1; TopBP1: topoisomerase II binding protein 1; ATR: ataxia telangiectasia and Rad3-related; Chk1: checkpoint kinase 1; FAK: focal adhesion kinase; CXCR4: C-X-C chemokine receptor type 4; MMP: matrix metalloproteinase; CXCL: C-X-C chemokine ligand; SOX2: sex-determining region Y-box transcription factor 2.

As we previously reviewed<sup>[146]</sup>, a series of studies have shown that GSK3β underpins the pivotal mechanisms for sustaining CSCs and the acquisition of cancer stemness phenotype in various cancer types, including colorectal cancer, prostate cancer, head and neck squamous cell carcinoma, glioblastoma, and leukemia. We have previously shown that kenpaullone, an adenosine triphosphate (ATP)-competitive GSK3β inhibitor, reversed the temozolomide resistance of glioblastoma patient-derived tumor stem cells<sup>[184]</sup>. In a recent preliminary study, we found that GSK3β inhibition counteracts tumor invasion and distant metastasis [Figure 2B] as well as the sphere formation of BxG400 cells, which acquire the highest resistance to gemcitabine established from the gemcitabine-sensitive pancreatic cancer BxPC-3 cells [Figure 2A]. These effects were associated with the suppression of C-X-C chemokine receptor type 4 (CXCR4)-mediated matrix metalloproteinase (MMP)-2 activation, and CXC ligand (CXCL) 1 and 8-induced activation of the signal transducer and activator of transcription (STAT)3/SRY-box transcription factor (SOX)2 axis (Figure 3; preliminary observations by Domoto *et al.*). Collectively, GSK3β potentially functions as a molecular hub that wires chemoresistance, tumor invasion, and cancer stemness phenotype, thereby aggravating pancreatic cancer towards the incurable/devastating disease stage.

### PRESUMPTIVE INVOLVEMENT OF GSK3β IN THE DESMOPLASTIC TUMOR STROMA AND THE PERMISSIVE ANTI-CANCER IMMUNE ENVIRONMENT

Desmoplastic tumor stroma and permissive (or tolerant) immunity against cancer make up the hostile TME<sup>[60-62]</sup> that is recognized as a formidable obstacle for palliative treatment with chemotherapeutics, radiation, and targeted agents in pancreatic cancer<sup>[70-73]</sup>. Here, we briefly discuss the prospective involvement

of GSK3 $\beta$  in the pancreatic cancer TME.

The primary cellular components of pancreatic cancer TME include pancreatic stellate cells (PSCs)<sup>[185]</sup> and cancer-associated fibroblasts (CAFs)<sup>[60,186,187]</sup>. Both stromal cell types in pancreatic cancer support tumor cells and produce dense fibrous stroma that forms a physical barrier for drug delivery, and their interaction with tumor cells mechanistically contributes to chemoresistance. PSCs, a minor and quiescent cellular population in healthy pancreas stroma, are activated by extracellular stimulants including tumor necrosis factor (TNF) $\alpha$ , transforming growth factor (TGF)- $\beta$ , and interleukin (IL)-1, IL-2, and IL-10 in the TME. Activated PSCs enable cancer cells to resist gemcitabine via the Notch pathway-mediated Hes1 overexpression<sup>[188]</sup>, exclusive expression of periostin in PSCs<sup>[189]</sup>, and the paracrine stromal cell-derived factor (SDF)-1 $\alpha$ -mediated activation of FAK/Akt and extracellular signal-regulated kinase (ERK)1/2, with subsequent activation of the IL-6/Janus kinase (JAK)/STAT3 pathway in the tumor cells in an autocrine manner<sup>[190]</sup>. The cellular origin of CAFs includes bone marrow-derived mesenchymal stem cells (MSCs), PSCs, and preexisting dormant fibroblasts. CAFs are reportedly activated by sonic hedgehog (SHH), TGF- $\beta$ , TNF $\alpha$ , and IL-1, IL-6, and IL-10. CAFs have been shown to support tumor cells through exosomal transfer and the paracrine signaling mediated by NF- $\kappa$ B and cytokines such as IL-6, thereby activating the downstream JAK/STAT3, mechanistic target of rapamycin (mTOR), and SHH pathways<sup>[184,186]</sup>. A previous study reported that CAFs with activation of the mTOR/4E-binding protein (BP)1 pathway and resultant secretion of IL-6 induced resistance to gemcitabine in pancreatic cancer cells<sup>[191]</sup>. While a previous study demonstrated that the GSK3 $\beta$  inhibitor BIO maintains proliferation and the undifferentiated state of immortalized pancreatic MSCs that share characteristics of bone marrow-derived MSCs and PSCs<sup>[192]</sup>, no studies have directly shown the role of GSK3 $\beta$  in PSCs or CAFs. As discussed above, the reported mechanisms by which PSCs and CAFs interact with and induce chemoresistance in pancreatic cancer cells involve the Notch, mTOR, and IL-6/JAK/STAT3 pathways. Previous reviews have described the roles of GSK3 $\beta$  in mediating Notch and mTOR signaling in different cancer types, including pancreatic cancer<sup>[193,194]</sup>. Previous studies have shown that GSK3 $\beta$  functions in the phosphorylation-dependent activation of STAT3<sup>[195,196]</sup> and that inhibition of GSK3 $\beta$  attenuates the progression of gastric and esophageal cancers by suppressing STAT3 activity<sup>[197,198]</sup>. Therefore, elucidating the conceivable contribution of GSK3 $\beta$  to the interaction of PSCs and CAFs with tumor cells may provide a new strategy for targeting the tumor-promoting stroma, thereby combatting chemoresistance in pancreatic cancer.

The permissive immune environment in pancreatic cancer is complex, primarily resulting from the failure in innate immunity exerted by natural killer (NK) T-cells and the suppression of adaptive immunity by the immune checkpoint machinery between CD8<sup>+</sup> T-cells and tumor cells, mediated by the programmed death (PD)-1/PD ligand (PD-L)1 axis and cytotoxic T-lymphocyte-associated protein (CTLA)-4. Despite substantial and excellent preclinical backing, most clinical studies have failed to prove the efficacy of antibody-based immune checkpoint blockades (reviewed in<sup>[18,19,199-202]</sup>). We<sup>[146]</sup> and a recent series of reviews<sup>[151,154,155]</sup> have detailed the reported evidence showing that GSK3 $\beta$  attenuates the ability of anti-tumor immunocellular arsenals such as NK cells, CD8<sup>+</sup> T-cell-derived pluripotent memory stem T-cells with cytotoxic capacity, and tumor-type specific genetically engineered chimeric antigen receptor (CAR)-T cells. The reported evidence also showed that GSK3 $\beta$  enhances PD-1 expression depending on the transcription factor TBX21 (Tbet) and PD-L1 expression in response to the inhibition of poly [ADP-ribose] polymerase (PARP)1, and that inhibition of GSK3 $\beta$  reverses the blockade of CD28's ability to bind and stimulate antigen-presenting immune cells by CTLA-4. Importantly, a recent study demonstrated that FAK suppresses antigen processing and presentation to promote immune evasion in pancreatic cancer<sup>[203,204]</sup>, suggesting a previously underexplored mechanistic link between desmoplastic tumor stroma and tumor cell-autonomous mechanisms of immune evasion<sup>[205-207]</sup>. If future studies provide a direct relationship



between tolerant anti-tumor immunity and the acquisition of chemoresistance, it will enhance the opportunity to combat chemoresistance in pancreatic cancer by targeting GSK3 $\beta$ .

