

Notch signaling in hepatocellular carcinoma: molecular targeting in an advanced disease

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ABSTRACT

The use of alternative therapeutic approaches in advanced carcinogenesis is a growing investigative base. One such cancer, primary liver cancer, is one of the most commonly occurring cancers worldwide and often presents in late stage disease consequently preventing traditional curative modalities. As a result, hepatocellular carcinoma (HCC), representing the majority of primary liver cancer, is the third most common cause of cancer-related deaths globally. Survival rates are linked to stage of presentation as well as concomitant cirrhosis limiting the 5-year survival in these patients to < 20%. Alternative strategies are in dire need as patients in this cohort have limited palliative options. Currently, sorafenib is the only approved systemic therapy; however, it has a limited survival advantage and low efficacy prompting the empirical need for further evaluation. Understanding of cancer therapy has led to an enhanced focus on the Notch pathway as a potential target for advanced HCC. Notch signaling is a critical component of development and cell fate and has been linked to various modalities including liver regeneration and as a key driver in carcinogenesis. In this review, we will provide a review of the current status of the Notch signaling in liver cancer and of Notch as an alternative potential strategy for advanced HCC.

Key words: Hepatocellular carcinoma; liver regeneration; Notch inhibitors; Notch signaling; sorafenib

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INTRODUCTION

Hepatocellular carcinoma (HCC) is a significant health concern representing the sixth most common cancer globally.^[1] Over the past 20 years, HCC has become one of the most frequent occurring tumors worldwide with the incidence in the United States steadily increasing.^[2-5] In addition, coupled with an increase in the incidence, HCC mortality has also increased substantially. Currently, it is the third most common cause of cancer-related deaths throughout the world.

Approximately one-third of patients are amendable to curative therapy through the use of localized radiofrequency ablation or resection.^[6,7] Moderate stage disease indicative of multifocal intrahepatic carcinogenesis has led to alternative approaches such as trans-arterial chemoembolization (TACE). TACE has provided a relatively efficacious avenue for patients in this category.^[7-10] Patients progressing to or presenting as late stage disease have limited treatment options. Approximately, 70% of patients will initially or eventually present at this late stage. Consequently, this leads to a 5-year survival in patients with HCC of < 20%.^[11] In addition, HCC is characteristically coupled with concomitant cirrhosis, further exacerbating disease morbidity and mortality.^[12] Therefore, there is an urgent and critical need to expand alternative and effective approaches to these patients in advanced, nonresectable disease. This need for additional therapy and the evolving understanding of molecular pathways has led to a concentrated focus on therapeutic molecular targeting in many organ-specific cancers as well as HCC.

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Several signaling pathways are of interest due to their specific oncogenic and/or tumor suppressor characteristics. For instance, the ras/raf pathway provides the only current approved therapeutic approach in advanced HCC through the use of sorafenib.^[13,14] Other traditional pathways such as PI3k/Akt/mammalian target of rapamycin,^[15-17] Wnt/ β -catenin,^[18] as well as Notch signaling^[19-21] have been investigated. Further delineation into the manipulation of these pathways is critical for future alternative strategies for HCC. One of these pathways of interest is Notch signaling. As a functionally conserved pathway, it is involved in the regulation of several cellular properties including differentiation, proliferation, homeostasis and survival. First studied in *Drosophila*, Notch was linked to neural development. Future studies were able to identify the homology between species as well as accurately describe the Notch transmembrane receptor and provide evidence in its role as cellular regulator of differentiation, proliferation, and survival.^[22-28] Continued work in the field of Notch signaling would inevitably showcase its role in a myriad of cellular processes centered on the development.

NOTCH SIGNALING

The Notch signaling pathway consists of Notch receptors, ligands, negative and positive modifiers, and transcription factors. In mammals, these efficient modules have several members and the interplay between these molecules is not yet fully understood, but its role in several processes is being teased out including regulation of metabolism, inflammation, liver regeneration and repair.^[29] Notch signaling is important from other conserved signaling pathways because its role in the mechanism of signal transduction is crucial. Compare to other intercellular signaling pathways such as Wnt, Hedgehog, and transforming growth factor- β , Notch is distinctive in several traits. First, the signaling of Notch is unique. It is comprised of both canonical and noncanonical signaling. The traditional canonical pathway occurs in a juxtacrine process that is unique to Notch. Cell-to-cell interaction is required for subsequent signaling. A transmitter cell releases one of the five major Notch ligands (Jagged 1, 2, and Delta-like 1, 3, 4) and binds to one of the four transmembrane Notch receptors (Notch 1-4) on the associate cell. Signaling is through several cleavage steps. The Notch receptor is cleaved by furin-like convertases in the trans-Golgi network, which results in two subunits of the mature/functional receptor. The extracellular Notch receptor subunit consists of a ligand-binding domain that is composed of epidermal growth factor-like repeats.^[30] In addition, mammalian Notch-1, -2 and -3 receptors contain cytokine response regions and transcriptional activation domains.^[30] Successful binding of the ligand to the representative receptor triggers a

cleavage cascade of the Notch intracellular domain (NICD) via γ -secretase with NICD translocated to the nucleus.^[31] NICD then binds with the RBP-Jk family activating the complex as well as recruiting co-activator MAML1 that initiates transcription of Notch downstream targets including hairy enhancer of split, hairy enhancer of split with YRPW motif families, p21, and Sox-9.^[32-36] Deactivation of the Notch signal is rapidly induced by phosphorylation and degradation. NICD is phosphorylated within the PEST domain by the CDK8 kinase and targeted for proteasomal degradation by E3 ubiquitin ligases that include Sel10/Fbw7.^[37,38] Transcription activation of the ternary complex is disassembled and reset for the next round of signaling. With no second messenger to amplify its signal, deactivation is acute and tightly regulated.

