

Review

Open Access



The clinical landscape of cutaneous neurofibromas in neurofibromatosis type 1

Rebecca M. Brown 

Department of Neurology, The University of Alabama at Birmingham, Birmingham, AL 35233, USA.

Correspondence to: Dr. Rebecca M. Brown, Department of Neurology, The University of Alabama at Birmingham, 1020 Faculty Office Tower, 510 20th Street South, Birmingham, AL 35233, USA. E-mail: rbrown23@uab.edu

How to cite this article: Brown RM. The clinical landscape of cutaneous neurofibromas in neurofibromatosis type 1. *Rare Dis Orphan Drugs J.* 2025;4:21. <https://dx.doi.org/10.20517/rdodj.2025.14>

Received: 3 Mar 2025 **First Decision:** 21 Apr 2025 **Revised:** 5 Jun 2025 **Accepted:** 30 Jun 2025 **Published:** 23 Jul 2025

Academic Editor: Daniel Scherman **Copy Editor:** Ping Zhang **Production Editor:** Ping Zhang

Abstract

Neurofibromatosis type 1 (NF1) is a hereditary tumor predisposition syndrome that predisposes patients to tumors derived from the neural crest cell population. One of the most prominent and well-recognized features is the proclivity for nerve sheath tumors of the skin known as cutaneous neurofibromas (CNs). These tumors are benign and have self-limited growth, but they exert a strong negative impact on patients' quality of life. The only effective treatments currently are procedural, and there are no available medications. This review addresses the cellular and molecular characteristics of cutaneous neurofibromas with a focus on identifying novel therapeutic targets that could complement existing approaches. Preclinical models, tumor evolution throughout the lifespan, genetic associations with tumor phenotype, and a brief history of interventional clinical trials are also discussed.

Keywords: Cutaneous neurofibroma, skin tumor, neurofibromatosis, NF1, peripheral nerve sheath tumor, rasopathy, neurocristopathy

INTRODUCTION

Neurofibromatosis Type 1 (NF1) is a relatively common autosomal dominant tumor-predilection syndrome and neurocristopathy affecting approximately 1 in 3,000 individuals regardless of sex or ethnicity. Its pathoetiology is dependent on the monoallelic loss of function of NF1, resulting in haploinsufficiency in every tissue of the body. Subsequent somatic loss of the remaining allele results in disease manifestations



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, sharing, adaptation, distribution and reproduction in any medium or format, for any purpose, even commercially, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.



such as peripheral nerve sheath tumors of the Schwann cell ontology, aberrant bone formation/reabsorption, and low-grade gliomas of the brain, as well as other tumors of neural crest-derived tissues^[1,2]. The protein product of *NF1*, neurofibromin, serves to positively regulate adenyl cyclase, inactivate rat sarcoma (Ras), inhibit mitosis and induce apoptosis, and inhibit cell adhesion and motility. With germline haploinsufficiency of neurofibromin, Ras signaling escalates, leading to downstream mitogen-activated protein kinase kinase (MEK) activation. Cutaneous neurofibromas (CNs) are benign tumors of the dermis that arise from biallelic loss of *NF1* in Schwann cell-like tumor cells and occur in 99% of *NF1* patients. They typically begin to develop in late childhood/early adolescence and have self-limited growth trajectories, but lifetime accumulation can ultimately involve the entire integument. CNs can remain small/barely perceptible, or they may grow to large sizes, sometimes reaching many centimeters in diameter. Typically, each patient exhibits a characteristic number and size distribution of tumors, which can range from just a few to tens of thousands. Although individual tumors eventually enter a state of senescence, their ultimate size cannot be precisely predicted.

CNs are primarily associated with physical deformity that can lead to embarrassment, a tendency to cover up the skin with bulky clothing, barriers to intimacy, and socioeconomic disparity due to fewer opportunities for client-facing jobs. CNs can be pruritic or painful and may catch on clothing or jewelry, resulting in bleeding, swelling, and irritation. *NF1* quality of life (QOL) metrics have identified that CNs play a major role in the negative impact of *NF1* on mood and social interaction^[3-5]. The scientific establishment has recognized the importance of understanding CN biology vis-à-vis other tumor types, and of identifying tolerable treatments with satisfactory outcomes for patients^[6]. This review article summarizes what is known about CNs, including appearance, tumor initiation, preclinical models, the ultrastructure of CNs, and barriers/challenges of developing further therapies.

CNS HAVE A RANGE OF APPEARANCES

The classical appearance of CNs is that of a skin-colored, mildly erythematous, or rarely hypermelanotic globular lesion with a broad base, typically measuring from a few millimeters to several centimeters in maximal cross-sectional diameter. However, there are many clinical variants [Figure 1]. Tumors may be pedunculated - resembling a ball on a chain (usually more erythematous than the surrounding unaffected skin) - sessile, flat, or globular. Small nascent or latent CNs may only be identifiable using ultrasound imaging. Some CNs present as plaque-like lesions with distinct borders but a more planar appearance, while others are diffuse or dispersed, covering larger anatomic regions and resulting in areas of thickened, irregular skin, with or without the characteristic myxoid or collagenous stroma. Subcutaneous neurofibromas typically have indistinct borders and a more violaceous coloration. On cross-sectional gross pathology, CNs appear as a typically well-delineated white rubbery nodular lesion within the dermis, with an overlying grenz zone of normal papillary dermis, often extending into the subdermis with a barbell-like contraction at the plane of surrounding skin. This can be contrasted with plexiform neurofibromas involving the cutis, which perioperatively are more likely to have a multi-nodular or band-like ultrastructure, and can be more adherent to the overlying skin. Some plexiform neurofibromas involving the skin appear as rugate elephantine hyperpigmented exophytic lesions, whereas others may be confused with a large subcutaneous neurofibroma.

Histopathologically, CNs include abnormal cutaneous nerve endings (myelinated and unmyelinated) and nerve-dissociated bipolar Schwann-like tumor cells lacking an organized basal lamina embedded in a generous supply of extracellular matrix (ECM) with the addition of hyperplastic fibroblasts. CNs can be distinguished from plexiform neurofibromas of the cutis based on the histologic presence of microscopic thickened vermian nerve bundles seen in PNs. However, this definitive feature may not be captured in the



Figure 1. Examples of cutaneous neurofibromas. Moderate to severe burden of average-sized cutaneous neurofibromas on a male with Fitzpatrick Skin Type 6 on (A) chest and abdomen, and (B) hands; (C) and (D) Moderate burden of average-sized cutaneous neurofibromas on the chest and hands, respectively, of a woman with Fitzpatrick Skin Type 1; (E) Mild case of cutaneous neurofibromas (arrow heads) on a background of excess skin freckling; (F) A moderate-severe case of cutaneous neurofibromas on the abdomen. Ruler indicates approximate size of the lesions.

sample analyzed by the pathologist; therefore, clinical acumen is important for differentiation. The nerves in CNs include both myelinated and non-myelinated axons. Compared with the surrounding skin, CNs are well-vascularized, containing small arterioles as well as arterial and venous capillaries typically consisting of a single endothelial cell layer with or without subendothelial smooth muscle cells or pericytes, surrounded by a vessel basal lamina and a thickened adventitia. The intima and media layers are also enlarged. On electron microscopy, there are corpuscle-like bodies surrounding small blood vessels or myelinated vessels, composed of lamellar Schwann and perineurial cells with long processes, fine collagen fibers, and basal lamina. A pyramidal cell type with long tripartite processes in connection with morphologically similar cells was identified on electron microscopy as a “covering cell”^[7].