## CONCLUSION

GSK3 $\beta$  potentially functions as a molecular hub that wires chemoresistance, tumor invasion, and cancer stemness phenotype, thereby aggravating pancreatic cancer towards the incurable/devastating disease stage.

## DECLARATIONS

### Authors' contributions

Conceptualization: Minamoto T

Literature search: Uehara M, Minamoto T

Preparation of figures: Domoto T, Minamoto T

Original drafting: Uehara M, Domoto T, Minamoto T

Review and editing: Takenaka S, Takeuchi O, Shimasaki T, Miyashita T, Minamoto T

All authors have read and agreed to the published version of the manuscript.

### Availability of data and materials

Not applicable.

### Financial support and sponsorship

This work was supported by Grants-in-Aid for Scientific Research from the Ministry of Education, Science, Sports, Technology and Culture and from the Japan Society for the Promotion of Science (to UM: 23K14644; TD: 22K07227, T. Miyashita: 23K08163; and T. Minamoto: 22H03144).

### Conflicts of interest

All authors declared that there are no conflicts of interest.

### Ethical approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Copyright

© The Author(s) 2024.

## REFERENCES

1. Kleeff J, Korc M, Apte M, et al. Pancreatic cancer. *Nat Rev Dis Primers* 2016;2:16022. [DOI PubMed](#)
2. Mizrahi JD, Surana R, Valle JW, Shroff RT. Pancreatic cancer. *Lancet* 2020;395:2008-20. [DOI PubMed](#)
3. Suker M, Beumer BR, Sadot E, et al. FOLFIRINOX for locally advanced pancreatic cancer: a systematic review and patient-level meta-analysis. *Lancet Oncol* 2016;17:801-10. [DOI PubMed PMC](#)
4. Goldstein D, El-Maraghi RH, Hammel P, et al. nab-Paclitaxel plus gemcitabine for metastatic pancreatic cancer: long-term survival from a phase III trial. *J Natl Cancer Inst* 2015;107:dju413. [DOI PubMed](#)
5. Wang-Gillam A, Li CP, Bodoky G, et al; NAPOLI-1 Study Group. Nanoliposomal irinotecan with fluorouracil and folinic acid in metastatic pancreatic cancer after previous gemcitabine-based therapy (NAPOLI-1): a global, randomised, open-label, phase 3 trial. *Lancet* 2016;387:545-57. [DOI PubMed](#)
6. Tempero MA, Malafa MP, Chiorean EG, et al. NCCN guidelines insights: pancreatic adenocarcinoma, version 1.2019. *J Natl Compr Canc Netw* 2019;17:202-10. [DOI PubMed](#)
7. Springfield C, Jäger D, Büchler MW, et al. Chemotherapy for pancreatic cancer. *Presse Med* 2019;48:e159-74. [DOI PubMed](#)
8. Christenson ES, Jaffee E, Azad NS. Current and emerging therapies for patients with advanced pancreatic ductal adenocarcinoma: a

- bright future. *Lancet Oncol* 2020;21:e135-45. DOI PubMed PMC
9. Bergman AM, Pinedo HM, Peters GJ. Determinants of resistance to 2',2'-difluorodeoxycytidine (gemcitabine). *Drug Resist Updat* 2002;5:19-33. DOI PubMed
  10. Binenbaum Y, Na'ara S, Gil Z. Gemcitabine resistance in pancreatic ductal adenocarcinoma. *Drug Resist Updat* 2015;23:55-68. DOI PubMed
  11. Zeng S, Pöttler M, Lan B, Grützmann R, Pilarsky C, Yang H. Chemoresistance in pancreatic cancer. *Int J Mol Sci* 2019;20:4504. DOI PubMed PMC
  12. Sun J, Russell CC, Scarlett CJ, McCluskey A. Small molecule inhibitors in pancreatic cancer. *RSC Med Chem* 2020;11:164-83. DOI PubMed PMC
  13. Carbone D, Pecoraro C, Panzeca G, et al. 1,3,4-Oxadiazole and 1,3,4-thiadiazole nortoposentin derivatives against pancreatic ductal adenocarcinoma: synthesis, cytotoxic activity, and inhibition of CDK1. *Mar Drugs* 2023;21:412. DOI PubMed PMC
  14. Pishvaian MJ, Blais EM, Brody JR, et al. Overall survival in patients with pancreatic cancer receiving matched therapies following molecular profiling: a retrospective analysis of the Know Your Tumor registry trial. *Lancet Oncol* 2020;21:508-18. DOI PubMed PMC
  15. Al-Share B, Hammad N, Diab M. Pancreatic adenocarcinoma: molecular drivers and the role of targeted therapy. *Cancer Metastasis Rev* 2021;40:355-71. DOI PubMed
  16. O'Kane GM, Lowery MA. Moving the needle on precision medicine in pancreatic cancer. *J Clin Oncol* 2022;40:2693-705. DOI PubMed
  17. Jiang H, Hegde S, Knolhoff BL, et al. Targeting focal adhesion kinase renders pancreatic cancers responsive to checkpoint immunotherapy. *Nat Med* 2016;22:851-60. DOI PubMed PMC
  18. Balachandran VP, Beatty GL, Dougan SK. Broadening the impact of immunotherapy to pancreatic cancer: challenges and opportunities. *Gastroenterology* 2019;156:2056-72. DOI PubMed PMC
  19. Ullman NA, Burchard PR, Dunne RF, Linehan DC. Immunologic strategies in pancreatic cancer: making cold tumors hot. *J Clin Oncol* 2022;40:2789-805. DOI PubMed PMC
  20. Hosein AN, Dougan SK, Aguirre AJ, Maitra A. Translational advances in pancreatic ductal adenocarcinoma therapy. *Nat Cancer* 2022;3:272-86. DOI PubMed
  21. Yang G, Guan W, Cao Z, et al. Integrative genomic analysis of gemcitabine resistance in pancreatic cancer by patient-derived xenograft models. *Clin Cancer Res* 2021;27:3383-96. DOI PubMed
  22. de Sousa Cavalcante L, Monteiro G. Gemcitabine: metabolism and molecular mechanisms of action, sensitivity and chemoresistance in pancreatic cancer. *Eur J Pharmacol* 2014;741:8-16. DOI PubMed
  23. Jia Y, Xie J. Promising molecular mechanisms responsible for gemcitabine resistance in cancer. *Genes Dis* 2015;2:299-306. DOI PubMed PMC
  24. Rajabpour A, Rajaei F, Teimoori-Toolabi L. Molecular alterations contributing to pancreatic cancer chemoresistance. *Pancreatol* 2017;17:310-20. DOI PubMed
  25. Adamska A, Elaskalani O, Emmanouilidi A, et al. Molecular and cellular mechanisms of chemoresistance in pancreatic cancer. *Adv Biol Regul* 2018;68:77-87. DOI PubMed
  26. Rauchwerger DR, Firby PS, Hedley DW, Moore MJ. Equilibrative-sensitive nucleoside transporter and its role in gemcitabine sensitivity. *Cancer Res* 2000;60:6075-9. PubMed
  27. Nakano Y, Tanno S, Koizumi K, et al. Gemcitabine chemoresistance and molecular markers associated with gemcitabine transport and metabolism in human pancreatic cancer cells. *Br J Cancer* 2007;96:457-63. DOI PubMed PMC
  28. Hagmann W, Jesnowski R, Löhr JM. Interdependence of gemcitabine treatment, transporter expression, and resistance in human pancreatic carcinoma cells. *Neoplasia* 2010;12:740-7. DOI PubMed PMC
  29. Hagmann W, Faissner R, Schnölzer M, Löhr M, Jesnowski R. Membrane drug transporters and chemoresistance in human pancreatic carcinoma. *Cancers* 2010;3:106-25. DOI PubMed PMC
  30. Saiki Y, Yoshino Y, Fujimura H, et al. DCK is frequently inactivated in acquired gemcitabine-resistant human cancer cells. *Biochem Biophys Res Commun* 2012;421:98-104. DOI PubMed
  31. Dash S, Ueda T, Komuro A, et al. MYC/glutamine dependency is a therapeutic vulnerability in pancreatic cancer with deoxycytidine kinase inactivation-induced gemcitabine resistance. *Mol Cancer Res* 2023;21:444-57. DOI PubMed
  32. Costantino CL, Witkiewicz AK, Kuwano Y, et al. The role of HuR in gemcitabine efficacy in pancreatic cancer: HuR up-regulates the expression of the gemcitabine metabolizing enzyme deoxycytidine kinase. *Cancer Res* 2009;69:4567-72. DOI PubMed PMC
  33. Nakahira S, Nakamori S, Tsujie M, et al. Involvement of ribonucleotide reductase M1 subunit overexpression in gemcitabine resistance of human pancreatic cancer. *Int J Cancer* 2007;120:1355-63. DOI PubMed
  34. Wang C, Zhang W, Fu M, Yang A, Huang H, Xie J. Establishment of human pancreatic cancer gemcitabine-resistant cell line with ribonucleotide reductase overexpression. *Oncol Rep* 2015;33:383-90. DOI PubMed
  35. Minami K, Shinsato Y, Yamamoto M, et al. Ribonucleotide reductase is an effective target to overcome gemcitabine resistance in gemcitabine-resistant pancreatic cancer cells with dual resistant factors. *J Pharmacol Sci* 2015;127:319-25. DOI PubMed
  36. Ng SSW, Tsao MS, Chow S, Hedley DW. Inhibition of phosphatidylinositol 3-kinase enhances gemcitabine-induced apoptosis in human pancreatic cancer cells. *Cancer Res* 2000;60:5451-5. PubMed
  37. Akada M, Crnogorac-Jurcevic T, Lattimore S, et al. Intrinsic chemoresistance to gemcitabine is associated with decreased expression