Noncanonical Notch signaling involves a multiple of parallel pathways as cross-talk between these pathways dominates the influence of Notch through paracrine regulation.^[31,39] One particular, well documented, example is the signaling of Notch and the Wnt/ β -catenin pathways. Both pathways can act in synergistic concert through traditional Notch signaling or opposing interactions.^[40] Conversely, antagonistic signaling is through noncanonical effects. A second crucial aspect particular to Notch signaling is the counteracting effects. Notch signals involve either the promotion or suppression of cell proliferation, cell death, and activation of differentiation programs. This happens in cells throughout development of the organism and during the maintenance of self-renewing adult tissues. Therefore, gain or loss of Notch signaling mechanisms has been directly linked to multiple human disorders. Even more conflicting in nature, opposing actions of Notch has been linked to similar disease processes in the liver.^[41,42]

NOTCH SIGNALING IN THE LIVER

The role of Notch signaling in the liver remained relatively unknown until the discovery and investigation of Alagille syndrome (AGS). As an autosomal dominant disease, AGS is characterized by ductopenia and cholestasis. Diminished development of intrahepatic bile ducts is the hallmark of AGS. Through genetic testing, near the turn of the century, it was demonstrated that mutations in the *Jagged 1* gene and to a lesser extent Notch-2 led to AGS.^[43-46] Therefore, it evidenced the role of Notch signaling in hepatogenesis, more specifically hepatic duct morphogenesis. Further research into the role of Notch in liver development focused on liver regeneration following injury. During injury and subsequent liver regeneration (i.e., partial liver resection) hepatocyte and cholangiocyte proliferative properties are often inhibited; therefore, precursor hepatic progenitor cells (HPCs) are activated in response to massive liver injury.^[47-49] HPCs

can differentiate or give rise to hepatocytes as well as cholangiocytes through Notch activation thus further strengthening the role of Notch in hepatogenesis and morphogenesis.^[50-52] During repair and regeneration and at the height of concentrated HPC involvement, several pathways are activated to assist in morphogenesis including the Notch pathway. In addition, activation of the Notch pathway (specifically Notch-1 and -2 isoforms) is in concert with parallel pathways during regeneration.^[40,53,54] These critical and emerging studies linking Notch activation to intrahepatic morphogenesis was a fundamental building block to the transition to reviewing the role of Notch in carcinogenesis.

NOTCH IN CARCINOGENESIS

As the expanding role of Notch signaling in the development of organogenesis continued, the role of Notch in carcinogenesis was ongoing. Notch-1 identification as an oncogene was first discovered through investigation into T-cell acute lymphoblastic leukemia (T-ALL). T-ALL gain of function mutations in Notch-1 led to overexpression and constitutive activation of Notch-1 receptor and thus enhanced proliferation.^[55] In addition, the oncogenic ability of Notch was also exhibited in colorectal cancer.^[56] According to Ambros, most Notch-mediated processes require a transient pulse of activity that in some cases lasts only as long as a fraction of the cell cycle degradation of the NICD.^[57] This is of particular interest as Notch transduction has a 1:1 ratio of input to output without the presence of second messengers. Therefore, constitutive activation will provide constant transduction and thus aberrant proliferation. In addition to mutations in Notch-1 signaling leading to oncogenic enhancement, alterations in Notch-1 signaling also has led to changes in angiogenesis.^[58]

The enhanced discoveries of the Notch signaling pathway lead to further advancement in additional solid tumors. Robinson *et al.*^[59] studied the overexpression of Notch-1 and -2 fusion proteins in benign breast epithelial cells. Subsequent constitutive expression resulted in altered growth characteristics while the inhibition of Notch signaling reduced the growth of the *Notch* gene fusion-expressing breast cancer xenografts. Zender *et al.*^[60] showed that overexpression of the Notch signaling pathway modulates the dysregulation of the oncogene cyclin E, resulting in the development of cholangiocellular carcinoma. In addition, inhibition of Notch activity blocks tumor cell proliferation and induces apoptosis in cholangiocellular carcinoma.^[60] A retrospective analysis in oral squamous cell carcinoma (OSCC) showed that the Notch pathway was defective in 66% of patients and the studies of mechanism

showed that the functional Notch-1 signaling inhibited proliferation of OSCC cell lines.^[61]

Notch-1 dysregulation is not the only isoform involved in carcinogenesis. Studies have demonstrated that up regulation of the Notch-3 isoform was required for induction of p21 expression in senescent cells.^[62] Inactivation of Notch-3 by γ -secretase inhibitor (GSI) or short interference RNA (siRNA) decreased cell proliferation and induced apoptosis in the chemoresistant ovarian cancer cells.^[63] Finally, Lu *et al.*^[64] have showed that the Notch-3 was positively correlated with Jagged-1 at the mRNA and protein levels. Therefore, they concluded that Notch-3 and Jagged-1 may play an important role in the initiation and proliferation of human nonfunctioning adenomas.^[64]

Despite these early indications of Notch as a potential target for cancer therapy, the reality of Notch signaling in carcinogenesis remains opaque. Although there is growing evidence as to Notch acting as an oncogenic process, other cancers exhibit Notch's role as a tumor suppressor in nature. For instance, in neuroendocrine tumors, Notch-1 acts as a tumor suppressor with overexpression leading to a reduction in cellular proliferation and growth.^[26,65-67] Furthermore, despite thorough studies involving either the activation or inhibition of the Notch pathway in the modulation of carcinogenesis, there is limited data surrounding the expression of Notch receptors and their link with cancer. Additionally, noncanonical pathway activation of Notch further confuses and complicates the underlying roles of this pathway during times of aberrant cellular growth.