These tumors contain a high percentage of infiltrating tumor-supporting cells with atypical proliferative and signaling capabilities associated with haploinsufficiency of neurofibromin from the germline mutation of NF1. Aside from spindle-shaped Schwann-like tumor cells that form the tumor stroma (immunohistochemically identified by Sox10 or S100 staining), there are plentiful fibroblasts (including endoneurial fibroblast-like cells) and perineurial-like cells, as well as endothelial cells and pericytes. There is a robust immune cell component including tumor-associated lymphocytes, dendritic cells, mast cells, and macrophages^[8-11]. Occasionally, extensive eosinophils are seen, although this is not characteristic.

Schwann-like cells do not normally proliferate within the dermis, and thus, the identification of tumor stromal cells in a cutaneous biopsy - characterized by fusiform cells irregularly distributed within the ECM on hematoxylin and eosin (H&E) staining or by immunohistochemical (IHC) detection of Schwann cell markers S100 and Sox-10 - is generally accepted as diagnostic of a tumor of the Schwann cell lineage. Beyond this classic neurofibroma histopathologic phenotype, additional more unusual morphologic variants also exist^[12]. CNs are difficult to distinguish from cutaneous schwannomas on routine H&E

staining, and the need for distinction may not be clinically apparent to the pathologist as both are benign tumors. If the patient does not yet have a genetic diagnosis and/or does not carry the typical features of NF1, two distinct tumors can be submitted for next-generation sequencing of somatic mutation analysis with attention to alterations of chromosome 22q11. If > 1 tumors have different mutations or alterations involving the NF2 gene and no identified alteration of NF1, this would indicate non-NF2-related schwannomatosis rather than NF1. Diagnosticians should carefully consider the possibility of schwannomatosis as a diagnosis for individuals who do not have an identified germline NF1 mutation and who do not fully meet diagnostic requirements for NF1, but who harbor internal nerve sheath tumors and have at least one resected skin lesion identified as a “neurofibroma” based on H&E.

THE EXTRACELLULAR MATRIX INCREASES INTERSTITIAL PRESSURE

The ECM makes up ~50% of the dry weight of the tumor, which is about half of that in skin but twice the concentration found in peripheral nerve endoneurium. The concentration of glycosaminoglycans, particularly hyaluronic acid, is up to 10 times higher in CNs than in the surrounding skin^[13]. In CNs, collagen is organized into densely packed, haphazardly splayed bundles, interspersed with intermittent elastin or reticular fibers that surround disorganized, spindle-shaped tumor cells. CNs are physically characterized by high internal tissue tension despite lacking a true capsule. This internal pressure becomes evident during resection at the skin plane, wherein the residual hypodermal tumor often protrudes or “balloons” outward through the incision, driven by the newly created pressure gradient.

CNs are mostly composed of ECM, including fibrillary collagen types (COL) I, II, III, IV, V, network collagen type VI (COLVI) and basement membrane collagen type XV, plasma-derived fibronectin, hyaluronic acid, and Laminin isotype subunit A^[14,15]. COLI and COLIII have a filamentous ultrastructure and are involved in fibrosis, and COLIII confers a promitotic cellular phenotype and may contribute to drug resistance^[16]. Cultured fibroblasts from CNs primarily produce COLI > COLIII, whereas COLIV is secreted by Schwann-like tumor cells into a basement membrane-like structure coating each cell^[13]. In normal skin, COLVI typically resides between the basement membrane and the interstitium. It is produced by Schwann cells to guide remyelination after nerve injury^[17,18] and is required for the organization of acetylcholine receptors at the neuromuscular junction^[19]. COLVI is also found abundantly in normal skin at the dermal-epidermal border, where it contributes to the near-impenetrable skin barrier to environmental exposures, as well as within skeletal muscle, the cerebral meninges, and in disease states such as glioblastoma and Alzheimer’s disease^[20,21].

ECM components are differentially produced by Schwann-derived tumor cells, perineurial cells, and tumor-associated fibroblasts that release autocrine and paracrine stromal cell-derived factor-1 (SDF1) and transforming growth factor beta (TGFβ) to increase ECM deposition^[22]. During synthesis, COLVI is first woven into a triple helical monomer and then joined into disulfide-bonded dimers and tetramers before secretion into the extracellular space, where subunits associate non-covalently to form a net-like beaded meshwork. Intracellular collagen-synthesizing and -modifying enzymes such as lysyl hydroxylases and prolyl hydroxylases are overexpressed in CNs. The abundant ECM generates a more rigid tumor microenvironment through collagen crosslinking with elastin. ECM binding also enhances focal adhesion kinase (FAK) and integrin signaling. Moreover, the macrostructure of the ECM impairs the diffusion of large molecules and leads to anisotropy and sequestering of pro-tumorigenic growth factors close to tumor cells. Within CNs, the subchains COL6A1, A2, and A3 are most abundantly expressed by CN-associated fibroblasts^[14]. COLIV is present in a patchy discontinuous pattern focused at the tumor periphery, potentially because of hypoxia or altered diffusion parameters at the tumor core. The hyper-coiled, heavily cross-linked, and chemically stable collagen components of the ECM have a half-life longer than the average

human lifespan if not actively degraded by Schwann cells and fibroblasts, blocks immune cell migration and drug diffusion, and induces pro-tumorigenic signaling.

The collagenolysis of COLVI is not yet fully understood; however, matrix metalloproteases (MMPs) 2, 7, 9, 11, and 14 are able to digest moieties^[23]. The major anabolic machinery for collagen resides within the cell cytoplasm. Collagens bind to integrin proteins at the cell membrane and a small peptide moiety is cleaved, which signals macro-pinocytosis, clathrin-mediated phagocytosis, or endocytosis of fragments released by MMPs, followed by cathepsin-mediated lysosomal digestion. COLIV exists as a heterotrimeric helix composed of six interchangeable isomeric subunits. When excreted, COLIV self-organizes into irregular hexagonal networks of tetramers or rope-like dimers that are stabilized through hydrophobic/hydrophilic interactions as well as a lock-in-key mechanism wherein conjugate beta hairpin structures plug into neighboring carboxy terminal domains.

In addition to providing structure and altering diffusion characteristics, ECM contributes to the tumor microenvironment by binding to cell surface proteins to enact diverse pro-tumorigenic signaling pathways within both stromal and tumor-associated NF1 heterozygous cells. COLVI binds to neural/glial antigen 2 (NG2), integrins, or the capillary morphogenesis gene 2 (CMG2/ANTXR2) receptor and activates the cyclic AMP response element-binding protein (CREB)/AKT/ β -catenin/T cell factor/lymphoid enhancer factor pathway^[24] (TCF-LEF) and phosphatidylinositol-3 kinase (PI3K) signaling^[20]. The C-terminal of COLVI (A3) can be proteolytically cleaved, releasing the signaling factor endotrophin, which promotes fibrosis and inflammation. COLVI ultrastructurally interacts with biglycan, fibronectin, decorin, von Willebrand factor (vWF), vWF-A domain-related protein (WARP), fibulin, heparin sulfate, and other collagens, notably COLI and COLIV. COLVI is implicated in the inhibition of apoptosis, the maintenance of a stem-like quality, the promotion of tumor growth, and the activation of autophagy^[25].