- of BNIP3 in pancreatic cancer. *Clin Cancer Res* 2005;11:3094-101. DOI PubMed
38. Duxbury MS, Ito H, Benoit E, Waseem T, Ashley SW, Whang EE. RNA interference demonstrates a novel role for integrin-linked kinase as a determinant of pancreatic adenocarcinoma cell gemcitabine chemoresistance. *Clin Cancer Res* 2005;11:3433-8. DOI PubMed PMC
  39. Bafna S, Kaur S, Momi N, Batra SK. Pancreatic cancer cells resistance to gemcitabine: the role of MUC4 mucin. *Br J Cancer* 2009;101:1155-61. DOI PubMed PMC
  40. Duxbury MS, Ito H, Benoit E, Waseem T, Ashley SW, Whang EE. A novel role for carcinoembryonic antigen-related cell adhesion molecule 6 as a determinant of gemcitabine chemoresistance in pancreatic adenocarcinoma cells. *Cancer Res* 2004;64:3987-93. DOI PubMed
  41. Giroux V, Malicet C, Barthet M, et al. p8 is a new target of gemcitabine in pancreatic cancer cells. *Clin Cancer Res* 2006;12:235-41. DOI PubMed
  42. Bhardwaj V, Tadinada SM, Lai JCK, Bhushan A. Failure of pancreatic cancer chemotherapy: consequences of drug resistance mechanisms. In: Srivastava SK, editor. *Pancreatic cancer - molecular mechanism and targets*. InTech; 2012. Available from: <https://www.intechopen.com/chapters/33494>. [Last accessed on 29 Jan 2024].
  43. Wey JS, Gray MJ, Fan F, et al. Overexpression of neuropilin-1 promotes constitutive MAPK signalling and chemoresistance in pancreatic cancer cells. *Br J Cancer* 2005;93:233-41. DOI PubMed PMC
  44. Duxbury MS, Ito H, Zinner MJ, Ashley SW, Whang EE. Inhibition of SRC tyrosine kinase impairs inherent and acquired gemcitabine resistance in human pancreatic adenocarcinoma cells. *Clin Cancer Res* 2004;10:2307-18. DOI PubMed
  45. Arlt A, Gehrz A, Mürköster S, et al. Role of NF- $\kappa$ B and Akt/PI3K in the resistance of pancreatic carcinoma cell lines against gemcitabine-induced cell death. *Oncogene* 2003;22:3243-51. DOI PubMed
  46. Kunnumakkara AB, Guha S, Krishnan S, Diagaradjane P, Gelovani J, Aggarwal BB. Curcumin potentiates antitumor activity of gemcitabine in an orthotopic model of pancreatic cancer through suppression of proliferation, angiogenesis, and inhibition of nuclear factor- $\kappa$ B-regulated gene products. *Cancer Res* 2007;67:3853-61. DOI PubMed
  47. Pan X, Arumugam T, Yamamoto T, et al. Nuclear factor- $\kappa$ B p65/reI $\alpha$  silencing induces apoptosis and increases gemcitabine effectiveness in a subset of pancreatic cancer cells. *Clin Cancer Res* 2008;14:8143-51. DOI PubMed PMC
  48. Singh S, Srivastava SK, Bhardwaj A, Owen LB, Singh AP. CXCL12-CXCR4 signalling axis confers gemcitabine resistance to pancreatic cancer cells: a novel target for therapy. *Br J Cancer* 2010;103:1671-9. DOI PubMed PMC
  49. Wang Z, Li Y, Kong D, et al. Acquisition of epithelial-mesenchymal transition phenotype of gemcitabine-resistant pancreatic cancer cells is linked with activation of the notch signaling pathway. *Cancer Res* 2009;69:2400-7. DOI PubMed PMC
  50. Yao J, Qian C. Inhibition of Notch3 enhances sensitivity to gemcitabine in pancreatic cancer through an inactivation of PI3K/Akt-dependent pathway. *Med Oncol* 2010;27:1017-22. DOI PubMed
  51. Yao J, An Y, Wie JS, et al. Cycloamine reverts acquired chemoresistance and down-regulates cancer stem cell markers in pancreatic cancer cell lines. *Swiss Med Wkly* 2011;141:w13208. DOI PubMed
  52. Meidhof S, Brabletz S, Lehmann W, et al. ZEB1-associated drug resistance in cancer cells is reversed by the class I HDAC inhibitor mocetinostat. *EMBO Mol Med* 2015;7:831-47. DOI PubMed PMC
  53. Shah AN, Summy JM, Zhang J, Park SI, Parikh NU, Gallick GE. Development and characterization of gemcitabine-resistant pancreatic tumor cells. *Ann Surg Oncol* 2007;14:3629-37. DOI PubMed
  54. Provenzano PP, Hingorani SR. Hyaluronan, fluid pressure, and stromal resistance in pancreas cancer. *Br J Cancer* 2013;108:1-8. DOI PubMed PMC
  55. Hamada S, Masamune A, Shimosegawa T. Inflammation and pancreatic cancer: disease promoter and new therapeutic target. *J Gastroenterol* 2014;49:605-17. DOI PubMed
  56. Bijlsma MF, van Laarhoven HWM. The conflicting roles of tumor stroma in pancreatic cancer and their contribution to the failure of clinical trials: a systematic review and critical appraisal. *Cancer Metastasis Rev* 2015;34:97-114. DOI PubMed
  57. Neesse A, Algül H, Tuveson DA, Gress TM. Stromal biology and therapy in pancreatic cancer: a changing paradigm. *Gut* 2015;64:1476-84. DOI PubMed
  58. Rath N, Olson MF. Regulation of pancreatic cancer aggressiveness by stromal stiffening. *Nat Med* 2016;22:462-3. DOI PubMed
  59. DuFort CC, DelGiorno KE, Hingorani SR. Mounting pressure in the microenvironment: fluids, solids, and cells in pancreatic ductal adenocarcinoma. *Gastroenterology* 2016;150:1545-57.e2. DOI PubMed PMC
  60. Whittle MC, Hingorani SR. Fibroblasts in pancreatic ductal adenocarcinoma: biological mechanisms and therapeutic targets. *Gastroenterology* 2019;156:2085-96. DOI PubMed PMC
  61. Herting CJ, Karpovsky I, Lesinski GB. The tumor microenvironment in pancreatic ductal adenocarcinoma: current perspectives and future directions. *Cancer Metastasis Rev* 2021;40:675-89. DOI PubMed
  62. Hingorani SR. Epithelial and stromal co-evolution and complicity in pancreatic cancer. *Nat Rev Cancer* 2023;23:57-77. DOI PubMed PMC
  63. Geller LT, Barzily-Rokni M, Danino T, et al. Potential role of intratumor bacteria in mediating tumor resistance to the chemotherapeutic drug gemcitabine. *Science* 2017;357:1156-60. DOI PubMed PMC
  64. Saiki Y, Hirota S, Horii A. Attempts to remodel the pathways of gemcitabine metabolism: recent approaches to overcoming tumours with acquired chemoresistance. *Cancer Drug Resist* 2020;3:819-31. DOI PubMed PMC
  65. Santofimia-Castaño P, Iovanna J. Combating pancreatic cancer chemoresistance by triggering multiple cell death pathways.