TARGETING NOTCH IN HEPATOCELLULAR CARCINOMA

Despite promising results of Notch mediation in multiple organ-specific cancers, there is limited and conflicting data on Notch signaling in HCC. In several studies, Notch-1 acts as a tumor suppressor.^[41,68] On the other hand, there is evidence that Notch is oncogenic in nature.^[69-71] As time passes, growing evidence may indicate that although individual HCC signatures may include Notch as a tumor suppressor, the majority of HCC Notch mediation is through overexpression and oncogenic activation. For instance, Villanueva *et al.*^[72] revealed that the conditional expression of NICD1 in a mouse model led to HCC in all test subjects within the 1st year. Biopsied tumors represented varying stages in the mice test group. Moreover, overexpression of Notch-1 was closely linked to insulin-like growth factor 2 and Sox-9 expression levels and interestingly, the NICD1 conditionally active mice genetic signature was evidenced in a subset of human patients with HCC.^[72] Further studies have shifted the tide toward Notch acting as an oncogene in liver carcinogenesis. Sox-9, a downstream target of Notch signaling cascade, is linked to cellular proliferation

and carries a worse prognosis. Overexpression of Sox-9 leads to a transition to HPC-type activity and consequently less differentiated cell types.^[73]

As we advance our understanding of the Notch signaling pathway, initial studies including several early phase I clinical trials are underway in various stepwise components within the pathway. Given the intricacy of the canonical Notch pathway, it comes as no surprise that there are multiple avenues to target Notch signaling [Figure 1]. Targeting either the ligands and/or the receptors, inhibiting cleavage of the active NICD, and preventing transcription of downstream targets are the major targeted aims in Notch mediation. In this review, we will briefly discuss the most common techniques aimed at inhibiting Notch signaling.

The most studied area is inhibition of GSI and the subsequent release of NICD. In fact, GSI examination is not inclusive to HCC. Rather, the extensive research of Notch signaling in neural development has led to GSI application for Alzheimer's disease.^[74] There has been countless preclinical and phase I clinical trials examining the efficacy and effectiveness of GSI mediation in many cancer types.^[75-77] Unfortunately, to date, there is no phase I evidence of the role of GSIs in

HCC. Moreover, given the pan-inhibition nature of GSIs, the toxicity profiles are relatively disconcerting given the off target effects, especially intestinal adverse effects specifically through down regulation of Notch-1 and -2 isoforms.^[29]

As a result of the nonspecific inhibition of GSIs, alternative strategies should be considered. One particular area of increasing interest is the use of monoclonal antibodies and decoys at both the ligand and receptor sites. Antibody and decoy (competitive antagonist) application has a more specific efficacy, thus limiting the dose-escalated toxicities and potentially providing a concentrated result. There are multiple monoclonal antibodies currently being tested in preclinical studies. Notch-1 receptor antibodies have shown promising results.^[78,79] Both Notch-2 and Notch-3 antibodies have transitioned to phase I clinical trials.^[79,80] In addition to Notch receptor blockade, antibodies against Notch signaling ligands have been investigated. Delta-like ligand four antibodies have shown interesting results from multiple avenues.^[58,81,82] Similarly, decoys provide excellent Notch inhibition and act as a competitive antagonist either at the Notch receptor or the ligand binding sites. Notch-1 decoys have been studied as well as Jagged-1 ligand decoys.^[83,84] Finally, prevention of NICD-mediated transcription is a novel process to modulate carcinogenesis. Peptides that block the transcription of NICD provide interesting applications to Notch signal inhibition.^[75] In addition to this review of potential Notch mediation through the alteration of multiple events, Espinoza and Miele^[75] recently compiled a comprehensive table and analysis including a majority of current preclinical and clinical studies using a myriad of Notch inhibitors that further details the current effectiveness of Notch alteration.

Traditional mechanisms of Notch inhibition have and will continue to be thoroughly investigated; however, there is growing interest in targeted gene inhibition, more specifically in the context of Notch signaling. Historically, AGS and the mechanism of hepatogenesis were further delineated through *Notch-2* gene manipulation; therefore, an approach to targeting specific genes within the Notch pathway may provide additional support in lieu of traditional Notch inhibitors. There are several approaches to gene silencing, two of which are frequently used and include using a small hairpin RNA or short hairpin RNA (shRNA) or siRNA. Mao *et al.*^[85] reported that the shRNA mediated knock-down of Notch-1 inhibited the breast cancer cell line MCF-7's proliferation and induced cell apoptosis through multiple mechanistic actions. One in particular, the down regulation of the anti-apoptotic protein nuclear factor-kappa B, proved effective and enhanced the anti-tumorigenic effect when combined with traditional chemotherapeutic agents such

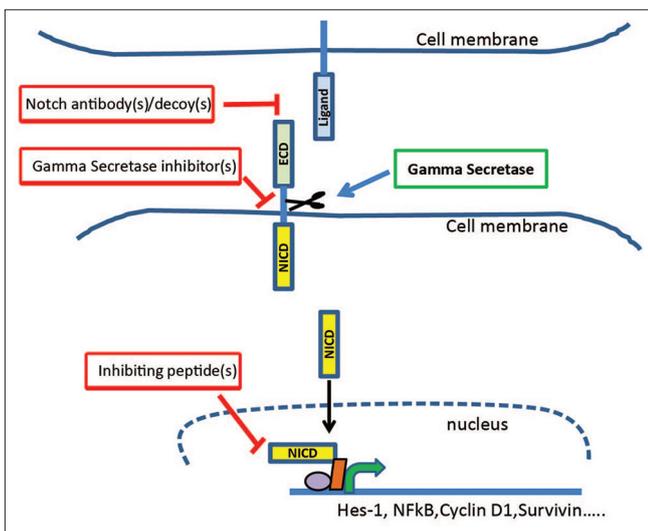


Figure 1: Canonical Notch signaling pathway and potential sites of inhibition. The Notch pathway is primed through cell-to-cell interaction distinguishing it from other regulatory pathways. Following ligand secretion from a transmitting cell, the ligand binds to one of the four Notch receptors on the receiver cell. Ligand-receptor binding facilitates the cleavage of the intracellular component of the transmembrane receptor via γ -secretase. Successful cleavage activates the Notch intracellular component domain (NICD) which translocates to the nucleus where is regulated a host of transcription factors. Given the intricacy of the pathway, there are multiple key regulatory steps poised for targeted therapy. First, monoclonal antibody and decoy administration at both the Notch ligand and receptor is in preliminary, preclinical investigation. Second, γ -secretase inhibitors are the most studied target within the pathway; however, there is limited data within hepatocellular carcinoma. Finally, inactivating transcriptional peptides are a novel trend focused on inhibiting the canonical transcription mediated by NICD