COLIV is less well-studied than COLVI in tumor biology. It has been proposed as an inducer of epithelial-mesenchymal transition, potentially heralding a poorer prognosis in certain cancers^[26]. COLIV has a known specificity for binding tumor cells and encouraging migration along the collagenous matrix. This interaction relies on integrin $\alpha_v\beta_1$ signaling at an ultrastructural binding site that includes arginine 461 in the alpha 2 subchain and asparagine 461 within the alpha 1 subchain. COLIV also binds to laminins, perlecan, and proteoglycans. Non-integrin binding mechanisms occur through cell-surface proteins CD44, discoidin domain receptors (DDR), heparin sulfate proteoglycans, glycoprotein VI, and the mannose receptor family. Injection of COLIV into the rat sciatic nerve in an *in vivo* injury model induced cellular proliferation, fibroblast immigration, and neurite extension, although it did not induce Schwann cell mitosis^[27]. COLIV may facilitate nerve regeneration, although the mechanism likely does not involve Schwann cell activation.

Hyaluronic acid and collagen deposition are associated with high tumor interstitial pressure, low oxygen tension, microvascular collapse, and hypoxia signaling in tumor cells^[28], and collagen density correlates negatively with T cell migration into tumors and positively with macrophage-dependent immunosuppression^[29]. The overall effect of this dense irregular ECM deposition is to block drug diffusion and immune cell migration, reduce glucose and oxygen supply, increase drug efflux via hypoxia inducible factor 1 α (HIF-1 α), increase survival (integrins), and reduce cell cycle arrest via FAK signaling^[20]. Through these downstream ECM-activated pathways, particularly COLVI, resistance has been documented in solid tumors to multiple targeted and chemotherapeutic agents. ECM-targeted drug co-administration may provide a means to increase the efficacy of MEK inhibitors and other targeted therapies for CNs^[30].

CNS APPEAR IN LATE CHILDHOOD AND CONTINUE TO DEVELOP THROUGHOUT LIFE

CNs typically begin to arise in late childhood or early adolescence and continue to accumulate throughout the lifetime, affecting 99% of adults with NF1^[31,32]. Their growth is self-limited; in most patients, they reach an average maximum diameter of ~5 mm, although sizes can range from a few millimeters to 10 cm or larger. Inter-individual and interfamilial phenotypes can be striking. An 8-year prospective study revealed that a group-averaged change in CN volume among adults varies in body region but is overall quite modest - on average ~3.5 mm³ per annum, with the fastest growth seen on the back and abdomen. Similarly, the average number of new neurofibromas observed was approximately one novel lesion every 4 years during the 8-year observation period, again with the highest rate of appearance on the back and abdomen^[33]. A five-year natural history study of CN accumulation is open at the National Institutes of Health, utilizing whole-body photography to track lesion development, which could provide much-needed data on predictive factors for CN initiation, growth rates, and variability (ClinicalTrials.gov ID NCT05581511).

Hormones have long been suspected to act as mitogens for cutaneous neurofibromas. This hypothesis was initially based on the observation that these tumors often appear during early adolescence, a developmental stage characterized by high circulating sex hormone levels^[34]. Numerous preclinical experiments support a potential role for hormones in CN tumorigenesis and growth. CNs express hormone receptors, and Schwann-like tumor cell proliferation *in vitro* is buttressed by adding hormones to the growth medium^[35-37]. Subjective assessments have also documented that postpartum women retrospectively perceived a faster growth rate of CNs. However, there are no compelling prospective human data suggesting that either exogenous or endogenous hormones significantly influence tumor burden or growth rate across different sexes, or among individuals receiving hormone or anti-hormone therapies. One self-report study noted that two women receiving depot contraceptives containing high levels of synthetic progesterone believed their CN growth rate increased during treatment^[38]. Therefore, while there is no scientific evidence to support avoiding hormone or anti-hormone therapies altogether, women may be advised to consider alternative forms of contraception rather than depot injections or implants, particularly given similar weak associations observed with other tumors, such as meningiomas.

CN appearance and volume increase are difficult to assess both clinically and experimentally^[39]. There can be a high degree of inter-investigator variability on manual measurements, confounded by the use of different analytic tools (different brands/types of calipers), different methods of measurement (tumor base vs. largest tumor girth), and the degree of pressure exerted by the calipers on these rubbery/soft, sometimes inflamed lesions. The moment of nascence of CNs is also difficult to pinpoint, partly because pools of atypical tumor-like Schwann cells in apparently normal skin could represent a microscopic/primordial form of CNs, but there is no means to determine their prospective potential for growing into a visible papule. Retrospective patient report is notably unreliable, and periodic inflammation or differences in lighting may furthermore alter the investigator's attention to the presence or size of a tumor.

Future endeavors should strive to identify more CN measures that are independent of human error and variability. Photography, particularly 3-D photography, together with artificial intelligence (AI) has the potential to provide a universally accepted objective measure of CN total body burden, but the analytic software is designed to detect macular lesions with pigmentary differences from surrounding skin, and has not yet been optimized to automatically detect and measure voluminous and numerous idiochromatic CNs against a background of potentially abnormal surrounding skin. Furthermore, the research community would benefit from developing a standardized calibrated caliper device that applies a low, consistent pressure to CNs to eliminate inconsistencies associated with different tool sizes, manufacturers, weights, gliding friction, and manually applied pressures. It is important to learn from patients what degree of CN

reduction is meaningful to them. Given that destructive treatments completely eliminate the tumor, resulting in flat skin, a drug therapy that only partially reduces the size of a CN may or may not be acceptable to the patient population as a whole.

THE GENETICS OF CN SEVERITY REMAINS POORLY UNDERSTOOD

There are many subclassifications of CNs based on their dermal vs. subdermal location, gross anatomy, and histopathology; however, the shared underlying mechanism for tumorigenesis is considered constant across different tumors and individuals: loss of the wild-type NF1 allele, leading to complete/near-complete deficiency of translated neurofibromin protein. Mutations affecting the NF1 ultrastructure or pathologic gene alterations directly targeting the Protein Kinase C (PKC) domain may impact CN severity, at least in certain ethnic populations^[40]. Additional congenital polymorphisms or mutations may influence overall CN burden, whereas somatically acquired genetic alterations could contribute to individual tumor heterogeneity. There are a few causative NF1 mutations known to be associated with the phenotype and number of CNs. For example, microdeletion of the NF1 gene results in an extreme disease phenotype characterized by a heavy burden of CNs^[41], while several other mutations have been identified that are associated with a reduced incidence of CNs^[42-45]. However, in the vast majority of patients, a specific pathogenic germline mutation does not adequately predict CN features. The literature surrounding this question has been unequivocal that individuals within the same family with the same NF1 alteration can exhibit disparate cutaneous manifestations. In contrast to this well-accepted discordance between genotype and phenotype, the author's clinical experience suggests that the general burden and appearance of cutaneous and subcutaneous neurofibromas within families are often similar. The generation of databases documenting quantitative features of CNs will help future research to hone in on genetic and environmental modifiers of CN clinical presentation^[46].