- Pancreatology* 2021;21:522-9. DOI PubMed
66. Garber K. Stromal depletion goes on trial in pancreatic cancer. *J Natl Cancer Inst* 2010;102:448-50. DOI PubMed
  67. Erkan M, Hausmann S, Michalski CW, et al. The role of stroma in pancreatic cancer: diagnostic and therapeutic implications. *Nat Rev Gastroenterol Hepatol* 2012;9:454-67. DOI PubMed
  68. Provenzano PP, Cuevas C, Chang AE, Goel VK, Von Hoff DD, Hingorani SR. Enzymatic targeting of the stroma ablates physical barriers to treatment of pancreatic ductal adenocarcinoma. *Cancer Cell* 2012;21:418-29. DOI PubMed PMC
  69. Vennin C, Murphy KJ, Morton JP, Cox TR, Pajic M, Timpson P. Reshaping the tumor stroma for treatment of pancreatic cancer. *Gastroenterology* 2018;154:820-38. DOI PubMed
  70. Neesse A, Bauer CA, Öhlund D, et al. Stromal biology and therapy in pancreatic cancer: ready for clinical translation? *Gut* 2019;68:159-71. DOI PubMed
  71. Huang H, Brekken RA. The next wave of stroma-targeting therapy in pancreatic cancer. *Cancer Res* 2019;79:328-30. DOI PubMed
  72. Hosein AN, Brekken RA, Maitra A. Pancreatic cancer stroma: an update on therapeutic targeting strategies. *Nat Rev Gastroenterol Hepatol* 2020;17:487-505. DOI PubMed PMC
  73. Carpenter ES, Steele NG, Pasca di Magliano M. Targeting the microenvironment to overcome gemcitabine resistance in pancreatic cancer. *Cancer Res* 2020;80:3070-1. DOI PubMed
  74. Ho WJ, Jaffee EM, Zheng L. The tumour microenvironment in pancreatic cancer - clinical challenges and opportunities. *Nat Rev Clin Oncol* 2020;17:527-40. DOI PubMed PMC
  75. Holohan C, Van Schaeybroeck S, Longley DB, Johnston PG. Cancer drug resistance: an evolving paradigm. *Nat Rev Cancer* 2013;13:714-26. DOI PubMed
  76. Vasan N, Baselga J, Hyman DM. A view on drug resistance in cancer. *Nature* 2019;575:299-309. DOI PubMed PMC
  77. Tyner JW, Haderk F, Kumaraswamy A, et al. Understanding drug sensitivity and tackling resistance in cancer. *Cancer Res* 2022;82:1448-60. DOI PubMed PMC
  78. Nguyen LV, Vanner R, Dirks P, Eaves CJ. Cancer stem cells: an evolving concept. *Nat Rev Cancer* 2012;12:133-43. DOI PubMed
  79. Batlle E, Clevers H. Cancer stem cells revisited. *Nat Med* 2017;23:1124-34. DOI PubMed
  80. Dean M, Fojo T, Bates S. Tumour stem cells and drug resistance. *Nat Rev Cancer* 2005;5:275-84. DOI PubMed
  81. Maugeri-Saccà M, Vigneri P, De Maria R. Cancer stem cells and chemosensitivity. *Clin Cancer Res* 2011;17:4942-7. DOI PubMed
  82. Steinbichler TB, Dudás J, Skvortsov S, Ganswindt U, Riechelmann H, Skvortsova II. Therapy resistance mediated by cancer stem cells. *Semin Cancer Biol* 2018;53:156-67. DOI PubMed
  83. Lytle NK, Barber AG, Reya T. Stem cell fate in cancer growth, progression and therapy resistance. *Nat Rev Cancer* 2018;18:669-80. DOI PubMed PMC
  84. Borah A, Raveendran S, Rochani A, Maekawa T, Kumar DS. Targeting self-renewal pathways in cancer stem cells: clinical implications for cancer therapy. *Oncogenesis* 2015;4:e177. DOI PubMed PMC
  85. Kuhlmann JD, Hein L, Kurth I, Wimberger P, Dubrovskaya A. Targeting cancer stem cells: promises and challenges. *Anticancer Agents Med Chem* 2016;16:38-58. DOI PubMed
  86. Clarke MF. Clinical and therapeutic implications of cancer stem cells. *N Engl J Med* 2019;380:2237-45. DOI PubMed
  87. Raj D, Aicher A, Heeschen C. Concise review: stem cells in pancreatic cancer: from concept to translation. *Stem Cells* 2015;33:2893-902. DOI PubMed
  88. Sancho P, Alcalá S, Usachov V, Hermann PC, Sainz B Jr. The ever-changing landscape of pancreatic cancer stem cells. *Pancreatology* 2016;16:489-96. DOI PubMed
  89. Hermann PC, Sainz B Jr. Pancreatic cancer stem cells: a state or an entity? *Semin Cancer Biol* 2018;53:223-31. DOI PubMed
  90. Patil K, Khan FB, Akhtar S, Ahmad A, Uddin S. The plasticity of pancreatic cancer stem cells: implications in therapeutic resistance. *Cancer Metastasis Rev* 2021;40:691-720. DOI PubMed PMC
  91. Pisco AO, Huang S. Non-genetic cancer cell plasticity and therapy-induced stemness in tumour relapse: 'What does not kill me strengthens me'. *Br J Cancer* 2015;112:1725-32. DOI PubMed PMC
  92. Quint K, Tonigold M, Di Fazio P, et al. Pancreatic cancer cells surviving gemcitabine treatment express markers of stem cell differentiation and epithelial-mesenchymal transition. *Int J Oncol* 2012;41:2093-102. DOI PubMed
  93. Zhang Y, Wei J, Wang H, et al. Epithelial mesenchymal transition correlates with CD24+CD44+ and CD133+ cells in pancreatic cancer. *Oncol Rep* 2012;27:1599-605. DOI PubMed
  94. Zhang Z, Duan Q, Zhao H, et al. Gemcitabine treatment promotes pancreatic cancer stemness through the Nox/ROS/NF-κB/STAT3 signaling cascade. *Cancer Lett* 2016;382:53-63. DOI PubMed
  95. Kuo YC, Kou HW, Hsu CP, Lo CH, Hwang TL. Identification and clinical significance of pancreatic cancer stem cells and their chemotherapeutic drug resistance. *Int J Mol Sci* 2023;24:7331. DOI PubMed PMC
  96. Welch DR, Hurst DR. Defining the hallmarks of metastasis. *Cancer Res* 2019;79:3011-27. DOI PubMed PMC
  97. Brabletz T, Kalluri R, Nieto MA, Weinberg RA. EMT in cancer. *Nat Rev Cancer* 2018;18:128-34. DOI PubMed
  98. LeBleu VS, Thiery JP. The continuing search for causality between epithelial-to-mesenchymal transition and the metastatic fitness of carcinoma cells. *Cancer Res* 2022;82:1467-9. DOI PubMed
  99. Beuran M, Negoï I, Paun S, et al. The epithelial to mesenchymal transition in pancreatic cancer: a systematic review. *Pancreatology* 2015;15:217-25. DOI PubMed
  100. Yang AD, Fan F, Camp ER, et al. Chronic oxaliplatin resistance induces epithelial-to-mesenchymal transition in colorectal cancer cell