as paclitaxel. Finally, Mao *et al.*^[85] were able to translate this to an *in vivo* study evidencing that genetic knockout of Notch-1 abrogated tumor xenograft growth. Zhao *et al.*^[86] reported that the knockdown of Notch-1 by RNA interference suppressed Akt activation, reduced glioma cell growth rate and induce cell apoptosis.

Notch-1 deletion has also been studied in HCC. Sun and colleagues investigated that knockout of Notch-1 inhibited cell proliferation and significantly suppressed tumor formation of LO2/HBx cells in a BALB/c nude mouse model *in vivo* through activation of apoptotic caspase cascades. In addition, they observed that this blockade arrest the cell cycle in the G0/G1 phase through the down regulation of cyclin D1, CDK4, E2F1 and the up regulation of p21.^[87] Wang *et al.*^[88] suggested that the inhibition of Notch1 by shRNA significantly suppressed the growth of HBx transformed human hepatic cells through G0/G1 cell cycle arrest and apoptosis. The mechanism, they suggested, may be linked to the promoted expression of P16 and decreased expression of Bcl-2.^[88]

Finally, the investigation into microRNAs as a potential strategy is growing in interest. MicroRNAs are small regulators of both post-translational and post-transcriptional markers. They are often at the center of abrogation in many cancer types.^[89,90] Given their stability, they are potential candidates for use in combination studies. For instance, there is increasing data on the use of microRNAs sensitizing HCC to traditional chemotherapy.^[91,92] In addition, genetic profiling of microRNAs in patients with HCC will assist as an alternative and supportive strategy in terms of disease-free progression and overall survival. For example, microRNA-224 expression is associated with a better prognosis and further evaluation into this subtype is currently ongoing.^[93,94]

Despite the plethora of early investigations into Notch inhibition, there are concerns that need to be addressed moving forward. First, there are relatively limited studies advancing in HCC research. As of this publication, there are no clinical trials utilizing Notch inhibition as an alternative strategy for HCC. In addition, in other solid tumor studies, no trial has advanced past phase I. Perhaps we are in the early stages of development, but given the stagnant advancement additional approaches should be addressed.

TRANSITION TO THE FUTURE

The exciting and staggering concept of Notch signaling is that it is still in the infantile stages of development. The majority of evolution in the understanding of this pathway has come within the last 20-30 years. Within that time, there have been

novel and potentially ground-breaking investigations into the role of Notch not only in HCC, but other cancer types. In addition, the study of Notch mediation has radiated toward different fields of medicine with the intent of delineating the roles of isoform-specific NICD. Additionally, the role of cellular homeostasis has interpretive results in a myriad of clinical and basic science indications and perhaps Notch will be at the forefront of these studies. However, the role of Notch in carcinogenesis, albeit, counterintuitive, is both exciting and complex. The early results prove modulation of this pathway could aid in the care of advanced, resistant, and aggressive cancer types. These Notch-based strategies will continue to be evaluated and will also be combined with other pathway mitigation to reduce toxicity profiles, as well as the chemoresistance. Combination with approved and current strategies will further the understanding and commitment to providing alternative and efficacious treatment options to patients with HCC.

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REFERENCES

1. European Association For The Study Of The Liver, European Organisation For Research And Treatment Of Cancer. EASL-EORTC clinical practice guidelines: management of hepatocellular carcinoma. *J Hepatol* 2012;56:908-43.
2. Altekruse SF, Henley SJ, Cucinelli JE, McGlynn KA. Changing hepatocellular carcinoma incidence and liver cancer mortality rates in the United States. *Am J Gastroenterol* 2014;109:542-53.
3. Llovet JM, Burroughs A, Bruix J. Hepatocellular carcinoma. *Lancet* 2003;362:1907-17.
4. Schütte K, Bornschein J, Malfertheiner P. Hepatocellular carcinoma – epidemiological trends and risk factors. *Dig Dis* 2009;27:80-92.
5. Siegel R, Ma J, Zou Z, Jemal A. Cancer statistics, 2014. *CA Cancer J Clin* 2014;64:9-29.
6. McGlynn KA, Tsao L, Hsing AW, Devesa SS, Fraumeni JF Jr. International trends and patterns of primary liver cancer. *Int J Cancer* 2001;94:290-6.
7. Llovet JM, Bruix J. Systematic review of randomized trials for unresectable hepatocellular carcinoma: chemoembolization improves survival. *Hepatology* 2003;37:429-42.
8. Bruix J, Sherman M. Practice Guidelines Committee, American Association for the Study of Liver Diseases. Management of hepatocellular carcinoma. *Hepatology* 2005;42:1208-36.
9. Sergio A, Cristofori C, Cardin R, Pivetta G, Ragazzi R, Baldan A, Girardi L, Cillo U, Burra P, Giacomini A, Farinati F. Transcatheter arterial chemoembolization (TACE) in hepatocellular carcinoma (HCC): the role of angiogenesis and invasiveness. *Am J Gastroenterol* 2008;103:914-21.
10. Miura JT, Gamblin TC. Transarterial chemoembolization for primary liver malignancies and colorectal liver metastasis. *Surg Oncol Clin N Am* 2015;24:149-66.