Multiple evidence-driven hypotheses have sought to determine the cell of origin that initiates CNs through Knudson's "Second Hit" in the remaining wild-type NF1 gene. Loss of function of the intact NF gene can occur through mutations in coding and noncoding regions as well as deletions, and are not always detectable using standard gene sequencing, which may explain why dual hits are not identified in all CNs^[47,48]. Chromosomal aberrancies resulting from deletions and amplifications are commonly identified in cutaneous neurofibromas including recurrent losses in chromosomes 1, 2q, 3p, 4p, 5q, 6q, 7q, 12q, 19p, and 20p, and gains in chromosomes 2p and 8q^[49].

LABORATORY MODELS OF CNS ARE IMPERFECT

Although innovations in pharmacotherapy for plexiform neurofibromas have greatly benefited from preclinical research, a truly anthropomorphic animal model for CNs has only recently become available. Early genetically engineered NF1 heterozygous mouse models failed to recapitulate the characteristic symptoms of NF1^[50]. Later, conditional knockout mice with biallelic NF1 deletion in specific embryonic subpopulations of neural crest-derived Schwann cells were able to reproduce certain features of NF1 but did not develop the characteristic skin tumors^[51]. Spontaneous NF1-like veterinary syndromes occur in canine and bovine animals, but not in rodents^[51]. It has been hypothesized that the many years required to develop tumors in humans cannot be modeled by a rapid cycling mammal with a relatively brief life expectancy, such as the mouse. Porcine models have been developed for multiple medical conditions, as pig physiology and lifespan more closely mimic that of humans. In 2018, genetically altered *NF1*^{+/*R1947X*} Ossabaw minipigs were developed that spontaneously developed large diffuse mass-like cutaneous neurofibromas in ~40% of pigs at 4 months of age^[52]. The same year, another group published a *NF1*^{+/*ex42del*} Yucatan mini-swine model in which 44% of animals develop cutaneous tumors of the neck between 11-17 months of age^[53]. Both models exhibited this non-discreet morphology with continuous enlargement in contrast to the nodular

self-limited growth most common with human CN, and interestingly, the findings were mostly associated with gonadally intact male pigs, suggesting some role for androgens in CN tumorigenesis within this model.

In vitro models of CNs include primary cell culture and co-culture of Schwann-like tumor cells and tumor-associated fibroblasts from human donors, genetically engineered mouse models, and minipigs. Skin-derived precursor cells can be isolated and grown in culture, and semi-immortalized cells from donated human CNs were generated by introducing human TERT and mouse CDK4 genes via gene transduction. Induced pluripotent stem cells have been derived from human and rodent tissues, and have been used to generate orthotopic xenografts in mice^[54]. Because two-dimensional culture conditions cannot replicate the complex 3-dimensional cellular and ECM organization of CNs, neurofibroma-sphere/organoid models^[55] and complex multicellular skin grafts^[56] have been utilized to better understand the tumor microenvironment and ultimately for preclinical drug testing.

Innovative research has demonstrated that the cell of origin could reside in various sources, including multipotent bulge hair follicle cells^[57], a specialized population of skin-derived precursor cells of neural crest origin^[58], boundary cap cells of the dorsal root ganglia^[59], dysplastic vascular pericytes^[7], or deviant endoneurial or cutaneous nerve axon-associated Schwann cells - potentially through misactivation of the injury-repair pathway^[60-63]. Confounding this research is the inherently plastic nature of many neural crest-derived tissues, which can be driven, either by physiological processes or experimentally, to give rise to most of the cell types found in CNs.

CN tumorigenesis and ultimate dormancy are likely multifactorial, contingent upon microenvironmental contributions from haploinsufficient tumor-associated nerve, immune, and dermal cells through the release of growth factors, chemokines, and cytokines. Pleiotropic effects from germline variants in different genes may also contribute.

THERE ARE NO APPROVED PHARMACOTHERAPIES FOR CNS

Currently, there are no FDA-approved medical treatments for CNs. While surgical resection of the exophytic component combined with evacuation of the hypodermal portion and careful suturing can achieve permanent tumor control and an aesthetically acceptable appearance^[64], this technique can only go so far. It does not address the ongoing development of new tumors nor the often substantial burden of existing tumors. Other destructive approaches include focused ultrasound^[65], CO₂ laser, radiofrequency ablation^[66], photodynamic therapy^[67,68], Er:YAG or Nd:YAG laser^[69], and electrodesiccation^[70]. The efficacy of these approaches varies depending on tumor size and number, tolerance for scarring/ pigmentation changes, and post-procedural pain. A number of topical and systemic medications have been tested, with the most promising results to date seen with MEK inhibitors^[70-72]. Most notably, NFX-179, a topical ointment applied daily for 28 days to five medium-sized CNs (5-10 mm) in 42 subjects, resulted in a $\geq 50\%$ reduction in tumor volume in 20% of tumors and patients. In addition, there was a 47% reduction in phosphorylated ERK levels measured via automated Western blot, with practically no adverse events. However, CN-related quality of life outcomes were not assessed in this Phase 2a trial. Overall, patient satisfaction regarding efficacy, pain, scarring, durability, and side effect profiles across all treatments evaluated to date leaves room for improvement.

A search of ClinicalTrials.gov at the time of writing identified eighteen clinical trials using the search term “cutaneous neurofibroma” in the indication field [Table 1]. Of these, the majority^[14] were sponsored by academic medical centers, while the remaining four were industry-sponsored (covering three drugs). Ten trials investigated novel indications/pharmacotherapies, including three drugs targeting MEK inhibition;

Table 1. Clinical trials of cutaneous neurofibroma from <https://ClinicalTrials.gov/>

Trial number	Study title	Status	Intervention	Mechanism
NCT06300502	Assessing the efficacy of repeat, monthly treatments of cutaneous neurofibromas (cNFs)	NOT YET RECRUITING	Kybella or Asclera injection with 1064 nm Nd:YAG laser or 755 nm alexandrite laser	Destructive
NCT06159166	Mirdametininib monotherapy in adults with neurofibromatosis 1 (NF1) and cNF	RECRUITING	Oral mirdametininib	MEK inhibition
NCT02728388	Photodynamic therapy for benign dermal neurofibromas- phase II	RECRUITING	Topical aminolevulinic acid	Destructive
NCT06132165	Efficacy of skin cooling in reducing pain associated with non-invasive treatments of neurofibromatosis type 1 cutaneous neurofibromas	RECRUITING	Deoxycholic acid or polidocanol injection with 1064 nm Nd:YAG laser or 755 nm alexandrite laser	Destructive
NCT05438290	DPCP to treat cutaneous neurofibromas associated with NF1	COMPLETED	Topical Diphencyprone	Immunotherapy
NCT04730583	Tolerability of device based therapies for neurofibromatosis type 1 cutaneous neurofibromas	COMPLETED	Kybella injection with 1064 nm Nd:YAG laser or 755 nm alexandrite laser	Destructive
NCT05119582	HIFU treatment of cutaneous neurofibromas in neurofibromatosis type 1: safety and efficacy	COMPLETED	TOOsonix system ONE-M device	Destructive
NCT05005845	NFX-179 topical gel treatment for adults with NF1 and cNF	COMPLETED	Topical NFX-179 gel	MEK inhibition
NCT04435665	NFX-179 topical gel treatment in adults with NF1 and cNF	COMPLETED	Topical NFX-179 Gel	MEK inhibition
NCT03090971	Use of topical liquid diclofenac following laser microporation of cutaneous neurofibromas in patients with NF1	COMPLETED	Laser microporation device and topical diclofenac sodium	Prostaglandin inhibition
NCT00865644	Topical imiquimod 5% cream for treatment of cutaneous neurofibromas in adults with neurofibromatosis 1	COMPLETED	Topical imiquimod 5% cream	Immunotherapy
NCT00657202	Ranibizumab for neurofibromas associated with neurofibromatosis 1	COMPLETED	Oral ranibizumab	VEGF inhibitor
NCT01031901	Topical rapamycin therapy to alleviate cutaneous manifestations of tuberous sclerosis complex (TSC) and NF1	COMPLETED	Topical sirolimus or rapamycin	mTOR inhibition
NCT02839720	Selumetinib in treating patients with neurofibromatosis type 1 and cutaneous neurofibroma	COMPLETED	Oral selumetinib sulfate	MEK inhibition
NCT01682811	Phase I photodynamic therapy (PDT) for benign dermal NF1	COMPLETED	Levulan injection or topical, with or without microneedling, with photodynamic therapy	Destructive
NCT06120036	Dosing and Tolerability of deoxycholic acid vs. polidocanol in the treatment of neurofibromatosis type 1 cutaneous neurofibromas	COMPLETED	Kybella injection, or Asclera injection	Destructive
NCT02332902	Everolimus for treatment of disfiguring cutaneous lesions in neurofibromatosis 1 CRAD001CUS232T	COMPLETED	Oral everolimus	mTOR inhibition
NCT00921037	First clinical study of erbium - yttrium aluminium garnet (YAG) laser vaporization of cutaneous neurofibromas	UNKNOWN	Erbium-YAG laser device	Destructive