- lines. *Clin Cancer Res* 2006;12:4147-53. DOI PubMed
101. Dallas NA, Xia L, Fan F, et al. Chemoresistant colorectal cancer cells, the cancer stem cell phenotype, and increased sensitivity to insulin-like growth factor-I receptor inhibition. *Cancer Res* 2009;69:1951-7. DOI PubMed PMC
  102. Kajiyama H, Shibata K, Terauchi M, et al. Chemoresistance to paclitaxel induces epithelial-mesenchymal transition and enhances metastatic potential for epithelial ovarian carcinoma cells. *Int J Oncol* 2007;31:277-83. PubMed
  103. Hiscox S, Morgan L, Barrow D, Dutkowskil C, Wakeling A, Nicholson RI. Tamoxifen resistance in breast cancer cells is accompanied by an enhanced motile and invasive phenotype: inhibition by gefitinib ('Iressa', ZD1839). *Clin Exp Metastasis* 2004;21:201-12. DOI PubMed
  104. Hiscox S, Jiang WG, Obermeier K, et al. Tamoxifen resistance in MCF7 cells promotes EMT-like behaviour and involves modulation of  $\beta$ -catenin phosphorylation. *Int J Cancer* 2006;118:290-301. DOI PubMed
  105. Li QQ, Xu JD, Wang WJ, et al. Twist1-mediated adriamycin-induced epithelial-mesenchymal transition relates to multidrug resistance and invasive potential in breast cancer cells. *Clin Cancer Res* 2009;15:2657-65. DOI PubMed
  106. Fischer KR, Durrans A, Lee S, et al. Epithelial-to-mesenchymal transition is not required for lung metastasis but contributes to chemoresistance. *Nature* 2015;527:472-6. DOI PubMed PMC
  107. Fatherree JP, Guarin JR, McGinn RA, Naber SP, Oudin MJ. Chemotherapy-induced collagen IV drives cancer cell motility through activation of src and focal adhesion kinase. *Cancer Res* 2022;82:2031-44. DOI PubMed PMC
  108. Debaugnies M, Rodriguez-Acebes S, Blondeau J, et al. RHOJ controls EMT-associated resistance to chemotherapy. *Nature* 2023;616:168-75. DOI PubMed PMC
  109. Alexander S, Friedl P. Cancer invasion and resistance: interconnected processes of disease progression and therapy failure. *Trends Mol Med* 2012;18:13-26. DOI PubMed
  110. van Staalduijn J, Baker D, ten Dijke P, van Dam H. Epithelial-mesenchymal-transition-inducing transcription factors: new targets for tackling chemoresistance in cancer? *Oncogene* 2018;37:6195-211. DOI PubMed
  111. Garg M. Emerging roles of epithelial-mesenchymal plasticity in invasion-metastasis cascade and therapy resistance. *Cancer Metastasis Rev* 2022;41:131-45. DOI PubMed
  112. Weiss F, Lauffenburger D, Friedl P. Towards targeting of shared mechanisms of cancer metastasis and therapy resistance. *Nat Rev Cancer* 2022;22:157-73. DOI PubMed PMC
  113. Ebos JML. Prodding the beast: assessing the impact of treatment-induced metastasis. *Cancer Res* 2015;75:3427-35. DOI PubMed
  114. Karagiannis GS, Condeelis JS, Oktay MH. Chemotherapy-induced metastasis: molecular mechanisms, clinical manifestations, therapeutic interventions. *Cancer Res* 2019;79:4567-76. DOI PubMed PMC
  115. Arumugam T, Ramachandran V, Fournier KF, et al. Epithelial to mesenchymal transition contributes to drug resistance in pancreatic cancer. *Cancer Res* 2009;69:5820-8. DOI PubMed PMC
  116. Zheng X, Carstens JL, Kim J, et al. Epithelial-to-mesenchymal transition is dispensable for metastasis but induces chemoresistance in pancreatic cancer. *Nature* 2015;527:525-30. DOI PubMed PMC
  117. Polyak K, Weinberg RA. Transitions between epithelial and mesenchymal states: acquisition of malignant and stem cell traits. *Nat Rev Cancer* 2009;9:265-73. DOI PubMed
  118. Sato R, Semba T, Saya H, Arima Y. Concise review: stem cells and epithelial-mesenchymal transition in cancer: biological implications and therapeutic targets. *Stem Cells* 2016;34:1997-2007. DOI PubMed
  119. Zheng X, Dai F, Feng L, Zou H, Feng L, Xu M. Communication between epithelial-mesenchymal plasticity and cancer stem cells: new insights into cancer progression. *Front Oncol* 2021;11:617597. DOI PubMed PMC
  120. Lambert AW, Weinberg RA. Linking EMT programmes to normal and neoplastic epithelial stem cells. *Nat Rev Cancer* 2021;21:325-38. DOI PubMed
  121. Ishiwata T. Cancer stem cells and epithelial-mesenchymal transition: novel therapeutic targets for cancer. *Pathol Int* 2016;66:601-8. DOI PubMed
  122. Singh A, Settleman J. EMT, cancer stem cells and drug resistance: an emerging axis of evil in the war on cancer. *Oncogene* 2010;29:4741-51. DOI PubMed PMC
  123. Shibue T, Weinberg RA. EMT, CSCs, and drug resistance: the mechanistic link and clinical implications. *Nat Rev Clin Oncol* 2017;14:611-29. DOI PubMed PMC
  124. Yoneyama H, Takizawa-Hashimoto A, Takeuchi O, et al. Acquired resistance to gemcitabine and cross-resistance in human pancreatic cancer clones. *Anticancer Drugs* 2015;26:90-100. DOI PubMed
  125. Machesky LM. Lamellipodia and filopodia in metastasis and invasion. *FEBS Lett* 2008;582:2102-11. DOI PubMed
  126. Paz H, Pathak N, Yang J. Invading one step at a time: the role of invadopodia in tumor metastasis. *Oncogene* 2014;33:4193-202. DOI PubMed PMC
  127. Cormier KW, Woodgett JR. Recent advances in understanding the cellular roles of GSK-3. *F1000Res* 2017;6:167. DOI PubMed PMC
  128. Patel P, Woodgett JR. Chapter eight - Glycogen synthase kinase 3: a kinase for all pathways? *Curr Top Dev Biol* 2017;123:277-302. DOI PubMed
  129. McCubrey JA, Cocco L. GSK-3 signaling in health. *Adv Biol Regul* 2017;65:1-4. DOI PubMed
  130. Beurel E, Grieco SF, Jope RS. Glycogen synthase kinase-3 (GSK3): regulation, actions, and diseases. *Pharmacol Ther* 2015;148:114-31. DOI PubMed PMC