11. Pons F, Varela M, Llovet JM. Staging systems in hepatocellular carcinoma. *HPB (Oxford)* 2005;7:35-41.
12. McGlynn KA, London WT. The global epidemiology of hepatocellular carcinoma: present and future. *Clin Liver Dis* 2011;15:223-43, vii.
13. Llovet JM, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, de Oliveira AC, Santoro A, Raoul JL, Forner A, Schwartz M, Porta C, Zeuzem S, Bolondi L, Greten TF, Galle PR, Seitz JF, Borbath I, Haussinger D, Giannaris T, Shan M, Moscovici M, Voliotis D, Bruix J; SHARP Investigators Study Group. Sorafenib in advanced hepatocellular carcinoma. *N Engl J Med* 2008;359:378-90.
14. Villanueva A, Llovet JM. Targeted therapies for hepatocellular carcinoma. *Gastroenterology* 2011;140:1410-26.
15. Nakanishi K, Sakamoto M, Yamasaki S, Todo S, Hirohashi S. Akt phosphorylation is a risk factor for early disease recurrence and poor prognosis in hepatocellular carcinoma. *Cancer* 2005;103:307-12.
16. Vanhaesebroeck B, Stephens L, Hawkins P. PI3K signalling: the path to discovery and understanding. *Nat Rev Mol Cell Biol* 2012;13:195-203.
17. Vivanco I, Sawyers CL. The phosphatidylinositol 3-Kinase AKT pathway in human cancer. *Nat Rev Cancer* 2002;2:489-501.
18. Lachenmayer A, Alsinet C, Savic R, Cabellos L, Toffanin S, Hoshida Y, Villanueva A, Minguez B, Newell P, Tsai HW, Barretina J, Thung S, Ward SC, Bruix J, Mazzaferro V, Schwartz M, Friedman SL, Llovet JM. Wnt-pathway activation in two molecular classes of hepatocellular carcinoma and experimental modulation by sorafenib. *Clin Cancer Res* 2012;18:4997-5007.
19. Ranganathan P, Weaver KL, Capobianco AJ. Notch signalling in solid tumours: a little bit of everything but not all the time. *Nat Rev Cancer* 2011;11:338-51.
20. Razumilava N, Gores GJ. Notch-driven carcinogenesis: the merging of hepatocellular cancer and cholangiocarcinoma into a common molecular liver cancer subtype. *J Hepatol* 2013;58:1244-5.
21. Rose SL, Kunnimalaiyaan M, Drenzek J, Seiler N. Notch 1 signaling is active in ovarian cancer. *Gynecol Oncol* 2010;117:130-3.
22. Austin J, Kimble J. glp-1 is required in the germ line for regulation of the decision between mitosis and meiosis in *C. elegans*. *Cell* 1987;51:589-99.
23. Cabrera CV. The generation of cell diversity during early neurogenesis in *Drosophila*. *Development* 1992;115:893-901.
24. Coffman C, Harris W, Kintner C. Xotch, the *Xenopus* homolog of *Drosophila* notch. *Science* 1990;249:1438-41.
25. Ellisen LW, Bird J, West DC, Soreng AL, Reynolds TC, Smith SD, Sklar J. TAN-1, the human homolog of the *Drosophila* notch gene, is broken by chromosomal translocations in T lymphoblastic neoplasms. *Cell* 1991;66:649-61.
26. Kunnimalaiyaan M, Chen H. Tumor suppressor role of Notch-1 signaling in neuroendocrine tumors. *Oncologist* 2007;12:535-42.
27. Priess JR, Schnabel H, Schnabel R. The glp-1 locus and cellular interactions in early *C. elegans* embryos. *Cell* 1987;51:601-11.
28. Yamamoto S, Schulze KL, Bellen HJ. Introduction to Notch signaling. *Methods Mol Biol* 2014;1187:1-14.
29. Morell CM, Strazzabosco M. Notch signaling and new therapeutic options in liver disease. *J Hepatol* 2014;60:885-90.
30. Mumm JS, Kopan R. Notch signaling: from the outside in. *Dev Biol* 2000;228:151-65.
31. Andersson ER, Lendahl U. Therapeutic modulation of Notch signalling – are we there yet? *Nat Rev Drug Discov* 2014;13:357-78.
32. Allenspach EJ, Maillard I, Aster JC, Pear WS. Notch signaling in cancer. *Cancer Biol Ther* 2002;1:466-76.
33. Fortini ME. Notch signaling: the core pathway and its posttranslational regulation. *Dev Cell* 2009;16:633-47.
34. Geisler F, Strazzabosco M. Emerging roles of Notch signaling in liver disease. *Hepatology* 2015;61:382-92.
35. Gridley T. Notch signaling and inherited disease syndromes. *Hum Mol Genet* 2003;12:R9-13.
36. Strazzabosco M, Fabris L. Notch signaling in hepatocellular carcinoma: guilty in association! *Gastroenterology* 2012;143:1430-4.
37. O'Neil J, Grim J, Strack P, Rao S, Tibbitts D, Winter C, Hardwick J, Welcker M, Meijerink JP, Pieters R, Draetta G, Sears R, Clurman BE, Look AT. FBW7 mutations in leukemic cells mediate NOTCH pathway activation and resistance to gamma-secretase inhibitors. *J Exp Med* 2007;204:1813-24.
38. Thompson BJ, Buonamici S, Sulis ML, Palomero T, Vilimas T, Basso G, Ferrando A, Aifantis I. The SCFFBW7 ubiquitin ligase complex as a tumor suppressor in T cell leukemia. *J Exp Med* 2007;204:1825-35.
39. D'Souza B, Meloty-Kapella L, Weinmaster G. Canonical and non-canonical Notch ligands. *Curr Top Dev Biol* 2010;92:73-129.
40. Boulter L, Govaere O, Bird TG, Radulescu S, Ramachandran P, Pellicoro A, Ridgway RA, Seo SS, Spee B, Van Rooijen N, Sansom OJ, Iredale JP, Lowell S, Roskams T, Forbes SJ. Macrophage-derived Wnt opposes Notch signaling to specify hepatic progenitor cell fate in chronic liver disease. *Nat Med* 2012;18:572-9.
41. Viatour P, Ehmer U, Saddic LA, Dorrell C, Andersen JB, Lin C, Zmoos AF, Mazur PK, Schaffer BE, Ostermeier A, Vogel H, Sylvester KG, Thorgeirsson SS, Grompe M, Sage J. Notch signaling inhibits hepatocellular carcinoma following inactivation of the RB pathway. *J Exp Med* 2011;208:1963-76.
42. Wu G, Wilson G, George J, Qiao L. Modulation of Notch signaling as a therapeutic approach for liver cancer. *Curr Gene Ther* 2015;15:171-81.
43. Oda T, Elkahloun AG, Pike BL, Okajima K, Krantz ID, Genin A, Piccoli DA, Meltzer PS, Spinner NB, Collins FS, Chandrasekharappa SC. Mutations in the human Jagged1 gene are responsible for Alagille syndrome. *Nat Genet* 1997;16:235-42.
44. Li PH, Shu SG, Yang CH, Lo FC, Wen MC, Chi CS. Alagille syndrome with interstitial 20p deletion derived from maternal ins (7;20). *Am J Med Genet* 1996;63:537-41.
45. Li L, Krantz ID, Deng Y, Genin A, Banta AB, Collins CC, Qi M, Trask BJ, Kuo WL, Cochran J, Costa T, Pierpont ME, Rand EB, Piccoli DA, Hood L, Spinner NB. Alagille syndrome is caused by mutations in human Jagged1, which encodes a ligand for Notch1. *Nat Genet* 1997;16:243-51.
46. McDaniel R, Warthen DM, Sanchez-Lara PA, Pai A, Krantz ID, Piccoli DA, Spinner NB. NOTCH2 mutations cause Alagille syndrome, a heterogeneous disorder of the notch signaling pathway. *Am J Hum Genet* 2006;79:169-73.
47. Fausto N, Campbell JS, Riehle KJ. Liver regeneration. *J Hepatol* 2012;57:692-4.
48. Riehle KJ, Dan YY, Campbell JS, Fausto N. New concepts in liver regeneration. *J Gastroenterol Hepatol* 2011;26 Suppl 1:203-12.
49. Michalopoulos GK. Liver regeneration: alternative epithelial pathways. *Int J Biochem Cell Biol* 2011;43:173-9.
50. Fiorotto R, Raizner A, Morell CM, Torsello B, Scirpo R, Fabris L, Spirli C, Strazzabosco M. Notch signaling regulates tubular morphogenesis during repair from biliary damage in mice. *J Hepatol* 2013;59:124-30.
51. Morell CM, Fiorotto R, Fabris L, Strazzabosco M. Notch signalling beyond liver development: emerging concepts in liver repair and oncogenesis. *Clin Res Hepatol Gastroenterol* 2013;37:447-54.
52. Tanimizu N, Miyajima A. Notch signaling controls hepatoblast differentiation by altering the expression of liver-enriched transcription factors. *J Cell Sci* 2004;117:3165-74.
53. Chen Y, Choi SS, Michelotti GA, Chan IS, Swiderska-Syn M, Karaca GF, Xie G, Moylan CA, Garibaldi F, Premont R, Suliman HB, Piantadosi CA, Diehl AM. Hedgehog controls hepatic stellate cell fate by regulating metabolism. *Gastroenterology* 2012;143:1319-29.e1.
54. Xie G, Karaca G, Swiderska-Syn M, Michelotti GA, Krüger L, Chen Y, Premont RT, Choi SS, Diehl AM. Cross-talk between Notch and Hedgehog regulates hepatic stellate cell fate in mice. *Hepatology* 2013;58:1801-13.