VEGF: Vascular endothelial growth factor; mTOR: mammalian target of rapamycin.

three different Mek inhibitors tested within 4 different trials; 2 drugs targeting mTOR, two immunotherapeutic agents, one prostaglandin inhibitor, and one vascular endothelial growth factor (VEGF) inhibitor. The remaining trials explore(d) concurrent injectable drugs with laser or photodynamic therapy. Methods to enhance drug penetration into CNs included laser microporation, microneedling, and direct intratumoral injection. At the time of writing, two trials were actively recruiting and one had not yet begun enrollment. None of these approaches had demonstrated a definitive, clinically acceptable reduction in CN size or number^[73]. Several studies have evaluated patient preferences regarding treatment outcomes. Unsurprisingly, many patients indicated a willingness to take oral or topical medication over an extended

period if it could reduce the number of CNs by 30% and prevent the formation of new tumors^[74].

Emerging data from clinical trials of systemic MEK inhibitors suggest that this drug class may play a role in slowing or halting the development of new CNs and/or reducing the size of existing CNs^[75]. Special consideration must be allotted to drug development for this benign tumor, as currently approved MEK inhibitors (mirdametinib and selumetinib) are designed to treat large, functionally impactful plexiform neurofibromas, and have a side effect profile that is unlikely to be acceptable for managing cutaneous manifestations. The primary detriment to quality of life associated with CNs derives from tumor disfigurement, while over 90% of patients on currently approved MEK inhibitors additionally experience a conspicuous acneiform rash. This visible adverse effect could exacerbate societal stigmatization and likely hinder the use of MEK inhibitors for treating CNs, despite potential benefits in tumor reduction. A 2025 study was the first to unequivocally demonstrate that systemic MEK inhibition can reduce CN tumor burden, achieving a maximum response of a 29% decrease in CN volume. However, similar to other trials, adverse events such as dry skin and rash were common^[76]. It is possible that a reduced dose of MEK inhibitors, administered prophylactically or longitudinally, could mitigate these adverse events while slowing tumor growth; however, this approach has not yet been explored.

Data released in 2025 provided the most compelling evidence to date for the treatment of CNs using the topical selumetinib formulation NFX-179^[72]. At the highest dose tested, CNs exhibited approximately a 50% reduction in phosphorylated ERK levels, a recognized biomarker of MEK inhibition. Decreases in tumor volume were observed in all groups, including the control group, but the highest drug dose led to an additional ~10% average reduction in tumor volume compared with the control. Most notably, around 20% of CNs treated with the highest dose of the topical formulation exhibited a $\geq 50\%$ reduction in volume. Future generations of MEK inhibitors, as well as novel molecularly targeted therapies and topical, biologic, or injectable treatments, may further expand the arsenal of approaches to address this psychologically and socioeconomically impactful aspect of NF1. Beyond MEK inhibition, several groups are also investigating the unique transcriptional and cellular fingerprints of CNs to identify multi-target therapeutic strategies^[77,78].

DISCUSSION

People with NF1 are hugely impacted by the visual appearance and physical symptoms of CNs, but current treatment options are largely limited to destructive techniques. Surgical specialties (general surgery, plastic surgery, dermatology, or neurosurgery) do not generally have the bandwidth to dedicate extensive operating room time to removing the substantial tumor burden that affects many NF1 patients. Electrodessication, although effective against small CNs measuring 5 mm or less, cannot treat larger lesions. Moreover, it is rarely offered, given the need for general anesthesia, specialized training and equipment, and its relatively low reimbursement rate compared with other procedures. Laser therapies can be very effective, but are limited by post-procedural pain. All destructive techniques carry the risk of scarring and pigmentation changes that may ultimately offset the cosmetic benefits. Neurologists and primary care doctors can help address the need for CN management by training in tumor resection, which can be performed in a clinical setting without the need for a sterile operating room. A well-vetted training course with certification by a respected institution such as the Children's Tumor Foundation could provide the education for doctors and advanced practice providers, and a recertification course providing continuing medical education credits could ensure a durable skillset.

The human relevance of preclinical CN models has been improving over the past decade, but still requires further refinement. Porcine skin closely resembles human skin in cellular morphology, architecture, and the thickness of the four dermal layers. However, minipigs have additional cell layers in the stratum spinosum

and granulosum and a higher proportion of apocrine versus eccrine sweat glands, potentially affecting topical drug diffusion. Indeed, an *in vitro* porcine skin permeability assay of a lipophilic compound predicted to have high skin permeability revealed striking differences: no permeability in porcine skin compared with human skin^[79]. These physiologic distinctions could influence outcomes and dose determination in preclinical trials of topical medications. The skin of juvenile minipigs best replicates adult human skin^[80]; however, CNs in minipig models arise in adulthood, when their skin is less biosimilar to that of humans. Thus, these models may be more appropriate for testing systemic therapies rather than topical drugs. Further basic research is needed to understand why murine and porcine models fail to spontaneously and accurately replicate CN morphology and distribution.

ECM is the major component of CNs and can block drug delivery by increasing tumor parenchymal back pressure against capillary extravasation and physically impeding large molecule diffusion. Additionally, the ECM may interfere with T cell and antibody-based therapies, as cytotoxic T cells must navigate this dense “briar patch” of ECM to reach their targets. Collagen turnover is tightly regulated through an interplay between extracellular zinc-dependent endopeptidase activity and intracellular recycling. MMPs initiate large-scale collagen breakdown by cleaving a small moiety of extracellular collagen after it binds to cell surface receptors. Internalization then occurs through phagocytosis, pinocytosis, endocytosis, or autophagocytosis, with subsequent degradation mainly within phagolysosomes by cathepsin cysteine proteases. While current CN-directed therapies aim to kill or induce senescence in tumor cells, the bulk of cutaneous neurofibromas is composed of ECM. Extracellular collagen, in the absence of enzymatic degradation by MMPs, has a longevity that dwarfs the average human life expectancy due to its dense helical structure and resistance to proteolysis. In CN tumor cells, MMP expression is downregulated in a Ras-independent manner via aryl hydrocarbon receptor-induced extracellular signal-regulated kinase (ERK) phosphorylation. Therefore, blocking Ras signaling with MEK inhibitors is not expected to restore normal collagen turnover. Hypothetically, even if all tumor cells within CNs were eliminated, the skin bumps themselves might remain.