131. Hoffmeister L, Diekmann M, Brand K, Huber R. GSK3: a kinase balancing promotion and resolution of inflammation. *Cells* 2020;9:820. [DOI](#) [PubMed](#) [PMC](#)
132. Pandey MK, DeGrado TR. Glycogen synthase kinase-3 (GSK-3)-targeted therapy and imaging. *Theranostics* 2016;6:571-93. [DOI](#) [PubMed](#) [PMC](#)
133. Khan I, Tantray MA, Alam MS, Hamid H. Natural and synthetic bioactive inhibitors of glycogen synthase kinase. *Eur J Med Chem* 2017;125:464-77. [DOI](#) [PubMed](#)
134. Palomo V, Martinez A. Glycogen synthase kinase 3 (GSK-3) inhibitors: a patent update (2014-2015). *Expert Opin Ther Pat* 2017;27:657-66. [DOI](#) [PubMed](#)
135. Saraswati AP, Ali Hussaini SM, Krishna NH, Babu BN, Kamal A. Glycogen synthase kinase-3 and its inhibitors: potential target for various therapeutic conditions. *Eur J Med Chem* 2018;144:843-58. [DOI](#) [PubMed](#)
136. Wei J, Wang J, Zhang J, Yang J, Wang G, Wang Y. Development of inhibitors targeting glycogen synthase kinase-3 $\beta$  for human diseases: strategies to improve selectivity. *Eur J Med Chem* 2022;236:114301. [DOI](#) [PubMed](#)
137. Luo J. Glycogen synthase kinase 3 $\beta$  (GSK3 $\beta$ ) in tumorigenesis and cancer chemotherapy. *Cancer Lett* 2009;273:194-200. [DOI](#) [PubMed](#) [PMC](#)
138. McCubrey JA, Davis NM, Abrams SL, et al. Diverse roles of GSK-3: tumor promoter-tumor suppressor, target in cancer therapy. *Adv Biol Regul* 2014;54:176-96. [DOI](#) [PubMed](#)
139. Tejeda-Muñoz N, Robles-Flores M. Glycogen synthase kinase 3 in Wnt signaling pathway and cancer. *IUBMB Life* 2015;67:914-22. [DOI](#) [PubMed](#)
140. Miyashita K, Nakada M, Shakoori A, et al. An emerging strategy for cancer treatment targeting aberrant glycogen synthase kinase 3 $\beta$ . *Anticancer Agents Med Chem* 2009;9:1114-22. [DOI](#) [PubMed](#)
141. McCubrey JA, Steelman LS, Bertrand FE, et al. GSK-3 as potential target for therapeutic intervention in cancer. *Oncotarget* 2014;5:2881-911. [DOI](#) [PubMed](#) [PMC](#)
142. Domoto T, Pyko IV, Furuta T, et al. Glycogen synthase kinase-3 $\beta$  is a pivotal mediator of cancer invasion and resistance to therapy. *Cancer Sci* 2016;107:1363-72. [DOI](#) [PubMed](#) [PMC](#)
143. Walz A, Ugolokov A, Chandra S, et al. Molecular pathways: revisiting glycogen synthase kinase-3 $\beta$  as a target for the treatment of cancer. *Clin Cancer Res* 2017;23:1891-7. [DOI](#) [PubMed](#) [PMC](#)
144. Nagini S, Sophia J, Mishra R. Glycogen synthase kinases: moonlighting proteins with theranostic potential in cancer. *Semin Cancer Biol* 2019;56:25-36. [DOI](#) [PubMed](#)
145. Duda P, Akula SM, Abrams SL, et al. Targeting GSK3 and associated signaling pathways involved in cancer. *Cells* 2020;9:1110. [DOI](#) [PubMed](#) [PMC](#)
146. Domoto T, Uehara M, Bolidong D, Minamoto T. Glycogen synthase kinase 3 $\beta$  in cancer biology and treatment. *Cells* 2020;9:1388. [DOI](#) [PubMed](#) [PMC](#)
147. Garcea G, Manson MM, Neal CP, et al. Glycogen synthase kinase-3 beta; a new target in pancreatic cancer? *Curr Cancer Drug Targets* 2007;7:209-15. [DOI](#) [PubMed](#)
148. Shimasaki T, Kitano A, Motoo Y, Minamoto T. Aberrant glycogen synthase kinase 3 $\beta$  in the development of pancreatic cancer. *J Carcinog* 2012;11:15. [DOI](#) [PubMed](#) [PMC](#)
149. Zhang Q, Bhojani MS, Ben-Josef E, et al. Glycogen synthase kinase 3 $\beta$  in pancreatic cancer and its implications in chemotherapy and radiation therapy. *J Carcinog Mutagen* 2013;4:147. [DOI](#) [PubMed](#) [PMC](#)
150. Pecoraro C, Faggion B, Balboni B, et al. GSK3 $\beta$  as a novel promising target to overcome chemoresistance in pancreatic cancer. *Drug Resist Updat* 2021;58:100779. [DOI](#) [PubMed](#)
151. Park R, Coveler AL, Cavalcante L, Saeed A. GSK-3 $\beta$  in pancreatic cancer: spotlight on 9-ING-41, its therapeutic potential and immune modulatory properties. *Biology* 2021;10:610. [DOI](#) [PubMed](#) [PMC](#)
152. Elmadbouh OHM, Pandol SJ, Edderkaoui M. Glycogen synthase kinase 3 $\beta$ : a true foe in pancreatic cancer. *Int J Mol Sci* 2022;23:14133. [DOI](#) [PubMed](#) [PMC](#)
153. Osolodkin DI, Palyulin VA, Zefirov NS. Glycogen synthase kinase 3 as an anticancer drug target: novel experimental findings and trends in the design of inhibitors. *Curr Pharm Des* 2013;19:665-79. [DOI](#) [PubMed](#)
154. Sahin I, Eturi A, De Souza A, et al. Glycogen synthase kinase-3 beta inhibitors as novel cancer treatments and modulators of antitumor immune responses. *Cancer Biol Ther* 2019;20:1047-56. [DOI](#) [PubMed](#) [PMC](#)
155. Augello G, Emma MR, Cusimano A, et al. The role of GSK-3 in cancer immunotherapy: GSK-3 inhibitors as a new frontier in cancer treatment. *Cells* 2020;9:1427. [DOI](#) [PubMed](#) [PMC](#)
156. Ougolkov AV, Fernandez-Zapico ME, Savoy DN, Urrutia RA, Billadeau DD. Glycogen synthase kinase-3 $\beta$  participates in nuclear factor  $\kappa$ B-mediated gene transcription and cell survival in pancreatic cancer cells. *Cancer Res* 2005;65:2076-81. [DOI](#) [PubMed](#)
157. Ougolkov AV, Fernandez-Zapico ME, Bilim VN, Smyrk TC, Chari ST, Billadeau DD. Aberrant nuclear accumulation of glycogen synthase kinase-3 $\beta$  in human pancreatic cancer: association with kinase activity and tumor dedifferentiation. *Clin Cancer Res* 2006;12:5074-81. [DOI](#) [PubMed](#) [PMC](#)
158. Mai W, Miyashita K, Shakoori A, et al. Detection of active fraction of glycogen synthase kinase 3 $\beta$  in cancer cells by nonradioisotopic in vitro kinase assay. *Oncology* 2007;71:297-305. [DOI](#) [PubMed](#)
159. Wilson W III, Baldwin AS. Maintenance of constitutive I $\kappa$ B kinase activity by glycogen synthase kinase-3 $\alpha/\beta$  in pancreatic cancer. *Cancer Res* 2008;68:8156-63. [DOI](#) [PubMed](#) [PMC](#)