55. Weng AP, Ferrando AA, Lee W, Morris JP 4th, Silverman LB, Sanchez-Irizarry C, Blacklow SC, Look AT, Aster JC. Activating mutations of NOTCH1 in human T cell acute lymphoblastic leukemia. *Science* 2004;306:269-71.
56. van Es JH, van Gijn ME, Riccio O, van den Born M, Vooijs M, Begthel H, Cozijnsen M, Robine S, Winton DJ, Radtke F, Clevers H. Notch/gamma-secretase inhibition turns proliferative cells in intestinal crypts and adenomas into goblet cells. *Nature* 2005;435:959-63.
57. Ambros V. Cell cycle-dependent sequencing of cell fate decisions in *Caenorhabditis elegans* vulva precursor cells. *Development* 1999;126:1947-56.
58. Ridgway J, Zhang G, Wu Y, Stawicki S, Liang WC, Chanthery Y, Kowalski J, Watts RJ, Callahan C, Kasman I, Singh M, Chien M, Tan C, Hongo JA, de Sauvage F, Plowman G, Yan M. Inhibition of Dll4 signalling inhibits tumour growth by deregulating angiogenesis. *Nature* 2006;444:1083-7.
59. Robinson DR, Kalyana-Sundaram S, Wu YM, Shankar S, Cao X, Ateeq B, Asangani IA, Iyer M, Maher CA, Grasso CS, Lonigro RJ, Quist M, Siddiqui J, Mehra R, Jing X, Giordano TJ, Sabel MS, Kleer CG, Palanisamy N, Natrajan R, Lambros MB, Reis-Filho JS, Kumar-Sinha C, Chinnaiyan AM. Functionally recurrent rearrangements of the MAST kinase and Notch gene families in breast cancer. *Nat Med* 2011;17:1646-51.
60. Zender S, Nischeleit I, Wuestefeld T, Sorensen I, Dauch D, Bozko P, El-Khatib M, Geffers R, Bektas H, Manns MP, Gossler A, Wilkens L, Plentz R, Zender L, Malek NP. A critical role for notch signaling in the formation of cholangiocellular carcinomas. *Cancer Cell* 2013;23:784-95.
61. Pickering CR, Zhang J, Yoo SY, Bengtsson L, Moorthy S, Neskey DM, Zhao M, Ortega Alves MV, Chang K, Drummond J, Cortez E, Xie TX, Zhang D, Chung W, Issa JP, Zweidler-McKay PA, Wu X, El-Naggar AK, Weinstein JN, Wang J, Muzny DM, Gibbs RA, Wheeler DA, Myers JN, Frederick MJ. Integrative genomic characterization of oral squamous cell carcinoma identifies frequent somatic drivers. *Cancer Discov* 2013;3:770-81.
62. Cui H, Kong Y, Xu M, Zhang H. Notch3 functions as a tumor suppressor by controlling cellular senescence. *Cancer Res* 2013;73:3451-9.
63. Rahman MT, Nakayama K, Rahman M, Katagiri H, Katagiri A, Ishibashi T, Ishikawa M, Iida K, Nakayama S, Otsuki Y, Miyazaki K. Notch3 overexpression as potential therapeutic target in advanced stage chemoresistant ovarian cancer. *Am J Clin Pathol* 2012;138:535-44.
64. Lu R, Gao H, Wang H, Cao L, Bai J, Zhang Y. Overexpression of the Notch3 receptor and its ligand Jagged1 in human clinically non-functioning pituitary adenomas. *Oncol Lett* 2013;5:845-51.
65. Greenblatt DY, Vaccaro AM, Jaskula-Sztul R, Ning L, Haymart M, Kunnimalaiyaan M, Chen H. Valproic acid activates notch-1 signaling and regulates the neuroendocrine phenotype in carcinoid cancer cells. *Oncologist* 2007;12:942-51.
66. Nakakura EK, Sriuranpong VR, Kunnimalaiyaan M, Hsiao EC, Schuebel KE, Borges MW, Jin N, Collins BJ, Nelkin BD, Chen H, Ball DW. Regulation of neuroendocrine differentiation in gastrointestinal carcinoid tumor cells by notch signaling. *J Clin Endocrinol Metab* 2005;90:4350-6.
67. Ning L, Greenblatt DY, Kunnimalaiyaan M, Chen H. Suberoyl bis-hydroxamic acid activates Notch-1 signaling and induces apoptosis in medullary thyroid carcinoma cells. *Oncologist* 2008;13:98-104.
68. Croquelois A, Blindenbacher A, Terracciano L, Wang X, Langer I, Radtke F, Heim MH. Inducible inactivation of Notch1 causes nodular regenerative hyperplasia in mice. *Hepatology* 2005;41:487-96.
69. Giovannini C, Baglioni M, Baron Toaldo M, Ventrucci C, D'Adamo S, Cipone M, Chieco P, Gramantieri L, Bolondi L. Notch3 inhibition enhances sorafenib cytotoxic efficacy by promoting GSK3b phosphorylation and p21 down-regulation in hepatocellular carcinoma. *Oncotarget* 2013;4:1618-31.
70. Giovannini C, Gramantieri L, Chieco P, Minguzzi M, Lago F, Pianetti S, Ramazzotti E, Marcu KB, Bolondi L. Selective ablation of Notch3 in HCC enhances doxorubicin's death promoting effect by a p53 dependent mechanism. *J Hepatol* 2009;50:969-79.