Future strategies should consider a dual-targeted or stepwise approach to address both the tumor cells and the persistent ECM. Potential strategies include the injection of ECM-degrading enzymes or systemic treatment with small molecules that upregulate lysosomal collagen degradation, either preceding or concurrent with Ras pathway inhibition. More attention should be given to the role of the ECM in drug-resistant tumors such as CNs, including studies on diffusion parameters in intact tumors to inform pharmacotherapy development, and investigations into the tumor’s internal pressure dynamics that could reduce drug penetration, especially within hypoxic and acidotic tumor cores. Targeting the ECM may not only improve treatment outcomes but could also serve as a biomarker of treatment efficacy. Annexin A2 mediates collagen secretion and subsequent adhesion to the basement membrane and could potentially be targeted to prevent COLVI deposition within CNs. Lysosome activators could also be evaluated as a means to expedite collagen digestion. C6M, a metabolite of COLVI degradation, can be detected in serum and might serve as a biomarker for the global response to CN-directed therapies targeting the ECM.

CONCLUSION

Developing new and better treatments for CNs is a major priority for the 0.03% of the global population affected by NF1. CNs induce anxiety and depression, negatively impact careers and relationships, cause pain and pruritus, and present a constant visual reminder of a relentlessly progressive disease despite our best current treatments. Fortunately, the NF research community has listened to patient demands, dedicating funding and focus to addressing CNs. Alongside the laborious development of pharmacotherapies and the continued refinement of animal models, access to simple surgical resections could be expanded by

subsidizing and regulating training courses for tumor removal in clinical settings. The ECM represents a previously untargeted component of CNs and may also play a role in larger tumors and malignancies associated with NF1. Focusing on drug delivery at the cellular level will be key to understanding why some pharmacotherapies fail, even when bulk tumor concentrations appear sufficient to elicit a predicted response.

DECLARATIONS

Authors' contributions

The author contributed solely to the article.

Availability of data and materials

Not applicable.

Financial support and sponsorship

None.

Conflicts of interest

Brown RM, MD, PhD, is a paid scientific advisor to Pasithea Therapeutics, which is developing a novel MEK inhibitor PAS-004.

Ethical approval and consent to participate

Not applicable.

Consent for publication

Written informed consent for publication of the clinical images was obtained from the patients.

Copyright

© The Author(s) 2025.

REFERENCES

1. Brown R. Management of central and peripheral nervous system tumors in patients with neurofibromatosis. *Curr Oncol Rep.* 2023;25:1409-17. [DOI](#) [PubMed](#)
2. Lalvani S, Brown RM. Neurofibromatosis type 1: optimizing management with a multidisciplinary approach. *J Multidiscip Healthc.* 2024;17:1803-17. [DOI](#) [PubMed](#) [PMC](#)
3. Ortonne N, Wolkenstein P, Blakeley JO, et al. Cutaneous neurofibromas: current clinical and pathologic issues. *Neurology.* 2018;91:S5-S13. [DOI](#)
4. Page PZ, Page GP, Ecosse E, Korf BR, Leplege A, Wolkenstein P. Impact of neurofibromatosis 1 on quality of life: a cross-sectional study of 176 American cases. *Am J Med Genet A.* 2006;140:1893-8. [DOI](#) [PubMed](#)
5. Wolkenstein P, Zeller J, Revuz J, Ecosse E, Leplège A. Quality-of-life impairment in neurofibromatosis type 1: a cross-sectional study of 128 cases. *Arch Dermatol.* 2001;137:1421-5. [DOI](#) [PubMed](#)
6. Wang J, Fu J, Zhou Y, Gao D, Qing J, Yang G. Global research trends in cutaneous neurofibromas: a bibliometric analysis from 2003 to 2022. *Skin Res Technol.* 2024;30:e13595. [DOI](#) [PubMed](#) [PMC](#)
7. Friedrich RE, Holstein AF, Middendorff R, Davidoff MS. Vascular wall cells contribute to tumorigenesis in cutaneous neurofibromas of patients with neurofibromatosis type 1. A comparative histological, ultrastructural and immunohistochemical study. *Anticancer Res.* 2012;32:2139-58. [PubMed](#)
8. Kallionpää RA, Ahramo K, Martikkala E, et al. Mast cells in human cutaneous neurofibromas: density, subtypes, and association with clinical features in neurofibromatosis 1. *Dermatology.* 2022;238:329-39. [DOI](#)
9. Kallionpää RA, Peltonen S, Le KM, et al. Characterization of immune cell populations of cutaneous neurofibromas in neurofibromatosis 1. *Lab Invest.* 2024;104:100285. [DOI](#)
10. Church C, Fay CX, Kriukov E, et al. snRNA-seq of human cutaneous neurofibromas before and after selumetinib treatment implicates role of altered Schwann cell states, inter-cellular signaling, and extracellular matrix in treatment response. *Acta Neuropathol Commun.* 2024;12:102. [DOI](#) [PubMed](#) [PMC](#)