160. Mai W, Kawakami K, Shakoori A, et al. Deregulated GSK3 sustains gastrointestinal cancer cells survival by modulating human telomerase reverse transcriptase and telomerase. *Clin Cancer Res* 2009;15:6810-9. DOI PubMed
161. Mamaghani S, Patel S, Hedley DW. Glycogen synthase kinase-3 inhibition disrupts nuclear factor-kappaB activity in pancreatic cancer, but fails to sensitize to gemcitabine chemotherapy. *BMC Cancer* 2009;9:132. DOI PubMed PMC
162. Gaisina IN, Gallier F, Ougolkov AV, et al. From a natural product lead to the identification of potent and selective benzofuran-3-yl-(indol-3-yl)maleimides as glycogen synthase kinase 3 $\beta$  inhibitors that suppress proliferation and survival of pancreatic cancer cells. *J Med Chem* 2009;52:1853-63. DOI PubMed PMC
163. Guzmán EA, Johnson JD, Linley PA, Gunasekera SE, Wright AE. A novel activity from an old compound: Manzamine A reduces the metastatic potential of AsPC-1 pancreatic cancer cells and sensitizes them to TRAIL-induced apoptosis. *Invest New Drugs* 2011;29:777-85. DOI PubMed PMC
164. Zhang JS, Koenig A, Harrison A, et al. Mutant K-Ras increases GSK-3 $\beta$  gene expression via an ETS-p300 transcriptional complex in pancreatic cancer. *Oncogene* 2011;30:3705-15. DOI PubMed PMC
165. Shimasaki T, Ishigaki Y, Nakamura Y, et al. Glycogen synthase kinase 3 $\beta$  inhibition sensitizes pancreatic cancer cells to gemcitabine. *J Gastroenterol* 2012;47:321-33. DOI PubMed
166. Marchand B, Tremblay I, Cagnol S, Boucher MJ. Inhibition of glycogen synthase kinase-3 activity triggers an apoptotic response in pancreatic cancer cells through JNK-dependent mechanisms. *Carcinogenesis* 2012;33:529-37. DOI PubMed
167. Zhou W, Wang L, Gou SM, et al. ShRNA silencing glycogen synthase kinase-3 beta inhibits tumor growth and angiogenesis in pancreatic cancer. *Cancer Lett* 2012;316:178-86. DOI PubMed
168. Mamaghani S, Simpson CD, Cao PM, et al. Glycogen synthase kinase-3 inhibition sensitizes pancreatic cancer cells to TRAIL-induced apoptosis. *PLoS One* 2012;7:e41102. DOI PubMed PMC
169. Kitano A, Shimasaki T, Chikano Y, et al. Aberrant glycogen synthase kinase 3 $\beta$  is involved in pancreatic cancer cell invasion and resistance to therapy. *PLoS One* 2013;8:e55289. DOI PubMed PMC
170. Zhang JS, Herreros-Villanueva M, Koenig A, et al. Differential activity of GSK-3 isoforms regulates NF- $\kappa$ B and TRAIL- or TNF $\alpha$  induced apoptosis in pancreatic cancer cells. *Cell Death Dis* 2014;5:e1142. DOI PubMed PMC
171. Ying X, Jing L, Ma S, et al. GSK3 $\beta$  mediates pancreatic cancer cell invasion in vitro via the CXCR4/MMP-2 pathway. *Cancer Cell Int* 2015;15:70. DOI PubMed PMC
172. Kunnimalaiyaan S, Gamblin TC, Kunnimalaiyaan M. Glycogen synthase kinase-3 inhibitor AR-A014418 suppresses pancreatic cancer cell growth via inhibition of GSK-3-mediated Notch1 expression. *HPB* 2015;17:770-6. DOI PubMed PMC
173. Ma S, Li Q, Pan F. CXCR4 promotes GSK3 $\beta$  expression in pancreatic cancer cells via the Akt pathway. *Int J Clin Oncol* 2015;20:525-30. DOI PubMed
174. Baumgart S, Chen NM, Zhang JS, et al. GSK-3 $\beta$  governs inflammation-induced NFATc2 signaling hubs to promote pancreatic cancer progression. *Mol Cancer Ther* 2016;15:491-502. DOI PubMed PMC
175. Liu B, Yang H, Pilarsky C, Weber GF. The effect of GPRC5a on the proliferation, migration ability, chemotherapy resistance, and phosphorylation of GSK-3 $\beta$  in pancreatic cancer. *Int J Mol Sci* 2018;19:1870. DOI PubMed PMC
176. Edderkaoui M, Chheda C, Soufi B, et al. An inhibitor of GSK3B and HDACs kills pancreatic cancer cells and slows pancreatic tumor growth and metastasis in mice. *Gastroenterology* 2018;155:1985-98.e5. DOI PubMed PMC
177. Kazi A, Xiang S, Yang H, et al. GSK3 suppression upregulates  $\beta$ -catenin and c-Myc to abrogate KRas-dependent tumors. *Nat Commun* 2018;9:5154. DOI PubMed PMC
178. Ding L, Madamsetty VS, Kiers S, et al. Glycogen synthase kinase-3 inhibition sensitizes pancreatic cancer cells to chemotherapy by abrogating the TopBP1/ATR-mediated DNA damage response. *Clin Cancer Res* 2019;25:6452-62. DOI PubMed PMC
179. Uehara M, Domoto T, Takenaka S, et al. Glycogen synthase kinase-3 $\beta$  participates in acquired resistance to gemcitabine in pancreatic cancer. *Cancer Sci* 2020;111:4405-16. DOI PubMed PMC
180. Carbone D, Parrino B, Cascioferro S, et al. 1,2,4-Oxadiazole topsentin analogs with antiproliferative activity against pancreatic cancer cells, targeting GSK3 $\beta$  kinase. *ChemMedChem* 2021;16:537-54. DOI PubMed
181. Abrams SL, Akula SM, Meher AK, et al. GSK-3 $\beta$  can regulate the sensitivity of MIA-PaCa-2 pancreatic and MCF-7 breast cancer cells to chemotherapeutic drugs, targeted therapeutics and nutraceuticals. *Cells* 2021;10:816. DOI PubMed PMC
182. Palanivel C, Chaudhary N, Seshacharyulu P, et al. The GSK3 kinase and LZTR1 protein regulate the stability of Ras family proteins and the proliferation of pancreatic cancer cells. *Neoplasia* 2022;25:28-40. DOI PubMed PMC
183. Chikano Y, Domoto T, Furuta T, et al. Glycogen synthase kinase 3 $\beta$  sustains invasion of glioblastoma via the focal adhesion kinase, Rac1, and c-Jun N-terminal kinase-mediated pathway. *Mol Cancer Ther* 2015;14:564-74. DOI PubMed
184. Kitabayashi T, Dong Y, Furuta T, et al. Identification of GSK3 $\beta$  inhibitor kenpaullone as a temozolomide enhancer against glioblastoma. *Sci Rep* 2019;9:10049. DOI PubMed PMC
185. Pothula SP, Xu Z, Goldstein D, Pirola RC, Wilson JS, Apte MV. Key role of pancreatic stellate cells in pancreatic cancer. *Cancer Lett* 2016;381:194-200. DOI PubMed
186. von Ahrens D, Bhagat TD, Nagrath D, Maitra A, Verma A. The role of stromal cancer-associated fibroblasts in pancreatic cancer. *J Hematol Oncol* 2017;10:76. DOI PubMed PMC
187. Helms E, Onate MK, Sherman MH. Fibroblast heterogeneity in the pancreatic tumor microenvironment. *Cancer Discov* 2020;10:648-56. DOI PubMed PMC
188. Cao F, Li J, Sun H, Liu S, Cui Y, Li F. HES 1 is essential for chemoresistance induced by stellate cells and is associated with poor