71. Gramantieri L, Giovannini C, Lanzi A, Chieco P, Ravaioli M, Venturi A, Grazi GL, Bolondi L. Aberrant Notch3 and Notch4 expression in human hepatocellular carcinoma. *Liver Int* 2007;27:997-1007.
72. Villanueva A, Alsinet C, Yanger K, Hoshida Y, Zong Y, Toffanin S, Rodriguez-Carunchio L, Sole M, Thung S, Stanger BZ, Llovet JM. Notch signaling is activated in human hepatocellular carcinoma and induces tumor formation in mice. *Gastroenterology* 2012;143:1660-9.e7.
73. Guo X, Xiong L, Sun T, Peng R, Zou L, Zhu H, Zhang J, Li H, Zhao J. Expression features of SOX9 associate with tumor progression and poor prognosis of hepatocellular carcinoma. *Diagn Pathol* 2012;7:44.
74. Siemers ER, Quinn JF, Kaye J, Farlow MR, Porsteinsson A, Tariot P, Zoulnouni P, Galvin JE, Holtzman DM, Knopman DS, Satterwhite J, Gonzales C, Dean RA, May PC. Effects of a gamma-secretase inhibitor in a randomized study of patients with Alzheimer disease. *Neurology* 2006;66:602-4.
75. Espinoza I, Miele L. Notch inhibitors for cancer treatment. *Pharmacol Ther* 2013;139:95-110.
76. Tammam J, Ware C, Efferson C, O'Neil J, Rao S, Qu X, Gorenstein J, Angagaw M, Kim H, Kenific C, Kunii K, Leach KJ, Nikov G, Zhao J, Dai X, Hardwick J, Scott M, Winter C, Bristow L, Elbi C, Reilly JF, Look T, Draetta G, Van der Ploeg L, Kohl NE, Strack PR, Majumder PK. Down-regulation of the Notch pathway mediated by a gamma-secretase inhibitor induces anti-tumour effects in mouse models of T-cell leukaemia. *Br J Pharmacol* 2009;158:1183-95.
77. Wei P, Walls M, Qiu M, Ding R, Denlinger RH, Wong A, Tsaparikos K, Jani JP, Hosea N, Sands M, Randolph S, Smeal T. Evaluation of selective gamma-secretase inhibitor PF-03084014 for its antitumor efficacy and gastrointestinal safety to guide optimal clinical trial design. *Mol Cancer Ther* 2010;9:1618-28.
78. Aste-Amézaga M, Zhang N, Lineberger JE, Arnold BA, Toner TJ, Gu M, Huang L, Vitelli S, Vo KT, Haytko P, Zhao JZ, Baleydiere F, L'Heureux S, Wang H, Gordon WR, Thoryk E, Andrawes MB, Tiyanont K, Stegmaier K, Roti G, Ross KN, Franlin LL, Wang H, Wang F, Chastain M, Bett AJ, Audoly LP, Aster JC, Blacklow SC, Huber HE. Characterization of Notch1 antibodies that inhibit signaling of both normal and mutated Notch1 receptors. *PLoS One* 2010;5:e9094.
79. Wu Y, Cain-Hom C, Choy L, Hagenbeek TJ, de Leon GP, Chen Y, Finkle D, Venook R, Wu X, Ridgway J, Schahin-Reed D, Dow GJ, Shelton A, Stawicki S, Watts RJ, Zhang J, Choy R, Howard P, Kadyk L, Yan M, Zha J, Callahan CA, Hymowitz SG, Siebel CW. Therapeutic antibody targeting of individual Notch receptors. *Nature* 2010;464:1052-7.
80. Li K, Li Y, Wu W, Gordon WR, Chang DW, Lu M, Scoggin S, Fu T, Vien L, Histen G, Zheng J, Martin-Hollister R, Duensing T, Singh S, Blacklow SC, Yao Z, Aster JC, Zhou BB. Modulation of Notch signaling by antibodies specific for the extracellular negative regulatory region of NOTCH3. *J Biol Chem* 2008;283:8046-54.
81. Noguera-Troise I, Daly C, Papadopoulos NJ, Coetzee S, Boland P, Gale NW, Lin HC, Yancopoulos GD, Thurston G. Blockade of Dll4 inhibits tumour growth by promoting non-productive angiogenesis. *Nature* 2006;444:1032-7.
82. Schemet JS, Jiang W, Kumar SR, Krasnoperov V, Trindade A, Bedito R, Djokovic D, Borges C, Ley EJ, Duarte A, Gill PS. Inhibition of Dll4-mediated signaling induces proliferation of immature vessels and results in poor tissue perfusion. *Blood* 2007;109:4753-60.
83. Funahashi Y, Hernandez SL, Das I, Ahn A, Huang J, Vorontchikhina M, Sharma A, Kanamaru E, Borisenko V, Desilva DM, Suzuki A, Wang X, Shawber CJ, Kandel JJ, Yamashiro DJ, Kitajewski J. A notch1 ectodomain construct inhibits endothelial notch signaling, tumor growth, and angiogenesis. *Cancer Res* 2008;68:4727-35.
84. Small D, Kovalenko D, Kacer D, Liaw L, Landriscina M, Di Serio C, Di Serio C, Prudovsky I, Maciag T. Soluble Jagged 1 represses the