11. McLean DT, Meudt JJ, Lopez Rivera LD, et al. Single-cell RNA sequencing of neurofibromas reveals a tumor microenvironment favorable for neural regeneration and immune suppression in a neurofibromatosis type 1 porcine model. *Front Oncol.* 2023;13:1253659. DOI PubMed PMC
12. Nagrani NS, Bhawan J. Histopathological variants of cutaneous neurofibroma: a compendious review. *Dermatopathology.* 2022;10:1-19. DOI PubMed PMC
13. Peltonen J, Penttinen R, Larjava H, Aho HJ. Collagens in neurofibromas and neurofibroma cell cultures. *Ann N Y Acad Sci.* 1986;486:260-70. DOI PubMed
14. Brosseau JP, Sathe AA, Wang Y, et al. Human cutaneous neurofibroma matrisome revealed by single-cell RNA sequencing. *Acta Neuropathol Commun.* 2021;9:11. DOI PubMed PMC
15. Jiang C, McKay RM, Le LQ. Tumorigenesis in neurofibromatosis type 1: role of the microenvironment. *Oncogene.* 2021;40:5781-7. DOI PubMed PMC
16. Harigai R, Sakai S, Nobusue H, et al. Tranilast inhibits the expression of genes related to epithelial-mesenchymal transition and angiogenesis in neurofibromin-deficient cells. *Sci Rep.* 2018;8:6069. DOI PubMed PMC
17. Chen P, Cescon M, Megighian A, Bonaldo P. Collagen VI regulates peripheral nerve myelination and function. *FASEB J.* 2014;28:1145-56. DOI PubMed
18. Chen P, Cescon M, Zuccolotto G, et al. Collagen VI regulates peripheral nerve regeneration by modulating macrophage recruitment and polarization. *Acta Neuropathol.* 2015;129:97-113. DOI
19. Cescon M, Gregorio I, Eiber N, et al. Collagen VI is required for the structural and functional integrity of the neuromuscular junction. *Acta Neuropathol.* 2018;136:483-99. DOI
20. Cescon M, Gattazzo F, Chen P, Bonaldo P. Collagen VI at a glance. *J Cell Sci.* 2015;128:3525-31. DOI
21. Cescon M, Rampazzo E, Bresolin S, et al. Collagen VI sustains cell stemness and chemotherapy resistance in glioblastoma. *Cell Mol Life Sci.* 2023;80:233. DOI PubMed PMC
22. Jiang C, Kumar A, Yu Z, et al. Basement membrane proteins in extracellular matrix characterize NF1 neurofibroma development and response to MEK inhibitor. *J Clin Invest.* 2023;133. DOI
23. Williams L, Layton T, Yang N, Feldmann M, Nanchahal J. Collagen VI as a driver and disease biomarker in human fibrosis. *FEBS J.* 2022;289:3603-29. DOI
24. Wang WN, Koguchi-Yoshioka H, Nimura K, et al. Distinct transcriptional profiles in the different phenotypes of neurofibroma from the same subject with neurofibromatosis 1. *J Invest Dermatol.* 2024;144:133-141.e4. DOI
25. Castagnaro S, Gamberotto L, Cescon M, Bonaldo P. Autophagy in the mesh of collagen VI. *Matrix Biol.* 2021;100-101:162-72. DOI PubMed
26. Silver FH. The role of connections between cellular and tissue mechanical elements and the importance of applied energy in mechanotransduction in cancerous tissue. *Biomolecules.* 2025;15:457. DOI PubMed PMC
27. Lu P, Chen Z, Wu M, et al. Type I collagen extracellular matrix facilitates nerve regeneration via the construction of a favourable microenvironment. *Burns Trauma.* 2024;12:tkae049. DOI PubMed PMC
28. Li X, Shepard HM, Cowell JA, et al. Parallel accumulation of tumor hyaluronan, collagen, and other drivers of tumor progression. *Clin Cancer Res.* 2018;24:4798-807. DOI PubMed PMC
29. Rømer AMA, Thorseth ML, Madsen DH. Immune modulatory properties of collagen in cancer. *Front Immunol.* 2021;12:791453. DOI PubMed PMC
30. Henke E, Nandigama R, Ergün S. Extracellular matrix in the tumor microenvironment and its impact on cancer therapy. *Front Mol Biosci.* 2019;6:160. DOI PubMed PMC
31. Huson SM, Harper PS, Compston DA. Von Recklinghausen neurofibromatosis: a clinical and population study in south-east Wales. *Brain.* 1988;111:1355-81. DOI
32. Duong TA, Bastuji-Garin S, Valeyrie-Allanore L, Sbidian E, Ferkal S, Wolkenstein P. Evolving pattern with age of cutaneous signs in neurofibromatosis type 1: a cross-sectional study of 728 patients. *Dermatology.* 2011;222:269-73. DOI PubMed
33. Cannon A, Chen MJ, Li P, et al. Cutaneous neurofibromas in neurofibromatosis type I: a quantitative natural history study. *Orphanet J Rare Dis.* 2018;13:31. DOI PubMed PMC
34. Dugoff L, Sujansky E. Neurofibromatosis type 1 and pregnancy. *Am J Med Genet.* 1996;66:7-10. DOI PubMed
35. Geller M, Mezitis SG, Nunes FP, et al. Progesterone and estrogen receptors in neurofibromas of patients with NF1. *Clin Med Pathol.* 2008;1:93-7. DOI PubMed PMC
36. Fishbein L, Zhang X, Fisher LB, et al. In vitro studies of steroid hormones in neurofibromatosis 1 tumors and Schwann cells. *Mol Carcinog.* 2007;46:512-23. DOI
37. Pennanen P, Peltonen S, Kallionpää RA, Peltonen J. The effect of estradiol, testosterone, and human chorionic gonadotropin on the proliferation of Schwann cells with $NF1^{+/+}$ or $NF1^{-/-}$ genotype derived from human cutaneous neurofibromas. *Mol Cell Biochem.* 2018;444:27-33. DOI PubMed
38. Lammert M, Mautner VF, Kluwe L. Do hormonal contraceptives stimulate growth of neurofibromas? *BMC Cancer.* 2005;5:16. DOI PubMed PMC
39. Cannon A, Jarnagin K, Korf B, et al. Clinical trial design for cutaneous neurofibromas. *Neurology.* 2018;91:S31-7. DOI PubMed PMC
40. Zhu B, Zheng T, Wang W, et al. Genotype-phenotype correlations of neurofibromatosis type 1: a cross-sectional study from a large