- prognosis in pancreatic cancer. *Oncol Rep* 2015;33:1883-9. DOI PubMed
189. Liu Y, Li F, Gao F, et al. Periostin promotes the chemotherapy resistance to gemcitabine in pancreatic cancer. *Tumour Biol* 2016;37:15283-91. DOI PubMed
  190. Zhang H, Wu H, Guan J, et al. Paracrine SDF-1 $\alpha$  signaling mediates the effects of PSCs on GEM chemoresistance through an IL-6 autocrine loop in pancreatic cancer cells. *Oncotarget* 2015;6:3085-97. DOI PubMed PMC
  191. Duluc C, Moatassim-Billah S, Chalabi-Dchar M, et al. Pharmacological targeting of the protein synthesis mTOR/4E-BP1 pathway in cancer-associated fibroblasts abrogates pancreatic tumour chemoresistance. *EMBO Mol Med* 2015;7:735-53. DOI PubMed PMC
  192. Cao H, Chu Y, Lv X, et al. GSK3 inhibitor-BIO regulates proliferation of immortalized pancreatic mesenchymal stem cells (iPMSCs). *PLoS One* 2012;7:e31502. DOI PubMed PMC
  193. Bertrand FE. The cross-talk of NOTCH and GSK-3 signaling in colon and other cancers. *Biochim Biophys Acta Mol Cell Res* 2020;1867:118738. DOI PubMed
  194. Evangelisti C, Chiarini F, Paganelli F, Marmiroli S, Martelli AM. Crosstalks of GSK3 signaling with the mTOR network and effects on targeted therapy of cancer. *Biochim Biophys Acta Mol Cell Res* 2020;1867:118635. DOI PubMed
  195. Beurel E, Jope RS. Differential regulation of STAT family members by glycogen synthase kinase-3. *J Biol Chem* 2008;283:21934-44. DOI PubMed PMC
  196. Beurel E, Jope RS. Lipopolysaccharide-induced interleukin-6 production is controlled by glycogen synthase kinase-3 and STAT3 in the brain. *J Neuroinflammation* 2009;6:9. DOI PubMed PMC
  197. Yoon J, Ko YS, Cho SJ, et al. Signal transducers and activators of transcription 3-induced metastatic potential in gastric cancer cells is enhanced by glycogen synthase kinase-3 $\beta$ . *APMIS* 2015;123:373-82. DOI PubMed
  198. Gao S, Li S, Duan X, et al. Inhibition of glycogen synthase kinase 3 beta (GSK3 $\beta$ ) suppresses the progression of esophageal squamous cell carcinoma by modifying STAT3 activity. *Mol Carcinog* 2017;56:2301-16. DOI PubMed PMC
  199. Sahin IH, Askan G, Hu ZI, O'Reilly EM. Immunotherapy in pancreatic ductal adenocarcinoma: an emerging entity? *Ann Oncol* 2017;28:2950-61. DOI PubMed PMC
  200. Heumann T, Azad N. Correction to: Next-generation immunotherapy for pancreatic ductal adenocarcinoma: navigating pathways of immune resistance. *Cancer Metastasis Rev* 2021;40:863-4. DOI PubMed
  201. Hester R, Mazur PK, McAllister F. Immunotherapy in pancreatic adenocarcinoma: beyond “copy/paste”. *Clin Cancer Res* 2021;27:6287-97. DOI PubMed PMC
  202. Bockorny B, Grossman JE, Hidalgo M. Facts and hopes in immunotherapy of pancreatic cancer. *Clin Cancer Res* 2022;28:4606-17. DOI PubMed
  203. Canel M, Sławińska AD, Lonergan DW, et al. FAK suppresses antigen processing and presentation to promote immune evasion in pancreatic cancer. *Gut* 2023;73:131-55. DOI PubMed PMC
  204. Blanco-Gomez A, Jorgensen C. FAK scaffolds immune escape in pancreatic cancer. *Gut* 2023;73:6-8. DOI PubMed
  205. Turley SJ, Cremasco V, Astarita JL. Immunological hallmarks of stromal cells in the tumour microenvironment. *Nat Rev Immunol* 2015;15:669-82. DOI PubMed
  206. Barrett RL, Puré E. Cancer-associated fibroblasts and their influence on tumor immunity and immunotherapy. *Elife* 2020;9:e57243. DOI PubMed PMC
  207. Kennel KB, Bozlar M, De Valk AF, Greten FR. Cancer-associated fibroblasts in inflammation and antitumor immunity. *Clin Cancer Res* 2023;29:1009-16. DOI PubMed PMC