- function of its transmembrane form to induce the formation of the Src-dependent chord-like phenotype. *J Biol Chem* 2001;276:32022-30.
85. Mao J, Song B, Shi Y, Wang B, Fan S, Yu X, Tang J, Li L. ShRNA targeting Notch1 sensitizes breast cancer stem cell to paclitaxel. *Int J Biochem Cell Biol* 2013;45:1064-73.
 86. Zhao N, Guo Y, Zhang M, Lin L, Zheng Z. Akt-mTOR signaling is involved in Notch-1-mediated glioma cell survival and proliferation. *Oncol Rep* 2010;23:1443-7.
 87. Sun Q, Wang R, Wang Y, Luo J, Wang P, Cheng B. Notch1 is a potential therapeutic target for the treatment of human hepatitis B virus X protein-associated hepatocellular carcinoma. *Oncol Rep* 2014;31:933-9.
 88. Wang F, Xia X, Wang J, Sun Q, Luo J, Cheng B. Notch1 signaling contributes to the oncogenic effect of HBx on human hepatic cells. *Biotechnol Lett* 2013;35:29-37.
 89. Gramantieri L, Fornari F, Callegari E, Sabbioni S, Lanza G, Croce CM, Bolondi L, Negrini M. MicroRNA involvement in hepatocellular carcinoma. *J Cell Mol Med* 2008;12:2189-204.
 90. Lujambio A, Lowe SW. The microcosmos of cancer. *Nature* 2012;482:347-55.
 91. Xu Y, Xia F, Ma L, Shan J, Shen J, Yang Z, Liu J, Cui Y, Bian X, Bie P, Qian C. MicroRNA-122 sensitizes HCC cancer cells to adriamycin and vincristine through modulating expression of MDR and inducing cell cycle arrest. *Cancer Lett* 2011;310:160-9.
 92. Bai S, Nasser MW, Wang B, Hsu SH, Datta J, Kutay H, Yadav A, Nuovo G, Kumar P, Ghoshal K. MicroRNA-122 inhibits tumorigenic properties of hepatocellular carcinoma cells and sensitizes these cells to sorafenib. *J Biol Chem* 2009;284:32015-27.
 93. Wang Y, Lee AT, Ma JZ, Wang J, Ren J, Yang Y, Tantoso E, Li KB, Ooi LL, Tan P, Lee CG. Profiling microRNA expression in hepatocellular carcinoma reveals microRNA-224 up-regulation and apoptosis inhibitor-5 as a microRNA-224-specific target. *J Biol Chem* 2008;283:13205-15.
 94. Gyöngyösi B, Végh E, Járay B, Székely E, Fassan M, Bodoky G, Schaff Z, Kiss A. Pretreatment MicroRNA Level and Outcome in Sorafenib-treated Hepatocellular Carcinoma. *J Histochem Cytochem* 2014;62:547-55.

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