- Chinese cohort. *J Neurol.* 2024;271:1893-900. DOI
41. Kehrer-Sawatzki H, Kluwe L, Salamon J, et al. Clinical characterization of children and adolescents with NF1 microdeletions. *Childs Nerv Syst.* 2020;36:2297-310. DOI PubMed PMC
 42. Pinna V, Lanari V, Daniele P, et al. p.Arg1809Cys substitution in neurofibromin is associated with a distinctive NF1 phenotype without neurofibromas. *Eur J Hum Genet.* 2015;23:1068-71. DOI PubMed PMC
 43. Jiang C, McKay RM, Lee SY, et al. Cutaneous neurofibroma heterogeneity: factors that influence tumor burden in neurofibromatosis type 1. *J Invest Dermatol.* 2023;143:1369-77. DOI PubMed PMC
 44. Upadhyaya M, Huson SM, Davies M, et al. An absence of cutaneous neurofibromas associated with a 3-bp inframe deletion in exon 17 of the NF1 gene (c.2970-2972 delAAT): evidence of a clinically significant NF1 genotype-phenotype correlation. *Am J Hum Genet.* 2007;80:140-51. DOI PubMed PMC
 45. Trevisson E, Morbidoni V, Forzan M, et al. The Arg1038Gly missense variant in the NF1 gene causes a mild phenotype without neurofibromas. *Mol Genet Genomic Med.* 2019;7:e616. DOI
 46. Lin MJ, Yao H, Vera K, et al. Cutaneous neurofibromas and quality of life in adults with neurofibromatosis type 1. *JAMA Dermatol.* 2024;160:1091-8. DOI PubMed PMC
 47. Thomas L, Spurlock G, Eudall C, et al. Exploring the somatic NF1 mutational spectrum associated with NF1 cutaneous neurofibromas. *Eur J Hum Genet.* 2012;20:411-9. DOI PubMed PMC
 48. Emmerich D, Zemojtel T, Hecht J, et al. Somatic neurofibromatosis type 1 (NF1) inactivation events in cutaneous neurofibromas of a single NF1 patient. *Eur J Hum Genet.* 2015;23:870-3. DOI PubMed PMC
 49. Asai A, Karnan S, Ota A, et al. High-resolution 400K oligonucleotide array comparative genomic hybridization analysis of neurofibromatosis type 1-associated cutaneous neurofibromas. *Gene.* 2015;558:220-6. DOI
 50. Jacks T, Shih TS, Schmitt EM, Bronson RT, Bernards A, Weinberg RA. Tumour predisposition in mice heterozygous for a targeted mutation in Nf1. *Nat Genet.* 1994;7:353-61. DOI PubMed
 51. Osum SH, Watson AL, Largaespada DA. Spontaneous and engineered large animal models of neurofibromatosis type 1. *Int J Mol Sci.* 2021;22:1954. DOI PubMed PMC
 52. Isakson SH, Rizzardi AE, Coutts AW, et al. Genetically engineered minipigs model the major clinical features of human neurofibromatosis type 1. *Commun Biol.* 2018;1:158. DOI PubMed PMC
 53. White KA, Swier VJ, Cain JT, et al. A porcine model of neurofibromatosis type 1 that mimics the human disease. *JCI Insight.* 2018;3:120402. DOI PubMed PMC
 54. Mo J, Anastasaki C, Chen Z, et al. Humanized neurofibroma model from induced pluripotent stem cells delineates tumor pathogenesis and developmental origins. *J Clin Invest.* 2021;131:139807. DOI PubMed PMC
 55. Mazuelas H, Magallón-Lorenz M, Fernández-Rodríguez J, et al. Modeling iPSC-derived human neurofibroma-like tumors in mice uncovers the heterogeneity of Schwann cells within plexiform neurofibromas. *Cell Rep.* 2022;38:110385. DOI
 56. Liao CP, Pradhan S, Chen Z, Patel AJ, Booker RC, Le LQ. The role of nerve microenvironment for neurofibroma development. *Oncotarget.* 2016;7:61500-8. DOI PubMed PMC
 57. Wu J, Williams JP, Rizvi TA, et al. Plexiform and dermal neurofibromas and pigmentation are caused by Nf1 loss in desert hedgehog-expressing cells. *Cancer Cell.* 2008;13:105-16. DOI PubMed PMC
 58. Le LQ, Shipman T, Burns DK, Parada LF. Cell of origin and microenvironment contribution for NF1-associated dermal neurofibromas. *Cell Stem Cell.* 2009;4:453-63. DOI PubMed PMC
 59. Radomska KJ, Culpier F, Gresset A, et al. Cellular origin, tumor progression, and pathogenic mechanisms of cutaneous neurofibromas revealed by mice with Nf1 knockout in boundary cap cells. *Cancer Discov.* 2019;9:130-47. DOI
 60. Brosseau JP, Pichard DC, Legius EH, et al. The biology of cutaneous neurofibromas: consensus recommendations for setting research priorities. *Neurology.* 2018;91:S14-20. DOI PubMed PMC
 61. Ge LL, Xing MY, Zhang HB, Wang ZC. Neurofibroma development in neurofibromatosis type 1: insights from cellular origin and schwann cell lineage development. *Cancers.* 2022;14:4513. DOI PubMed PMC
 62. Zheng H, Chang L, Patel N, et al. Induction of abnormal proliferation by nonmyelinating schwann cells triggers neurofibroma formation. *Cancer Cell.* 2008;13:117-28. DOI
 63. Parrinello S, Noon LA, Harrisingh MC, et al. NF1 loss disrupts Schwann cell-axonal interactions: a novel role for semaphorin 4F. *Genes Dev.* 2008;22:3335-48. DOI
 64. Chamseddin BH, Hernandez L, Solorzano D, Vega J, Le LQ. Robust surgical approach for cutaneous neurofibroma in neurofibromatosis type 1. *JCI Insight.* 2019;5:128881. DOI PubMed PMC
 65. Wozniak B, Bove T, Zawada T, Calik J. Treatment of cutaneous neurofibromas in patients with neurofibromatosis type 1. *Case Rep Dermatol.* 2023;15:194-201. DOI PubMed PMC
 66. Yoshida Y, Sato N, Furumura M, Nakayama J. Treatment of pigmented lesions of neurofibromatosis 1 with intense pulsed-radio frequency in combination with topical application of vitamin D3 ointment. *J Dermatol.* 2007;34:227-30. DOI PubMed
 67. Quirk B, Olasz E, Kumar S, Basel D, Whelan H. Photodynamic therapy for benign cutaneous neurofibromas using aminolevulinic acid topical application and 633 nm red light illumination. *Photobiomodul Photomed Laser Surg.* 2021;39:411-7. DOI PubMed PMC
 68. Peltonen S, Jannic A, Wolkenstein P. Treatment of cutaneous neurofibromas with carbon dioxide laser: technique and patient experience. *Eur J Med Genet.* 2022;65:104386. DOI PubMed
 69. Chamseddin BH, Le LQ. Management of cutaneous neurofibroma: current therapy and future directions. *Neurooncol Adv.*

- 2020;2:i107-16. [DOI](#) [PubMed](#) [PMC](#)
70. Levine SM, Levine E, Taub PJ, Weinberg H. Electrosurgical excision technique for the treatment of multiple cutaneous lesions in neurofibromatosis type 1. *J Plast Reconstr Aesthet Surg.* 2008;61:958-62. [DOI](#) [PubMed](#)
 71. Verma SK, Riccardi VM, Plotkin SR, et al. Considerations for development of therapies for cutaneous neurofibroma. *Neurology.* 2018;91:S21-30. [DOI](#)
 72. Sarin KY, Bradshaw M, O'Mara C, et al. Effect of NFX-179 MEK inhibitor on cutaneous neurofibromas in persons with neurofibromatosis type 1. *Sci Adv.* 2024;10:eadk4946. [DOI](#) [PubMed](#) [PMC](#)
 73. Ly I, Romo CG, Gottesman S, et al. Target product profile for cutaneous neurofibromas: clinical trials to prevent, arrest, or regress cutaneous neurofibromas. *J Invest Dermatol.* 2023;143:1388-96. [DOI](#) [PubMed](#)
 74. Guiraud M, Bouroubi A, Beauchamp R, et al. Cutaneous neurofibromas: patients' medical burden, current management and therapeutic expectations: results from an online European patient community survey. *Orphanet J Rare Dis.* 2019;14:286. [DOI](#) [PubMed](#) [PMC](#)
 75. Passos J, Soares MP, Salgado D, et al. A single-center case study series assessing the effect of selumetinib use in patients with neurofibromatosis-related plexiform neurofibromas. *Neurooncol Adv.* 2024;6:vdae177. [DOI](#) [PubMed](#) [PMC](#)
 76. Gross AM, Reid OH, Baldwin LA, et al. Treatment of cutaneous neurofibromas in neurofibromatosis type 1 with mek inhibitor selumetinib: a nonrandomized clinical trial. *JAMA Dermatol.* 2025;161:533-7. [DOI](#) [PubMed](#) [PMC](#)
 77. Banerjee J, Allaway RJ, Taroni JN, et al. Integrative analysis identifies candidate tumor microenvironment and intracellular signaling pathways that define tumor heterogeneity in NF1. *Genes.* 2020;11:226. [DOI](#) [PubMed](#) [PMC](#)
 78. Mazuelas H, Magallón-Lorenz M, Uriarte-Arrazola I, et al. Unbalancing cAMP and Ras/MAPK pathways as a therapeutic strategy for cutaneous neurofibromas. *JCI Insight.* 2024;9. [DOI](#) [PubMed](#) [PMC](#)
 79. In M, Richardson KC, Loewa A, Hedtrich S, Kaessmeyer S, Plendl J. Histological and functional comparisons of four anatomical regions of porcine skin with human abdominal skin. *Anat Histol Embryol.* 2019;48:207-17. [DOI](#) [PubMed](#)
 80. Liu Y, Chen JY, Shang HT, et al. Light microscopic, electron microscopic, and immunohistochemical comparison of Bama minipig (*Sus scrofa domestica*) and human skin. *Comp Med.* 2010;60:142-8. [PubMed](#) [PMC](#)